

Evidence for key enzymatic controls on metabolism of Arctic river organic matter

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Abstract

Permafrost thaw in the Arctic driven by climate change is mobilizing ancient terrigenous organic carbon (OC) into fluvial networks. Understanding the controls on metabolism of this OC is imperative for assessing its role with respect to climate feedbacks. In this study, we examined the effect of inorganic nutrient supply and dissolved organic matter (DOM) composition on aquatic extracellular enzyme activities (EEAs) in waters draining the Kolyma River Basin (Siberia), including permafrost-derived OC. Reducing the phenolic content of the DOM pool resulted in dramatic increases in hydrolase EEAs (e.g., phosphatase activity increased >28-fold) supporting the idea that high concentrations of polyphenolic compounds in DOM (e.g., plant structural tissues) inhibit enzyme synthesis or activity, limiting OC degradation. EEAs were significantly more responsive to inorganic nutrient additions only after phenolic inhibition was experimentally removed. In controlled mixtures of modern OC and thawed permafrost endmember OC sources, respiration rates per unit dissolved OC were 1.3–1.6 times higher in waters containing ancient carbon, suggesting that permafrost-derived OC was more available for microbial mineralization. In addition, waters containing ancient permafrost-derived OC supported elevated phosphatase and glucosidase activities. Based on these combined results, we propose that both composition and nutrient availability regulate DOM metabolism in Arctic aquatic ecosystems. Our empirical findings are incorporated into a mechanistic conceptual model highlighting two key enzymatic processes in the mineralization of riverine OM: (i) the role of phenol oxidase activity in reducing inhibitory phenolic compounds and (ii) the role of phosphatase in mobilizing organic P. Permafrost-derived DOM degradation was less constrained by this initial ‘phenolic-OM’ inhibition; thus, informing reports of high biological availability of ancient, permafrost-derived DOM with clear ramifications for its metabolism in fluvial networks and feedbacks to climate.

Keywords: aquatic, Arctic, biogeochemistry, dissolved organic matter, enzymes, global change, organic matter decomposition, permafrost

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Introduction

Arctic climatic warming is occurring faster than the global average, with air temperature increases of 4–7 °C expected for the coming century (ACIA, 2004; Trenberth *et al.*, 2007). Rapid warming results in the thaw and mobilization of extensive amounts of OC currently locked up in frozen permafrost soils, releasing it into the contemporary carbon cycle (Schoor *et al.*, 2008; Schaefer *et al.*, 2011). Permafrost soils are estimated to contain on the order of 1672 Pg C, of which approximately 25% is stored as frozen yedoma soils in the Siberian Arctic (Zimov *et al.*, 2006; Tarnocai *et al.*, 2009).

Permafrost soils thus contain carbon stocks approximately twice the size of the entire current atmospheric carbon pool and almost five times estimated live forest biomass (Tarnocai *et al.*, 2009; Pan *et al.*, 2011). Examining the reactivity and fate of permafrost-derived OC is imperative in understanding northern high-latitude watershed and Arctic ocean carbon dynamics as well as potential climate feedbacks.

River networks efficiently mineralize terrigenous OC, acting as important sites for heterotrophic metabolism and greenhouse gas release to the atmosphere (Cole *et al.*, 2007; Battin *et al.*, 2008; Aufdenkampe *et al.*, 2011). Thawing permafrost and thermokarst processes significantly affect Arctic hydrology and geomorphology (Walvoord *et al.*, 2012) resulting in the direct and indirect release of ancient OC into aquatic ecosystems.

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Studies have highlighted permafrost-derived organic matter to not be safeguarded from microbial degradation by physical protection mechanisms (Fan *et al.*, 2008) and to display geochemical and optical properties indicative of less decomposed OC fractions (Hugelius *et al.*, 2012). Evidence suggests that permafrost OC may be readily utilized by modern microbial communities (Dutta *et al.*, 2006; Uhlir *et al.*, 2007; Waldrop *et al.*, 2010; Vonk *et al.*, 2013a). Furthermore, changing hydrologic conditions resulting from permafrost degradation may result in shifts in nutrient availability as well as DOM composition (Frey & McClelland, 2009). Both inorganic nutrient concentrations and DOM quality have been linked to aquatic OC mineralization in Arctic river systems (Holmes *et al.*, 2008; Mann *et al.*, 2012; Wickland *et al.*, 2012). Yet, mechanisms regulating the degradation of DOM including permafrost-derived OC remain unclear.

Induction of extracellular enzymes (EEAs) allows heterotrophic bacteria to degrade complex organic compounds into smaller molecules for assimilation, and may be an initial rate limiting step for decomposition and nutrient mineralization (Asmar *et al.*, 1994; Sinsabaugh, 1994). Hydrolase enzymes catalyze the hydrolysis of covalently bonded compounds (e.g., peptides/phosphate), whereas oxidative enzymes are capable of cleaving the aromatic nuclei of phenolic compounds, and thus can be produced to degrade polyphenolic compounds often associated with terrestrially derived DOM (Sinsabaugh, 2010). Stepwise degradation of DOM therefore appears to involve a plethora of enzymes, with their activities often used as indicators of freshwater nutrient status or the presence of complex substrates (Sinsabaugh & Moorhead, 1994; Allison & Vitousek, 2005). Environmental constraints that act to minimize the activities of specific enzymes have been posited to be responsible for inhibiting peat decomposition, resulting in what has been considered an enzymatic 'latch' upon global carbon stores (Freeman *et al.*, 2001a).

In this study, we undertook a series of controlled experiments that examined microbial EEA controls on the utilization of aquatic DOM, including permafrost-derived DOM in the Kolyma watershed, Siberia. We examined: (i) the effect of inorganic nutrient supply and DOM composition on OC metabolism and (ii) EEAs in waters containing modern carbon or thawed permafrost endmember OC sources, recognizing potential microbial inhibitors and accelerators. Here, we report on and synthesize several controlled laboratory experiments in conjunction with *in situ* field observations collected over a 2-year period (2011–2012) that provide the empirical basis for our conceptual model of Arctic riverine DOM degradation.

Materials and methods

Site description & sampling

The Kolyma River Basin covers 652 924 km² of North-East Siberia, and represents the largest watershed on earth underlain by continuous permafrost (Fig. 1). Sampling was focused in the lower basin region and was conducted during the summer months of June/July 2011 and 2012. The sample region is highly diverse containing mixed forest and tundra vegetation and mountain to floodplain landscapes (Fig. 1). The area receives relatively low annual precipitation rates (ca. 185 mm yr⁻¹; Welp *et al.*, 2005), but experiences large mean monthly air temperature variations, ranging from ca. 12 °C in summer to ca. -35 °C during winter. Waters from four diverse endmembers were selected and collected for manipulation and incubation experiments (Table 1; Fig. 1). The Kolyma River mainstem was sampled at two different locations (Sites 1 & 5, Fig. 1). Site 1 waters were collected from the Kolyma mainstem approximately 2 km upstream from the city of Cherskiy. The second Kolyma River mainstem site (Site 5; Fig. 1) was collected approximately 120 km upstream from the first (Fig. 1). Both Kolyma River sites represented an integrated signature of waters draining a large proportion of the Kolyma watershed. Site 2 was a series of first-order streams draining a yedoma exposure known as Duvanni Yar (DY), which was sampled in both years 2010 and 2011 and hereafter referred to in the text as DY-A and DY-B, respectively. This exposure has previously been extensively studied and is known to contain soil deposits ranging from between 13 000 and 45 000 years in age (Vasil'chuk & Vasil'chuk, 1997). The

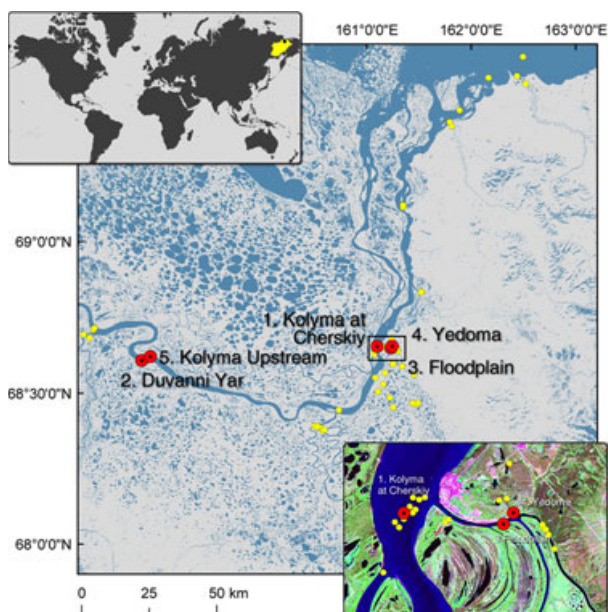


Fig. 1 Study site and sampling locations. Main study sites are represented by red circles with site numbers and names as Table 1. Yellow dots represent sampling locations for waters used to assess ecosystem scale patterns in EEAs (Table S1).

Table 1 Initial chemical and optical measurements of four endmember waters used in PVP and nutrient experiments (1–4) and Kolyma incubation waters mixed with permafrost waters (5). Site locations are displayed in Figure 1. Site DY was sampled on two different occasions in 2010 (DY-A) and 2011 (DY-B)

Exp.	Site no. & description	DOC (mg C l ⁻¹)	DOC:TDN (molar)	NO ₃ ⁻ (μg N l ⁻¹)	NH ₄ ⁺ (μg N l ⁻¹)	PO ₄ ⁻ (μg P l ⁻¹)	SUVA ₂₅₄ (l mg C ⁻¹ m ⁻¹)
Endmember waters (PVP & nutrient)	(1) Kolyma River mainstem at Cherskiy	3.8	10.9	9.9	27.7	2.9	3.2
	(2) Duvanni Yar (DY-B) (permafrost waters)	67.1	6.1	2462	1007	3.1	1.1
	(3) Floodplain stream (FPS)	10.7	15.4	2.0	1.5	2.5	3.1
	(4) Yedomas stream (Y4)	23.3	28.8	1.8	0.4	8.7	3.0
Incubation waters*	(5) Kolyma River upstream of Duvanni Yar	5.3	22.9	–	1.2	2.7	2.2
	(Mixture of 5 & DY-A)	6.6	19.0	–	2.0	3.6	2.2
	(Mixture of 5 & DY-A) Kolyma + 1% DY-A	19.6	16.9	–	51.4	5.0	1.6
	(Mixture of 5 & DY-A) Kolyma + 10% DY-A						

*NH₄⁺ and PO₄⁻ concentrations were measured using OPA and molybdate colorimetric methods, respectively.

small streams are fed by ice-wedge meltwaters draining a cut permafrost bank and thus represent an excellent yedomas permafrost endmember with waters containing OC aged between 20 000 and 30 000 years bp (Vonk *et al.*, 2013a). The remaining two sites (site 3-FPS & site 4-Y4) were small tributary streams draining into the Pantaleikha River; a major tributary of the Kolyma River (Fig. 1). FPS is a small lowland stream draining predominantly floodplain sediments, whereas Y4 represents an upland stream draining largely yedomas soils (Site 3 & 4; Fig. 1). In addition to these four endmember sites, samples were also collected from 42 different waters during 66 different sampling events across the lower basin region (streams, rivers, and lakes) over 2011 and 2012, to assess ecosystem scale patterns in EEAs and respiration rates (Fig. 1; Table S1).

Samples for dissolved organic carbon (DOC) and nutrient analyses were filtered immediately after collection (precombusted 450 °C 0.7 μm, GF/F), and then acidified with HCl (pH < 2). Filtered, unacidified waters were collected for optical analyses and biological oxygen demand (BOD) assays. Enzyme activity assays were also measured upon GF/F filtered waters during the laboratory-based and incubation experiments. EEAs in survey samples were measured on unfiltered surface waters, reducing preparation and assay measurement times. Optical analyses were always conducted within 48 h of collection. Enzyme and BOD assays were started immediately after filtration. All samples were stored cool (4 °C) and in the dark until processing began. All measurements, except nutrient analyses, were conducted on-site at the North-East Science Station, Siberia.

Dissolved organic carbon, total nitrogen, and optical analyses

Dissolved organic carbon (DOC) and total dissolved nitrogen (TDN) were measured on a Shimadzu TOC-V analyzer equipped with an inline chemiluminescence detection unit

(TNM-1) using established protocols (Mann *et al.*, 2012). Chromophoric dissolved organic matter measurements were made using a dual-beam spectrophotometer (Shimadzu UV-1800) and referenced against blank nanopure water spectra. UV-visible scans between 200 and 800 nm were collected at room temperature in 10 mm quartz cells. Specific UV absorbance (SUVA₂₅₄) was calculated as the decadal absorption coefficient at 254 nm divided by the DOC concentration (L mg C⁻¹ m⁻¹). SUVA₂₅₄ has been shown to be a sensitive indicator of DOM aromaticity (Weishaar *et al.*, 2003; Spencer *et al.*, 2012).

Extracellular enzyme assays and biological oxygen demand

The potential enzyme activities of up to six extracellular enzymes, involved in catalyzing the cleavage of a range of organic matter compounds, were investigated (Table 2). Assays were conducted on freshly collected samples using *p*-nitrophenol (pnp) linked substrates (Sigma Aldrich) and measured on a 96-well plate reader (Biotek Powerwave XS2). Phenol oxidase was assayed using L-3, 4-dihydroxyphenylalanine (L-DOPA) and leucine aminopeptidase using L-leucine *p*-nitroanilide (Table 2). Substrate concentrations of 5 mM were used for all enzymes, except LAP and NAG due to their solubility (2 mM). At least four replicate wells were averaged across each sample and triplicate subsamples used for each sample treatment. The variation across wells was typically <5% (max. 10%) and linear activity kinetics were assured by measurements at 4+ timepoints per assay. Each well received 200 μl of sample waters with 50 μl of substrate and enzyme. Substrate and sample blank controls were always run on the same plate and were subtracted from sample values. Plates were incubated at 20 °C and measured periodically over 1–8 h. Potential activities were expressed as the rate of pnp accumulation per hour (nmol l⁻¹ h⁻¹). Protocols for pnp-linked assays are available from the Enzymes in the

Table 2 Enzyme assays measured, their function and the substrates used. α -Gluc and NAGase activities were only measured during laboratory incubation experiments

Enzyme	Function	Substrate
Phenol Oxidase (POx)	Degradation of lignin	L-dihydroxyphenylalanine
Alkaline Phosphatase (APase)	Releases ester-bound phosphate	4-nitrophenyl phosphate disodium
Leucine aminopeptidase (LAP)	Degrades protein into amino acids	L-leucine p-nitroanilide
β -glucosidase (β -gluc)	Release glucose from cellulose decomposition	pnp- β -D- glucopyranoside
α -Glucosidase (α -gluc)	Releases glucose from soluble saccharides	pnp- α -D-glucopyranoside
Glucosaminidase (NAGase)	Degradation of chitin	pnp-N-acetyl glucosaminidase

Environment RCN webpage (enzymes.nrel.colostate.edu). We acknowledge that by using these fixed substrate concentrations we did not measure *in situ* activity rates per se, instead we assessed potential activity rates that are comparable across sites or treatments and influenced only by DOM composition, which was our stated aim. We avoided directly comparing EEAs measured in experimental waters with those measured in survey waters due to the differences in sample preparation.

Sensitive oxygen demand assays were conducted on filtered water samples over 5 day-incubation periods (BOD_{5 d}). Total oxygen loss over the 5-day period was used as an indicator of relative DOM bioavailability. Oxygen loss was converted into estimates of stream and river respiration, assuming a respiratory quotient of 1. Oxygen concentrations were measured using self-stirring BOD probes (YSI, ProOBOD, ± 0.1 mg l⁻¹) in triplicate glass BOD bottles (300 ml). Assays were run at room temperature (ca. 20 °C) and kept dark in-between measurements.

DOM composition and inorganic nutrient experiments

To examine the effect of DOM quality and inorganic nutrient concentrations upon EEA expression in our four endmember waters, the response of a suite of enzymes was followed after amendments to examine dissolved phenolic content and additions of inorganic nitrogen (N), phosphorus (P), or combined N & P. The dissolved phenolic content of waters were reduced by selectively removing them using polyvinylpyrrolidone (PVP; $\geq 95\%$, molecular weight 40 KD; Fisher Scientific, Pittsburgh, PA, USA) at a final concentration of 0.01 g ml⁻¹ (Carpenter *et al.*, 1976; Freeman *et al.*, 2001a). PVP-phenolic complexes were not extracted, but blanks containing PVP alone showed no effect upon enzyme activity rates. Potassium phosphate and potassium nitrate were used as a source of P and N, respectively, and added to yield a final concentration approximately 10 times mean natural concentrations in the Kolyma River Basin as determined using data from 2010 (<http://www.thepolarisproject.org/data/>). Waters were well mixed and split into two treatments, with one left unamended and the other treated with PVP to reduce dissolved phenolic content. Subsamples of both unamended and PVP waters were then amended by additions of inorganic N, P, N, and P, or none to act as a control. All treatment waters were incubated in the dark, at a fixed temperature of 20 °C, for 24 h before EEA measurements. Using this approach, we were able to assess the

independent and combined effects of phenolic content and inorganic nutrient content upon EEA expression.

To investigate the direct response of dissolved phenolics upon freshwater respiration rates, we conducted a separate BOD_{5d} assay on Kolyma River waters (Site 1; Fig. 1) containing different dissolved phenolic concentrations. Respiration rates were measured in three independent treatments; an unamended control and two identical waters amended with different final PVP concentrations (treatment A: 0.010 g ml⁻¹ & treatment B: 0.005 g ml⁻¹).

Incubation experiments

To assess OC loss and EEA expression during degradation of Kolyma waters mixed with ancient OC (DY-A waters) we conducted a controlled incubation experiment. Waters were collected from the Kolyma River mainstem (Site 5; Fig. 1) and four small tributary streams draining the DY exposure (Site 2; Fig. 1; Vonk *et al.*, 2013a). Samples were filtered (precombusted 450 °C 0.7 μ m, GF/F), and a composite sample (DY-A) produced by mixing the filtrate of the four smaller streams. The composite DY-A water contained a Δ^{14} C-DOC signature of -933 representing an OC pool aged $21\,700 \pm 85$ years before present (Vonk *et al.*, 2013a). Studies of major Arctic rivers including the Kolyma River, by contrast, report generally modern Δ^{14} C-DOC ages (Neff *et al.*, 2006; Raymond *et al.*, 2007). Three treatments containing 0%, 1.0%, and 10% dilutions of the DY-A composite with Kolyma River water were then prepared and incubated in triplicate in BOD glass bottles (20 °C in the dark).

Optical parameters, EEA's and nutrient concentrations (NH₄⁺ & PO₄⁻) were measured at the start, and after 5 days of incubation. Oxygen concentrations were measured frequently over the incubation period and continued for a total of 9 days. Ammonium concentrations were measured immediately on-site using the fluorometric method developed by Holmes *et al.* (1999) and modified by Taylor *et al.* (2007). Phosphate concentrations were measured immediately using the molybdate colorimetric method (U.S. Environmental Protection Agency, 1984). No flocculation was observed in optical or visual inspections in any of the treatments over the incubation period.

Statistical analyses

Paired sample *t*-tests were used to assess pre- and post-treatment differences in control waters with and without PVP

treatments, and EEAs pre- and postincubation. Effects of inorganic nutrient additions on EEAs were assessed using one-way analysis of variance (ANOVA) to test for significant differences among treatments vs. the control within each experiment. Significant responses were investigated using pairwise comparisons between group means after Bonferroni correction ($P < 0.05$). Data were normalized using log transformations where nonnormal distributions were observed before analyses. Linear regressions were used to assess correlations between parameters and the coefficient of determination (R^2) calculated. All statistical analyses were conducted using SPSS 20 (IBM, NY, USA).

Results

Endmember and incubation water properties

The initial geochemical properties of each of the four endmember waters studied varied considerably (Table 1). DOC concentrations were lowest in Kolyma mainstem waters (3.8 mg C l^{-1}), intermediate in FPS and Y4 waters ($10.7, 23.3 \text{ mg C l}^{-1}$, respectively), and highest in the DY-B waters draining yedoma deposits (67.1 mg C l^{-1}) (Table 1). TDN and inorganic nitrogen concentrations also ranged dramatically between samples, DY-B waters containing high concentrations of NO_3^- and NH_4^+ causing a low DOC : TDN ratio of 6.1 (Table 1). Highest DOC : TDN concentrations, indicating low inorganic N concentrations were observed in Y4 (28.8) and FPS waters (15.4). PO_4^- concentrations were generally low ($2.5\text{--}8.7 \text{ } \mu\text{g P l}^{-1}$) in all of the sample waters (Table 1). DOM composition also varied between endmember waters (Table 1). DOM aromaticity and molecular weight, inferred from SUVA_{254} values, was similar across sites except for DY-B waters, which contained very low aromatic content waters (low SUVA_{254} ; Table 1).

Initial incubation waters displayed a gradient in DOC content, nutrient concentration, and DOM quality (Table 1). DOC concentrations increased from 5.3 mg C l^{-1} in Kolyma River waters to 6.6 and 19.6 mg C l^{-1} with 1% and 10% DY-A additions, respectively. Additions of DY-A waters resulted in an increase in inorganic N concentrations and associated reduction in DOC : TDN (Table 1). PO_4^- concentrations increased with DY-A additions, increasing from $2.7 \text{ } \mu\text{g P l}^{-1}$ in Kolyma waters, to $5.0 \text{ } \mu\text{g P l}^{-1}$ in the 10% DY-A treatment (Table 1). Successive DY-A water additions also led to a reduction in DOM molecular weight and total aromaticity (lower SUVA_{254} ; Table 1).

Effect of phenolic removal in endmember waters on EEAs & respiration

After experimental manipulations to reduce phenolic content via PVP amendments, mean potential phenol

oxidase (POx) activities declined significantly in each of the endmember waters (52–62% losses) except the Kolyma River (Table 3; Table S2). Kolyma River waters, by contrast, demonstrated a significant rise in POx expression with phenolic reduction resulting in a 210% increase in activities (Table 3). Hydrolase activities displayed markedly different responses to phenolic removal in each of the endmember waters. Largest responses to phenolic removal were observed in APase activities, with increases of 28- and 48-fold in Kolyma River and FPS samples, respectively (Table 3). No response was observed in APase activity rates in DY-B waters after phenolic removal ($P = 0.24$). LAP activities approximately doubled in Kolyma waters after PVP amendments, but resulted in small reductions (50% loss) or no significant response at each of the other three sites. Furthermore, substantial stimulation of β -gluc activities (>25-fold) was observed in Kolyma River waters with phenolic removal (Table 3).

The removal of dissolved phenolics led to dramatic increases in the observed mean respiration rate of Kolyma River mainstem waters (Fig. 2). Unamended Kolyma mainstem waters displayed low respiration rates throughout the entire incubation period ($0.78 \text{ gC-O}_2 \text{ l}^{-1} \text{ g}^{-1} \text{ DOC}$ after 5 days; Fig. 2). PVP-treated Kolyma waters demonstrated consistently higher respiration rates over 7- to 8-fold (treatment B) and 13- to 16-fold (treatment A) greater than measured in untreated waters (Fig. 2). Significantly higher respiration rates persisted over the entire 5-day incubation period in both PVP treatments.

Effect of inorganic nutrient supply upon enzyme activities

Inorganic nutrient additions generally had little effect upon EEAs relative to control waters in unamended endmember waters ('Unamended' Table 4). Activities of APase were significantly lower in DY-B waters after combined N & P additions. The activity of LAP increased significantly, but relatively modestly (ca. 1.5-fold), with combined N and P supplements in the Kolyma River sample (Table 4).

Table 3 Effect of phenolic removal on freshwater enzyme activities (% change). Arrows represent direction of significant % change in EEAs ($P < 0.05$). NS represents nonsignificant response

	POx	APase	LAP	β -gluc
Kolyma	↑ 210	↑ 2800	↑ 120	↑ 2400
Duvanni Yar	↓ 52	NS	NS	↑ 77
FPS	↓ 62	↑ 4800	↓ 58	↓ 100
Y4	↓ 55	↑ 370	NS	NS

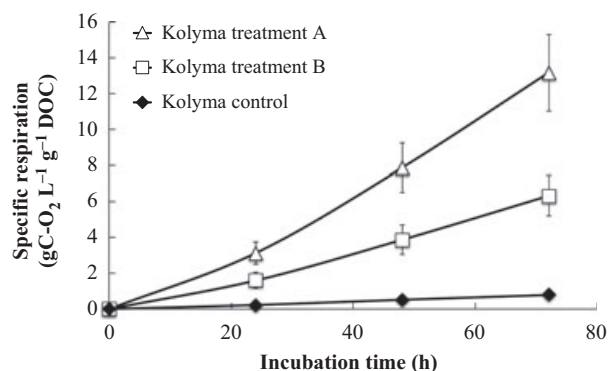


Fig. 2 Cumulative microbial respiration from identical Kolyma waters amended to reduce dissolved phenolic content. Unamended waters (black diamond), PVP treatments A = 0.010 g ml⁻¹ (open triangle) & B = 0.005 g ml⁻¹ (open square). Respiration values were normalized to DOC concentration to account for differences in sample OC concentrations. Values are means \pm 1 SE ($n = 3$). High specific respiration rates per unit DOC indicate a greater proportion of labile OC.

Table 4 Effect of inorganic nitrogen (N), phosphorus (P), and combined additions (N+P) upon potential enzyme activities in unamended (no PVP) and treated waters (PVP added)

Enzyme	Site	Unamended			PVP treated		
		N	P	N & P	N	P	N & P
POx	Kolyma	NS	NS	NS	NS	↓**	↓**
	Duvanni Yar	NS	NS	NS	NS	NS	NS
	FPS	NS	NS	NS	NS	↑*	↑**
	Y4	NS	NS	NS	NS	NS	NS
APase	Kolyma	NS	NS	NS	NS	↑**	↑**
	Duvanni Yar	NS	NS	↓**	NS	NS	NS
	FPS	NS	NS	NS	↑**	↓*	↓**
	Y4	NS	NS	NS	↑**	↑*	↑*
LAP	Kolyma	NS	NS	↑**	NS	↑*	↑**
	Duvanni Yar	NS	NS	NS	↓**	NS	↑**
	FPS	NS	NS	NS	NS	↑**	↑**
	Y4	NS	NS	NS	NS	↑*	↑*
β-gluc	Kolyma	NS	NS	NS	NS	↓**	↓**
	Duvanni Yar	NS	NS	NS	NS	NS	NS
	FPS	NS	NS	NS	NS	↑**	↑**
	Y4	NS	NS	NS	NS	↑*	↑*

Arrow represents direction of significant changes (* $P < 0.05$, ** $P < 0.01$ after Bonferroni correction). NS equals nonsignificant response ($P > 0.05$).

In contrast to unamended samples, waters containing lower concentrations of dissolved phenolic compounds ('PVP treated' Table 4) showed strong responses in EEAs to inorganic nutrient additions. Enzymes associated with N (LAP) and P (APase) cycling were most responsive to nutrient amendments. LAP activity was consistently higher in all endmember waters after

combined N and P additions and in all, except for DY-B, in response to P additions alone. The activity of LAP decreased significantly, with only N fertilization in the DY-B sample (Table 4).

The effect of inorganic nutrients on APase activities was often considerable, but differed between the four endmember waters. APase activities in Kolyma River water increased by ca. 3.5-fold with P and combined N & P fertilization. Similar patterns were observed in Y4 waters, alongside additional increases of APase activities with N amendments alone (ca. 2.5-fold). APase activities also increased in FPS waters with N additions alone, but declined significantly with P and combined additions (Table 4). DY-B waters showed no significant change in APase activities in response to nutrient additions after PVP amendment.

Activities of β-gluc significantly increased in response to both P and combined additions in Y4 and FPS waters post-PVP amendment. In contrast, β-gluc activities were suppressed in the Kolyma River sample with P amendments (P, N+P). Activities of POx also varied in response to both P and N & P amendments, causing a decrease in activities in the Kolyma River sample and significant increases in FPS waters after PVP treatment (Table 4).

Incubation experiments of ancient organic carbon

Respiration rates and EEAs measured within the 0%, 1.0%, and 10% DY-A permafrost treatments differed dramatically over the incubation period (Fig. 3; Table S2). Respiration rates were higher in both treatments amended with DY-A waters (1.0 & 10%), relative to Kolyma River mainstem waters, after only 1 day of incubation (Fig. 3a; $P < 0.01$). Normalized respiration rates (g C-O₂ l⁻¹ g⁻¹ DOC), accounting for differences in initial DOC concentrations, were also consistently higher in DY-A supplemented waters than Kolyma waters after 76 h onwards ($P < 0.01$; Fig. 4). However, no significant differences in respiratory losses were observed between the 1% and 10% DY-A treatments when C-normalized over the entire incubation period (Fig. 4).

Enzyme activity potentials of LAP and APase measured after 5 days of incubation, significantly increased across all treatments relative to preincubation rates ($P < 0.05$; Fig. 3b–d). LAP and APase activities increased approximately 3- to 4-fold over the 5-day period in both the Kolyma River and 1% DY-A treatments. The 10% DY-A sample, by contrast, supported far higher enzyme synthesis with LAP activities increasing over 6-fold and APase activities by 15-fold during the 5-day period. Enzyme activities associated with glucose and chitin degradation (α-gluc, β-gluc, & NAGase)

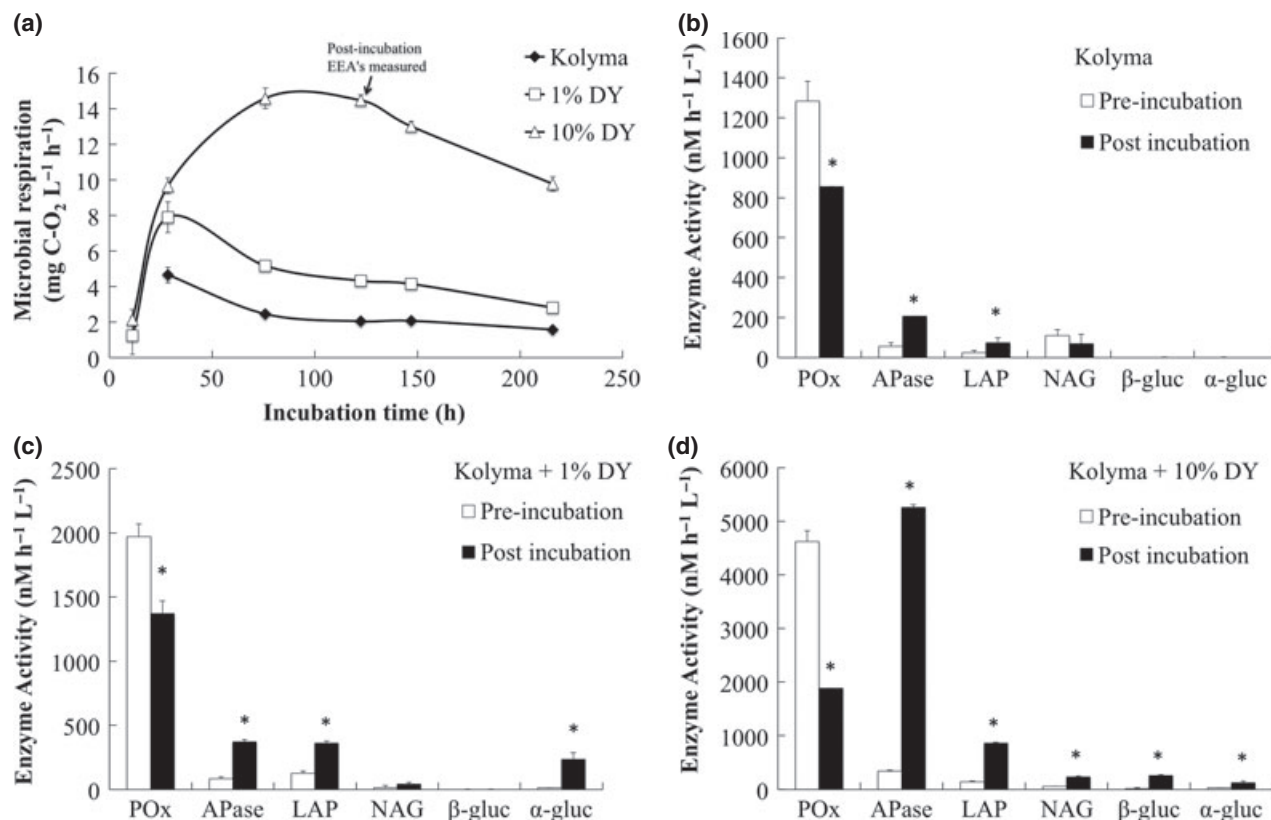


Fig. 3 (a) Microbial respiration in Kolyma, 1% and 10% DY-A treatments in laboratory incubations over 9 days. Values are means \pm 1 SE ($n = 3$). EEAs measured at time 0 and 5 days (arrow). Enzyme activity potentials measured pre- and post 5-day incubation in (b) Kolyma waters (c) Kolyma and 1% DY-A waters, and (d) Kolyma and 10% DY-A waters. Note different y -axis scales. Significant differences between pre- and postincubation activities are indicated by * ($P < 0.05$).

were either undetectable or showed no significant increase in Kolyma River waters. Kolyma River water containing 1.0% DY-A, by contrast, showed significant differences in α -gluc activities with increases of 16-fold over the incubation (Fig. 3c). Kolyma River with 10% DY-A waters also displayed significant differences in α -gluc activities (4-fold increase), as well as significant increases in the activity potentials of β -gluc and NAG-ase (18- & 4-fold increases, respectively, Fig. 3d). In contrast to the hydrolase enzymes, POx activities declined consistently and significantly over the incubation period in all of the treatments ($P < 0.05$). Final NH_4^+ concentrations, after the 5-day incubations were very similar across each treatment (9.7, 8.7 and 10 $\mu\text{g l}^{-1}$, respectively). Final PO_4^- concentrations were extremely low in each of the treatments ranging from 2 $\mu\text{g l}^{-1}$ (1% treatment) to below the detection limit (ca. 0.5 $\mu\text{g l}^{-1}$) in Kolyma and 10% samples.

Ecosystem scale patterns in DOM bioavailability

Enzyme activities varied greatly across diverse waters (streams, rivers, and lakes) within the lower Kolyma

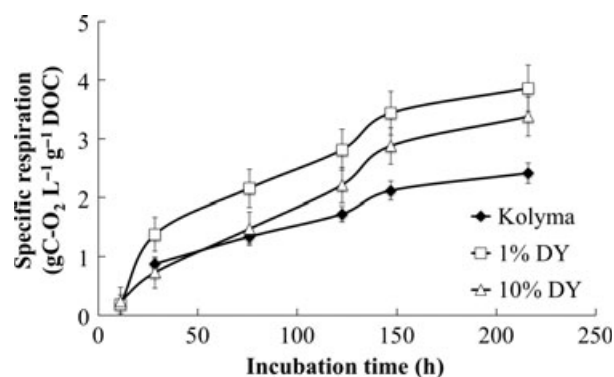


Fig. 4 Cumulative microbial respiration from Kolyma, 1% DY-A and 10% DY-A waters during incubations. Respiration values were normalized to DOC concentration to account for differences in initial sample OC concentrations. Values are means \pm 1 SE ($n = 3$).

Basin watershed (Table S1). Activities of POx, APase, and β -gluc in surface waters were all significantly correlated with respiration rates as determined by biological oxygen demand assays, but only POx explained a large proportion of the variance ($R^2 = 0.41, 0.15, 0.06$,

$P < 0.05$, $n = 65$, respectively) (Fig. 5a). DOC and TDN concentrations, as well as DOM compositional indices (SUVA₂₅₄) also correlated with respiration rates but showed low explanatory power in explaining variance in rates ($R^2 = 0.21, 0.16, 0.10$, $P < 0.05$, $n = 63$, respectively). Inorganic PO_4^- concentrations were highly correlated with respiration rates ($R^2 = 0.53$, $P < 0.01$, $n = 65$; Fig. 5b). Nitrate and ammonium concentrations showed no significant relationship with respiration ($P > 0.05$; Table S1).

Discussion

The bioavailability of permafrost-derived OC will prove critical in determining the fate and response of northern high-latitude ecosystems to climate-induced hydrologic and biogeochemical change. As important mediators of nutrient cycling and decomposition pathways, extracellular enzymes can provide useful information on the constraints limiting the continued breakdown of OC in aquatic ecosystems.

The significant increase in hydrolase enzyme activities we report across diverse waters after reductions in dissolved phenolic content highlights the inhibitory effect polyphenolic compounds can play upon

enzyme expression and thus OC decomposition pathways (Fig. 6, phenolic inhibition). In addition, the marked and dynamic response of hydrolase enzyme activities to inorganic nutrient supply, after reductions in the dissolved phenolic content of waters, demonstrates that the microbial community responds to variability in exogenous nutrient availability only after initial composition constraints have been removed (Fig. 6, nutrient availability). This may have been caused by a shift in the microbial community or an upregulation of enzyme production by the initial microbial population. The immobilization of extracellular enzyme activities has previously been reported in soils as a result of mineral stabilization and the formation of polyphenolic-enzyme complexes (Ladd, 1985). Furthermore, a phenolic inhibition (or 'latch') mechanism has also been proposed to be critical in preserving carbon stocks in low oxygen terrestrial peatland systems (Freeman *et al.*, 2001a; Fenner & Freeman, 2011), suggesting that this preservation mechanism could be globally pervasive. A number of natural pathways may act to reverse the formation of polyphenolic-enzyme complexes in freshwaters. These include the autodigestion of phenolic compounds via alternate enzymes (e.g., POx), dilution processes or ultraviolet photolytic decomposition (Boavida & Wetzel, 1998; Spencer *et al.*, 2009; Mann *et al.*, 2012; Fig. 6).

Extracellular POx enzymes have been shown to be crucial in enabling the degradation of lignin and other polyphenols, acting as one of the few enzymes capable of removing inhibitory phenolic compounds (McLatchey & Reddy, 1998). Previous studies in temperate soils have reported that POx activities were limited by oxygen availability, and suggested that soil OC degradation rates were therefore limited by low environmental oxygen concentrations (Freeman *et al.*, 2001a; Fenner & Freeman, 2011). Ecosystem scale relationships observed here between freshwater POx activities and respiration rates in waters across the Kolyma Basin (Fig. 5a), suggest that the activity of POx may play a crucial role in regulating the biological reactivity of DOM across aquatic ecosystem types. Relationships between DOM phenolic content and bioavailability have also been shown across a permafrost gradient in the Yenisei River (Siberia), with lower phenolic concentrations and higher bioavailability in tributaries with greater active layer depths (Kawahigashi *et al.*, 2004). If initial POx activities are largely a function of soil enzyme concentrations, changes in vegetation type, dominant flow paths or soil enzyme production may dramatically influence aquatic OC bioavailability. POx activities may also be expected to increase in warmer environments; experiments in peatland soils have

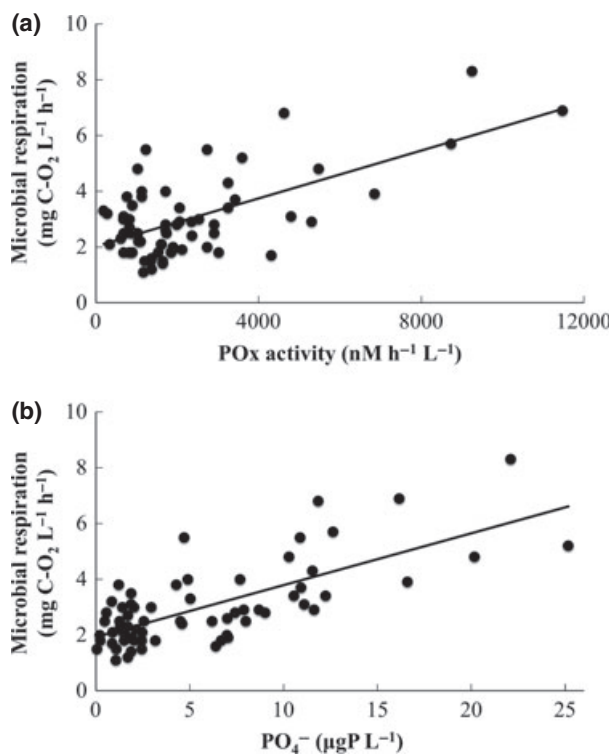


Fig. 5 Ecosystem scale patterns in microbial respiration measured over 2011 and 2012 plotted against (a) phenol oxidase (POx) activity ($R^2 = 0.41$, $P < 0.05$, $n = 65$), and (b) inorganic phosphate (PO_4^-) concentrations ($R^2 = 0.53$, $P < 0.01$, $n = 65$).

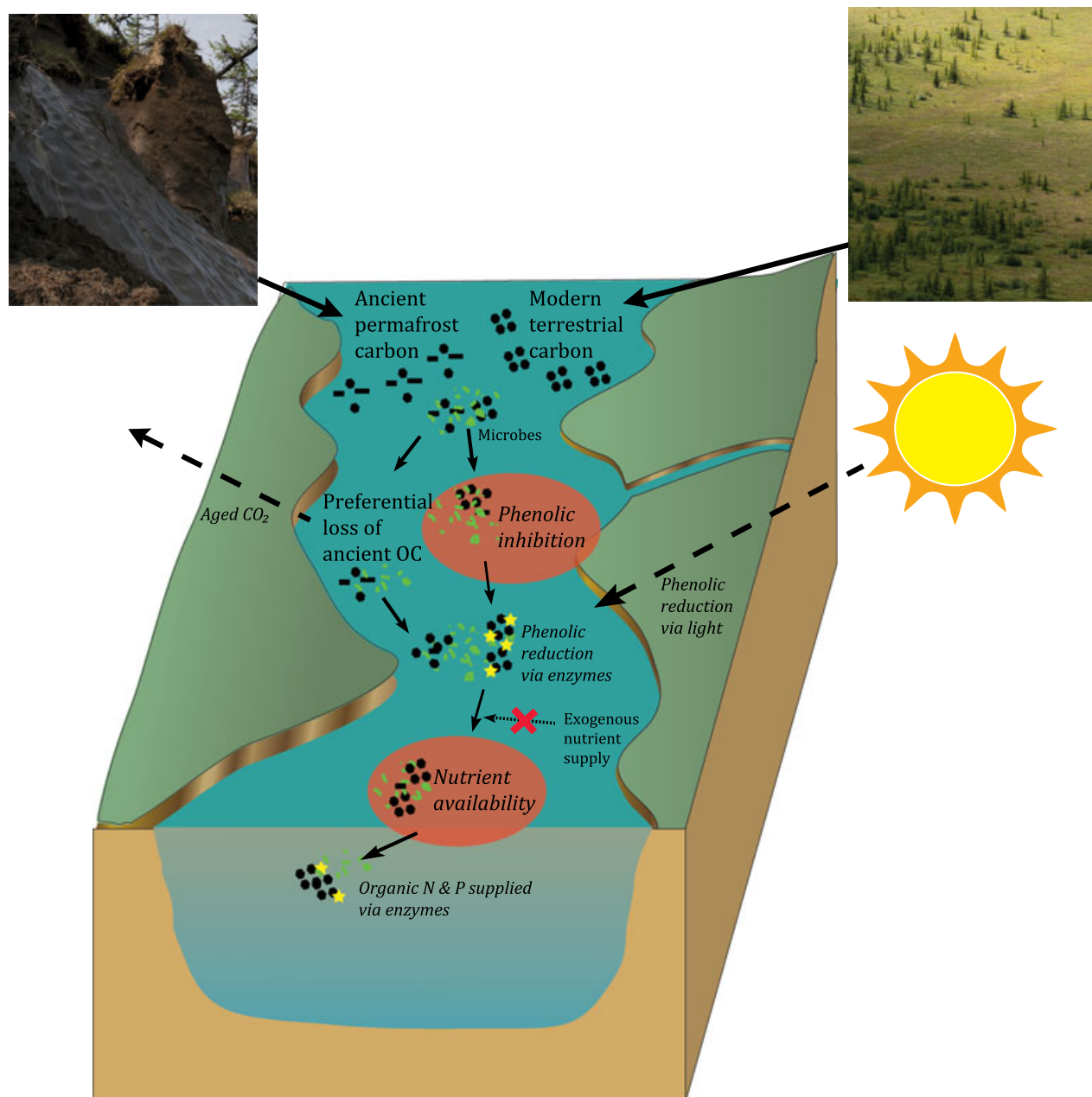


Fig. 6 Conceptual diagram of key controls upon metabolism of Arctic river organic matter (red circles). 'Phenolic inhibition' refers to the compositional constraints caused by inhibitory phenolic compounds. Inhibition from phenolics can be removed via enzymatic processes (stars) or photolysis (dotted line). Secondary constraints upon metabolism relate to limited 'nutrient availability' after exogenous nutrient supply has been exhausted, restricting continued organic carbon degradation. Hydrolase enzyme activity can release organic nutrients promoting continued degradation (stars) but only after initial phenolic constraints have been removed. Inset photographs courtesy of Chris Linder (<http://www.chrislinder.com/>).

demonstrated increased POx activities in warmer soils accompanying increased DOC release with higher phenolic compound content (Freeman *et al.*, 2001b). Recent findings from Arctic inland surface waters report >40% increases in the susceptibility of dissolved ancient OC to microbial degradation when exposed to UV light, as compared to dark controls (Cory *et al.*, 2013). These

results provide evidence that photochemical alteration can result in increased bioavailability, likely as a result of phenolic removal (Spencer *et al.*, 2009; Stubbins *et al.*, 2010) and easing of biochemical constraints upon degradation (e.g., extracellular enzyme production).

After compositional constraints are removed, nutrient availability became the dominant factor

determining continued aquatic OC losses. Across our study sites, APase activities were closely associated with the availability of phosphate (after phenolic removal; Table 4; Fig. 5b). APase activities generally increased when inorganic N availability increased and were suppressed when inorganic PO_4^- was in ample supply. These results agree with experiments in soils (Clarholm, 1993), and suggest APase activities are a sensitive indicator of the P status of freshwaters across the Kolyma watershed. Contrastingly, LAP and β -gluc activities responded little to N and P addition alone, but increased when both N and P were abundant. These enzymes were therefore likely produced in response to higher carbon processing rates, or a reduction in carbon availability. The rapid synthesis of APase enzymes in our endmember waters and observed during incubations of ancient OC, in association with the strong relationships between PO_4^- concentration and respiration rates (Table 3; Fig. 5b) indicate that phosphate availability plays a dominant role in influencing long-term OC bioavailability rates. The release of organic P via APase activities will therefore become the limiting factor once exogenous inorganic P has been depleted (Fig. 6, nutrient availability), yet APase activities will continue to be depressed and unresponsive until phenolic constraints have been removed (Tables 3 and 4; Fig. 6, phenolic inhibition).

A major concern of warming air temperatures across Arctic regions is that it may lead to enhanced permafrost thaw, resulting in the transfer of previously sequestered ancient OC to the atmosphere via respiration in soils, or via increased mobilization to inland waters and metabolism in the aquatic environment. Yedoma-derived OC (DY) displayed higher respiration rates per unit DOC than OC containing largely modern carbon, indicating that ancient OC was inherently more bioavailable over short time-scales. These results are in good agreement with a recent study that traced dissolved OC loss, rather than O_2 removal, at similar sites (Vonk *et al.*, 2013a). These findings suggest that ancient permafrost carbon inputs to the stream network may be extensively processed within Arctic stream and river networks, resulting in a flux of aged CO_2 to the atmosphere, where it may serve to accelerate future climate warming. Despite the largely modern ^{14}C -DOC signatures observed in many northern river systems during non-baseflow conditions (e.g. Evans *et al.*, 2007; Raymond *et al.*, 2007), selective in stream degradation of DOC may therefore result in the loss of an ancient DOC fraction that is masked by larger continued modern DOC inputs.

The limited response of DY waters to phenolic removal and the high initial enzyme activities we

report suggest that yedoma influenced waters contain lower initial dissolved polyphenolic concentrations, or contain such elevated initial enzyme concentrations that they were unable to be fully complexed by phenolics within the DOM pool (Wetzel, 1992). Alternatively, DY waters may contain high concentrations of intrinsic bacteria, limiting the need for an initial growth phase before EEA production (Rivkina *et al.*, 1998). Successive DY additions to Kolyma mainstem waters, mimicking changing contributions that may be expected with increasing permafrost thaw, resulted in a shift in the overall OC composition toward less aromatic, low-molecular-weight DOM, as inferred from lower SUVA_{254} (Table 1). These reductions in DOM aromaticity and molecular weight led to the observed increase in respiration rates (Figs 3 and 4).

Recent studies in the Yukon River Basin have highlighted that an increase in the contribution of organics from deeper soil horizons and groundwaters in permafrost regions is expected to result in the export of OC with lower aromaticity (SUVA_{254}), lower lignin carbon-normalized yields, and with higher contributions of hydrophilic relative to hydrophobic acids (Striegl *et al.*, 2007; Spencer *et al.*, 2008; O'Donnell *et al.*, 2010, 2012). Ice-wedge complexes, constituting up to 50% of the total volume of yedoma permafrost, have also been shown to contain low-molecular-weight, low aromaticity DOM which is readily available to microbial degradation (Vonk *et al.*, 2013b). Furthermore, a linear relationship between $\Delta^{14}\text{C}$ -DOC and SUVA_{254} values has also been reported in the Kolyma Basin, suggesting older OC pools typically display lower DOM aromaticity (Neff *et al.*, 2006). In the context of the results presented here it seems apparent that yedoma-derived OC is highly biologically labile in stream networks in comparison to modern terrestrial OC inputs, primarily as a result of its composition (Fig. 6, phenolic inhibition). As continued warming in the Arctic will lead to more extensive permafrost thaw it seems likely to result in a shift toward more readily available OC for metabolism, leading to increased mineralization of ancient organic carbon in downstream receiving aquatic ecosystems. Under this scenario, it is apparent that the aquatic microbial processing of permafrost-derived OC may play an important role in permafrost carbon and climate feedbacks with respect to global climate change.

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References

- ACIA (2004) *Impacts of a Warming Planet: Arctic Climate Impact Assessment*. Cambridge University Press, Cambridge, UK.
- Allison SD, Vitousek PM (2005) Responses of extracellular enzymes to simple and complex nutrient inputs. *Soil Biology and Biochemistry*, **37**, 937–944.
- Asmar F, Eiland F, Nielson NE (1994) Effect of extracellular-enzyme activities on solubilization rate of soil organic nitrogen. *Biology and Fertility of Soils*, **17**, 32–38.
- Aufdenkampe AK, Mayorga E, Raymond PA *et al.* (2011) Riverine coupling of biogeochemical cycles between land, oceans, and atmosphere. *Frontiers in Ecology and the Environment*, **9**, 53–60.
- Battin TJ, Kaplan LA, Findlay S *et al.* (2008) Biophysical controls on organic carbon fluxes in fluvial networks. *Nature Geoscience*, **1**, 95–100. doi: 10.1038/ngeo101.
- Boavida MJ, Wetzel RG (1998) Inhibition of phosphatase activity by dissolved humic substances and hydrolytic reactivation by natural ultraviolet light. *Freshwater Biology*, **40**, 285–293. doi:10.1046/j.1365-2427.1998.00349.x.
- Carpenter A, Siggia S, Carter S (1976) Separation and/or concentration of phenolic materials from dilute aqueous solutions. *Analytical chemistry*, **48**, 225–228.
- Clarholm M (1993) Microbial biomass P, labile P, and acid phosphatase activity in the humus layer of a spruce forest, after repeated additions of fertilizers. *Biology and Fertility of Soils*, **16**, 287–292.
- Cole JJ, Prairie YT, Caraco NF *et al.* (2007) Plumbing the global carbon cycle: integrating inland waters into the terrestrial carbon budget. *Ecosystems*, **10**, 172–185.
- Cory RM, Crump BC, Dobkowski JA, Kling GW (2013) Surface exposure to sunlight stimulates CO₂ release from permafrost soil carbon in the Arctic. *Proceedings of the National Academy of Sciences*, doi: 10.1073/pnas.1214104110, published ahead of print February 11, 2013.
- Dutta K, Schuur EAG, Neff JC, Zimov SA (2006) Potential carbon release from permafrost soils of Northeastern Siberia. *Global Change Biology*, **12**, 2336–2351.
- Evans CD, Freeman C, Cork LG *et al.* (2007) Evidence against recent climate-induced destabilisation of soil carbon from 14C analysis of riverine dissolved organic matter. *Geophysical Research Letters*, **34**, L07407. doi:10.1029/2007GL029431.
- Fan Z, Neff JC, Harden JW, Wickland KP (2008) Boreal soil carbon dynamics under a changing climate: a model inversion approach. *Journal of Geophysical Research*, **113**, G04016.
- Fenner N, Freeman C (2011) Drought-induced carbon loss in peatlands. *Nature Geoscience*, **4**, 895–900.
- Freeman C, Ostle N, Kang H (2001a) An enzymic “latch” on a global carbon store. *Nature*, **409**, 149.
- Freeman C, Evans CD, Monteith DT, Reynolds B, Fenner N (2001b) Export of organic carbon from peat soils. *Nature*, **412**, 785. doi:10.1038/35090628.
- Frey KE, McClelland JW (2009) Impacts of permafrost degradation on arctic river biogeochemistry. *Hydrological Processes*, **23**, 169–182.
- Holmes RM, Aminot A, Kerouel R, Hooker BA, Peterson BJ (1999) A simple and precise method for measuring ammonium in marine and freshwater ecosystems. *Canadian Journal of Fisheries and Aquatic Sciences*, **56**, 1801–1808.
- Holmes RM, McClelland JW, Raymond PA, Frazer BB, Peterson BJ, Stieglitz M (2008) Lability of DOC transported by Alaskan rivers to the Arctic Ocean. *Geophysical Research Letters*, **35**, L03402.
- Hugelius G, Routh J, Kuhry P, Crill P (2012) Mapping the degree of decomposition and thaw remobilization potential of soil organic matter in discontinuous permafrost terrain. *Journal of Geophysical Research: Biogeosciences*, **117**, G2030.
- Kawahigashi M, Kaiser K, Kalbitz K, Rodionov A, Guggenberger G (2004) Dissolved organic matter in small streams along a gradient from discontinuous to continuous permafrost. *Global Change Biology*, **10**, 1576–1586.
- Ladd JN (1985) Soil Enzymes. *Soil organic matter and biological activity* (eds. Vaughan D, Malcolm RE), pp. 175–221. Springer, Dordrecht, Netherlands.
- Mann PJ, Davydova A, Zimov N *et al.* (2012) Controls on the composition and lability of dissolved organic matter in Siberia’s Kolyma River basin. *Journal of Geophysical Research*, **117**, G01028.
- McLatchey GP, Reddy KR (1998) Regulation of organic matter decomposition and nutrient release in a wetland soil. *Journal of Environmental Quality*, **27**, 1268–1274.
- Neff JC, Finlay JC, Zimov SA, Davydov SP, Carrasco JJ, Schuur EAG, Davydova AI (2006) Seasonal changes in the age and structure of dissolved organic carbon in Siberian rivers and streams. *Geophysical Research Letters*, **33**, L23401.
- O’Donnell JA, Aiken GR, Kane ES, Jones JB (2010) Source water controls on the character and origin of dissolved organic matter in streams of the Yukon River basin, Alaska. *Journal of Geophysical Research*, **115**, G03025.
- O’Donnell JA, Aiken GR, Walvoord MA, Butler KD (2012) Dissolved organic matter composition of winter flow in the Yukon River basin: implications of permafrost thaw and increased groundwater discharge. *Global Biogeochemical Cycles*, **26**, GB0E06. doi:10.1029/2012GB004341.
- Pan Y, Birdsey RA, Fang J *et al.* (2011) A large and persistent carbon sink in the world’s forests. *Science*, **333**, 988–993.
- Raymond PA, McClelland JW, Holmes RM *et al.* (2007) Flux and age of dissolved organic carbon exported to the Arctic Ocean: a carbon isotopic study of the five largest arctic rivers. *Global Biogeochemical Cycles*, **21**, GB4011.
- Rivkina E, Gilichinsky D, Wagerer S, Tiedje J, McGrath J (1998) Biogeochemical activity of anaerobic microorganisms from buried permafrost sediments. *Geomicrobiology*, **15**, 187–193.
- Schaefer K, Zhang T, Bruhwiler L, Barrett AP (2011) Amount and timing of permafrost carbon release in response to climate warming. *Tellus B*, **63B**, 165–180.
- Schuur EAG, Bockheim J, Canadell JG *et al.* (2008) Vulnerability of permafrost carbon to climate change: implications for the global carbon cycle. *BioScience*, **58**, 701.
- Sinsabaugh RS (1994) Enzymic analysis of microbial pattern and process. *Biology and Fertility of Soils*, **17**, 69–74.
- Sinsabaugh RL (2010) Phenol oxidase, peroxidase and organic matter dynamics of soil. *Soil Biology and Biochemistry*, **42**, 391–404.
- Sinsabaugh RL, Moorhead DL (1994) Resource allocation to extracellular enzyme production: a model for nitrogen and phosphorus control of litter decomposition. *Soil Biology and Biochemistry*, **26**, 1305–1311.
- Spencer RGM, Aiken GR, Wickland KP, Striegl RG, Hernes PJ (2008) Seasonal and spatial variability in dissolved organic matter quantity and composition from the Yukon River Basin, Alaska. *Global Biogeochemical Cycles*, **22**, GB4002. doi:10.1029/2008GB003231.
- Spencer RGM, Stubbins A, Hernes PJ *et al.* (2009) Photochemical degradation of dissolved organic matter and dissolved lignin phenols from the Congo River. *Journal of Geophysical Research – Biogeosciences*, **114**, G03010. doi:10.1029/2009JG000968.
- Spencer RGM, Butler KD, Aiken GR (2012) Dissolved organic carbon and chromophoric dissolved organic matter properties of rivers in the U.S.A. *Journal of Geophysical Research – Biogeosciences*, **117**, G03001. doi:10.1029/2011JG001928.
- Striegl RG, Dornblaser MM, Aiken GR, Wickland KP, Raymond PA (2007) Carbon export and cycling by the Yukon, Tanana, and Porcupine rivers, Alaska, 2001–2005. *Water Resources Research*, **43**, W02411.
- Stubbins A, Spencer RGM, Chen H *et al.* (2010) Illuminated darkness: the molecular signatures of Congo River dissolved organic matter and its photochemical alteration as revealed by ultrahigh precision mass spectrometry. *Limnology and Oceanography*, **55**, 1467–1477.
- Tarnocai C, Canadell JG, Schuur EAG, Kuhry P, Mazhitova G, Zimov S (2009) Soil organic carbon pools in the northern circumpolar permafrost region. *Global Biogeochemical Cycles*, **23**, GB2023.
- Taylor BW, Keep CF, Hall RO, Koch BJ, Tronstad LM, Flecker AS, Ulseth AJ (2007) Improving the fluorometric ammonium method: matrix effects, background fluorescence, and standard additions. *Journal of the North American Benthological Society*, **26**, 167–177.
- Trenberth KE, Jones PD, Ambenje P *et al.* (2007) Observations: surface and atmospheric climate change. In: *Climate Change 2007. The Physical Science Basis. Contribution of WG I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change*. (eds Solomon S, Qin D, Manning M, Chen Z, Marquis MC, Averyt KB, Tignor M, Miller HL), pp. 235–336 + plus annex online. Cambridge University Press, Cambridge, UK.
- Uhlirva E, Santruckova H, Davydov SP (2007) Quality and potential biodegradability of soil organic matter preserved in permafrost of Siberian tussock tundra. *Soil Biology and Biochemistry*, **39**, 1978–1989.
- U.S. Environmental Protection Agency (1984) *Methods for Chemical Analysis of Water and Wastes*. Report EPA-600/4-79-020, Environmental Monitoring and Support Laboratory, Office for Research and Development, Cincinnati, OH, Washington, D. C.
- Vasil’chuk YK, Vasil’chuk AC (1997) Radiocarbon Dating and Oxygen Isotope Variations in Late Pleistocene Syngenetic Ice-Wedges, Northern Siberia. *Permafrost and Periglacial Processes*, **8**, 335–345.
- Vonk JE, Mann PJ, Davydov S *et al.* (2013a) High biolability of ancient permafrost carbon upon thaw. *Geophysical Research Letters*, **40**, 2689–2693. doi: 10.1002/grl.50348.

- Vonk JE, Mann PJ, Dowdy KL *et al.* (2013b) Dissolved organic carbon loss from Yedoma permafrost amplified by ice wedge thaw. *Environmental Research Letters*, **8**. doi: 10.1088/1748-9326/8/3/035023.
- Waldrop MP, Wickland KP, White R III, Berhe AA, Harden JW, Romanovsky VE (2010) Molecular investigations into a globally important carbon pool: permafrost-protected carbon in Alaskan soils. *Global Change Biology*, **16**, 2543–2554. doi: 10.1111/j.1365-2486.2009.02141.x.
- Walvoord MA, Voss CI, Wellman TP (2012) Influence of permafrost distribution on groundwater flow in the context of climate-driven permafrost thaw: example from Yukon Flats Basin, Alaska, USA. *Water Resources Research*, **48**. doi: 10.1029/2011WR011595.
- Weishaar JL, Aiken GR, Bergamaschi BA, Fram MS, Fujii R, Mopper K (2003) Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition and reactivity of dissolved organic carbon. *Environmental Science & Technology*, **37**, 4702–4708.
- Welp LR, Randerson JT, Finlay JC, Davydov SP, Zimova GM, Davydova AI, Zimov SA (2005) A high-resolution time series of oxygen isotopes from the Kolyma River: implications for the seasonal dynamics of discharge and basin-scale water use. *Geophysical Research Letters*, **32**, 1–5. doi: 10.1029/2005GL022857.
- Wetzel RG (1992) Gradient-dominated ecosystems: sources and regulatory functions of dissolved organic matter in freshwater ecosystems. *Hydrobiologia*, **229**, 181–198.
- Wickland KP, Aiken GR, Butler K, Dornblaser MM, Spencer RGM, Striegl RG (2012) Biodegradability of dissolved organic carbon in the Yukon River and its tributaries: seasonality and importance of inorganic nitrogen. *Global Biogeochemical Cycles*, **26**, GB0E03.
- Zimov SA, Schuur EAG, Chapin FS III (2006) Permafrost and the global carbon budget. *Science*, **312**, 1612–1613.

Supporting Information

Additional Supporting Information may be found in the online version of this article:

Table S1. EEAs and sample characteristics for surface waters measured throughout the Lower Kolyma Basin (Fig. 1).

Table S2. EEAs for DOM composition and inorganic nutrient addition experiments (Tables 3 and 4) and ancient organic carbon incubations (Fig. 3).