



# Molecular Cancer Research

## Inflammation, a Key Event in Cancer Development

Haitian Lu, Weiming Ouyang and Chuanshu Huang

*Mol Cancer Res* 2006;4:221-233. Published OnlineFirst March 23, 2006.

**Updated version** Access the most recent version of this article at:  
doi:[10.1158/1541-7786.MCR-05-0261](https://doi.org/10.1158/1541-7786.MCR-05-0261)

**Cited Articles** This article cites by 220 articles, 84 of which you can access for free at:  
<http://mcr.aacrjournals.org/content/4/4/221.full.html#ref-list-1>

**Citing articles** This article has been cited by 47 HighWire-hosted articles. Access the articles at:  
<http://mcr.aacrjournals.org/content/4/4/221.full.html#related-urls>

**E-mail alerts** [Sign up to receive free email-alerts](#) related to this article or journal.

**Reprints and Subscriptions** To order reprints of this article or to subscribe to the journal, contact the AACR Publications Department at [pubs@aacr.org](mailto:pubs@aacr.org).

**Permissions** To request permission to re-use all or part of this article, contact the AACR Publications Department at [permissions@aacr.org](mailto:permissions@aacr.org).

## Subject Review

# Inflammation, a Key Event in Cancer Development

Haitian Lu, Weiming Ouyang, and Chuanshu Huang

*Nelson Institute of Environmental Medicine, New York University School of Medicine, Tuxedo, New York*

### Abstract

**Several recent studies have identified nuclear factor- $\kappa$ B as a key modulator in driving inflammation to cancers. Besides this transcription factor, essential in regulating inflammation and cancer development, an inflammatory microenvironment inhabiting various inflammatory cells and a network of signaling molecules are also indispensable for the malignant progression of transformed cells, which is attributed to the mutagenic predisposition of persistent infection-fighting agents at sites of chronic inflammation. As a subverted host response to inflammation-induced tumors, the inflammatory cells and regulators may facilitate angiogenesis and promote the growth, invasion, and metastasis of tumor cells. Thus far, research regarding inflammation-associated cancer development has focused on cytokines and chemokines as well as their downstream targets in linking inflammation and cancer. Moreover, other proteins with extensive roles in inflammation and cancer, such as signal transducers and activators of transcription, Nrf2, and nuclear factor of activated T cells, are also proposed to be promising targets for future studies. The elucidation of their specific effects and interactions will accelerate the development of novel therapeutic interventions against cancer development triggered by inflammation. (Mol Cancer Res 2006;4(4):221–33)**

### Introduction

The link between inflammation and cancers, rather than a recent concern, was noticed ~150 years ago. As early as 1863, Virchow indicated that cancers tended to occur at sites of chronic inflammation (1). Lately, it turned out that acute inflammation contributed to the regression of cancer (2). However, accumulated epidemiologic studies support that chronic inflammatory diseases are frequently associated with increased risk of cancers (1–3). The investigation aiming at the

relationship between inflammation and cancers first led to the determination whether the reactive oxygen and nitrogen species generated by inflammatory cells, such as leukocytes recruited to the inflammatory foci to kill infectious agents, may cause mutagenic assaults and result in tumor initiation (4). Now, it has been realized that the development of cancers from inflammation might be a process driven by inflammatory cells as well as a variety of mediators, including cytokines, chemokines, and enzymes, which altogether establish an inflammatory microenvironment (3). Although this host response may suppress tumors, it may also facilitate cancer development via multiple signaling pathways (5). This review focuses on critical molecular players during the development from inflammation to carcinogenesis. We also discuss several potential targets, such as inducible nitric oxide synthase (iNOS), cyclooxygenase-2 (COX-2), and hypoxia-inducible factor-1 $\alpha$  (HIF-1 $\alpha$ ), which have functions in both inflammatory responses and cancer development.

### Inflammation: From Acute to Chronic

Inflammation is a physiologic process in response to tissue damage resulting from microbial pathogen infection, chemical irritation, and/or wounding (2). At the very early stage of inflammation, neutrophils are the first cells to migrate to the inflammatory sites under the regulation of molecules produced by rapidly responding macrophages and mast cells prestationed in tissues (3, 6). As the inflammation progresses, various types of leukocytes, lymphocytes, and other inflammatory cells are activated and attracted to the inflamed site by a signaling network involving a great number of growth factors, cytokines, and chemokines (3, 6). All cells recruited to the inflammatory site contribute to tissue breakdown and are beneficial by strengthening and maintaining the defense against infection (3).

There are also mechanisms to prevent inflammation response from lasting too long (7). A shift from antibacterial tissue damage to tissue repair occurs, involving both proinflammatory and anti-inflammatory molecules (7). Prostaglandin E<sub>2</sub> (8), transforming growth factor- $\beta$  (9), and reactive oxygen and nitrogen intermediates (6) are among those molecules with a dual role in both promoting and suppressing inflammation. The resolution of inflammation also requires a rapid programmed clearance of inflammatory cells: neighboring macrophages, dendritic cells, and backup phagocytes do this job by inducing apoptosis and conducting phagocytosis (10–12). The phagocytosis of apoptotic cells also promotes an anti-inflammatory response, such as enhancing the production of anti-inflammatory mediator transforming growth factor- $\beta$  (13–15). However, if inflammation resolution is dysregulated, cellular response changes to the pattern of chronic inflammation. In

Received 12/14/05; revised 1/27/06; accepted 1/30/06.

**Grant support:** NIH/National Cancer Institute grants CA112557, CA103180, and CA094964 and NIH/National Institute of Environmental Health Sciences grant ES012451 (C. Huang).

The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked advertisement in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

**Requests for reprints:** Chuanshu Huang, Nelson Institute of Environmental Medicine, New York University School of Medicine, 57 Old Forge Road, Tuxedo, NY 10987. Phone: 845-731-3519; Fax: 845-351-2118. E-mail: chuanshu@env.med.nyu.edu

Copyright © 2006 American Association for Cancer Research.  
doi:10.1158/1541-7786.MCR-05-0261

chronic inflammation, the inflammatory foci are dominated by lymphocytes, plasma cells, and macrophages with varying morphology (2). Macrophages and other inflammatory cells generate a great amount of growth factors, cytokines, and reactive oxygen and nitrogen species that may cause DNA damage (3). If the macrophages are activated persistently, they may lead to continuous tissue damage (16). A microenvironment constituted by all the above elements inhabits the sustained cell proliferation induced by continued tissue damage, thus predisposes chronic inflammation to neoplasia (1).

### Cancer Development: An Overview

Cancer defines malignant neoplasms characterized by metastatic growth. It may occur in almost every organ and tissue relating to a variety of etiologic factors, such as genomic instability and environmental stress (2). A two-stage carcinogenesis model is first conceptualized in a mouse model of skin cancer (17). In this model, carcinogenesis is initiated by carcinogen-triggered irreversible genetic alteration and then promoted by dysregulated gene expression of initiated cells that resulted from epigenetic mechanisms and host-selective pressure (3). Once the proliferation advantage is obtained, cancer cells enter the progression stage in which their population expands rapidly (4). This model was subjected to criticism because it oversimplifies and failed to apply to all types of cancer (18). However, cancer development is still accepted as a multistep process, during which genetic alterations confer specific types of growth advantage; therefore, it drives the progressive transformation from normal cells to malignant cancer cells (19). Malignant growth is characterized by several key changes: self-sufficiency of growth signals, insensitivity to antigrowth signals, escaping from apoptosis, unregulated proliferation potential, enhanced angiogenesis, and metastasis (19). Each of these shifts is complicated and accomplished by combined efforts of various signaling processes. In later discussion, we will find out that inflammation may contribute to the formation of these cancer phenotypes.

### Inflammation and Cancer: Evidence from Epidemiology and Clinical Studies

The association between inflammation and cancer was illustrated by epidemiologic and clinical studies (1, 3, 20). For instance, the risk of colorectal cancer was 10-fold greater if linked with inflammatory bowel disease, such as ulcerative colitis and Crohn's disease (21, 22). Moreover, the control of colitis by certain anti-inflammatory agent reduced colon cancer incidence (23, 24). In the context of the respiratory system, it was also suggested that cancer risk is positively associated with the severity and duration of inflammatory diseases (25, 26). For example, dysplastic progression in nasopharyngeal carcinoma was attributed to EBV (3, 16).

The cause of inflammation may be microbial infection or a noninfective physical and/or chemical irritant (2). In the gastrointestinal tract, gastric *Helicobacter pylori* infection is the leading cause of adenocarcinoma and mucosa-associated lymphoid tissue lymphoma (3, 16). In the bile tract, cholangiocarcinoma was followed by chronic inflammatory

infiltrate induced by *Clonorchis sinensis* infection (16). Within the hepatic system, chronic hepatitis caused by hepatitis B and C viruses predisposes into hepatocellular carcinoma, the third leading cause of cancer mortality globally (27). Moreover, human papillomavirus infection is the leading cause of penile and anogenital cancers. Schistosomiasis and human herpesvirus type 8 may increase the risk of bladder cancer and Kaposi's sarcoma, respectively (1, 28).

Chronic inflammation not caused by infection may also contribute to carcinogenesis. The risk of esophageal cancer, pancreatic cancer, and gallbladder cancer may be increased by inflammatory diseases, such as esophagitis, Barrett's metaplasia, and chronic pancreatitis (16, 29). Possible associations were also found in Marjolin's ulcer and skin carcinoma (16), asbestos and mesothelioma (16), silica, cigarette smoke, and bronchial cancer (16), chronic asthma and lung cancer (30-32), sarcoidosis and lung, skin, and liver cancer (33), ulcerative lichen planus and verrucous carcinoma (34, 35), foreskin inflammation/phimosis and penile cancer (36), and pelvic inflammatory disease or ovarian epithelial inflammation and ovarian cancer (16, 37). Chronic prostatitis, resulting from either persistent bacteria infection or noninfective stimuli, was associated with prostate cancer (38). Therefore, there is increasing evidence that supports the association between chronic inflammation and cancer development.

### Mechanisms for the Association between Inflammation and Cancer

How does chronic inflammation develop to tumors? What are the important driving forces in this process? Studies from an animal model suggested a sequence of histopathologic events from chronic gastritis to gastric carcinogenesis (39). Chronic inflammation is characterized by sustained tissue damage, damage-induced cellular proliferation, and tissue repair (39). Cell proliferation in this context is usually correlated with "metaplasia," a reversible change in cell type (39). "Dysplasia," a disorder of cellular proliferation leading to atypical cells production, follows and is regarded as the previous event of carcinoma because it was usually found adjacent to the site of neoplasm (21). A recent study added further details into the above model and proposed a new paradigm that might be applied to all epithelial cancers (40). Within a mouse model of gastric cancer derived from *H. pylori* infection-induced chronic inflammation, bone marrow-derived cells were recruited to the site of chronic inflammation when the tissue-based stem cell compartment was exhausted by sustained chronic injury. These engrafted bone marrow-derived cells are more pliable and have a greater potential to develop into cancer through the putative "metaplasia, dysplasia, and cancer" process (40). However, a detailed mechanism applicable to all types of cancers is still unclear.

### Mutagenic Potential of Inflammation

The chronic inflammation microenvironment is predominated by macrophages (3, 6). Those macrophages, together with other leukocytes, generate high levels of reactive oxygen and nitrogen species to fight infection (41). However, in a setting of continuous tissue damage and cellular proliferation, the

persistence of these infection-fighting agents is deleterious (4). They may produce mutagenic agents, such as peroxynitrite, which react with DNA and cause mutations in proliferating epithelial and stroma cells (41, 42). Macrophages and T lymphocytes may release tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and macrophage migration inhibitory factor to exacerbate DNA damage (43). Migration inhibitory factor impairs p53-dependent protective responses, thus causing the accumulation of oncogenic mutations (44). Migration inhibitory factor also contributes to tumorigenesis by interfering Rb-E2F pathway (45). Within an ileocolitis-associated mouse cancer model, the high susceptibility to inflammation and cancer in hydroperoxide-reducing enzyme-deficient mice suggested that intracellular hydroperoxides might also contribute to tumor initiation (46).

### *Role of Inflammatory Cells in Tumor Development*

Other than a single mutation, more genetic and epigenetic events are required to drive from initiated cells to malignant tumors (19). Some of these events are also found to be related to chronic inflammation. For instance, angiogenesis, a critical process in tumor progression (47), associates with chronic inflammation, such as psoriasis, rheumatoid arthritis, and fibrosis (19). In addition, the tumor inflammatory microenvironment can facilitate the breakage of the basement membrane, a process required for the invasion and migration of tumor cells (3). A wide population of leukocytes and other types of immune cells infiltrate to the developing tumor site and establish the tumor inflammatory microenvironment (5). Macrophages, neutrophils, eosinophils, dendritic cells, mast cells, and lymphocytes are also found to be key components in the epithelial-originated tumors (5, 16, 48).

The infiltration of immune cells to tumors may repress tumor growth (49-57). However, the increasing concern is that inflammatory cells act as tumor promoters in inflammation-associated cancers (3, 58, 59). Accumulated mutations in epithelial cells lead to dysregulation of their growth and migration. These dysregulated epithelial cells may also signal to recruit leukocytes (47). In addition, tumor cells may also produce cytokines and chemokines to attract immune cells to facilitate cancer development (3, 5, 47). In clinical studies, increased tumor-associated macrophages (TAM) density was found to associate with poor prognosis (60-65), whereas the role of other immune cells (e.g., dendritic cells, mast cells, and neutrophils) in tumor development is still under investigation because of inconsistent results (66-72).

TAMs contribute to tumor development through several mechanisms. TAMs release interleukin (IL)-10 and prostaglandin  $E_2$ , which suppress antitumor response (73). TAMs may also facilitate tumor growth by releasing angiogenic factors, such as vascular endothelial growth factor (VEGF), endothelin-2, and urokinase-type plasminogen activator (43, 74-80). A positive feedback loop may exist because both VEGF and endothelin-2 have chemotactic effects on TAMs (74, 81, 82). TAMs may produce IL-1, which up-regulates VEGF transcription (83). TAMs may also facilitate tumor cell invasion and metastasis by releasing matrix metalloproteinases (MMP-2 and MMP-9), which degrade the extracellular matrix and the

basement membrane (43, 84). In addition, TAMs may induce TNF- $\alpha$  and iNOS, the role of which links inflammation to cancer and will be discussed later in detail. Moreover, TAMs release epidermal growth factor and other epidermal growth factor receptor family ligands to promote tumor cell proliferation and migration (61, 81, 85-87). A paracrine loop of epidermal growth factor might exist for macrophages to synergistically interact with tumors, enhancing metastasis (88). Several macrophage-associated inflammatory factors identified recently might contribute to the increase of tumor susceptibility (89). Activated mast cells generate angiogenic growth factors, such as VEGF/vascular permeability factor and basic fibroblast growth factor, specific angiogenic regulators histamine and heparin, MMP-9, and mast cell-specific proteases MCP-4 and MCP-6 (47, 69, 90-92). Therefore, activated mast cells are suggested to be involved in tumor angiogenesis, invasion, and metastasis. Inflammatory mast cells also enhance tumor progression by releasing cytokines and chemokines (47). Tumor-associated neutrophils enhance tumor angiogenesis, invasion, and metastasis in a similar manner to TAMs and mast cells (47, 93, 94). Neutrophils may also play a role in genetic instability of tumors (95).

T lymphocytes are recruited to tumors by a series of chemokines. At the premalignant lesion stage in a skin cancer model, the knockout of T cells resulted in the decreased leukocyte infiltration and reduced level of MMP-9 (96). Consistently, the increase of CD4<sup>+</sup> T cells was positively correlated with poor prognosis in both renal cell cancer and colorectal cancer (97, 98). Future studies should address the role of lymphocytes in cancer development. Recently, it was shown that transforming growth factor- $\beta$  signaling in T lymphocytes suppressed colon tumor growth by inhibiting IL-6 (99).

### *Key Molecular Players in Linking Inflammation to Cancer*

To address the details of transition from inflammation to cancers and the further development of inflammation-associated cancers, it is necessary to investigate specific roles of key regulatory molecules involved in this process (Table 1).

**Cytokines.** Cytokines, including IL, TNF- $\alpha$ , growth factors, and differentiation factors (colony-stimulating factors), are secreted or membrane-bound molecules that play a regulatory role in the growth, differentiation, and activation of immune cells (100). Cytokine signaling could contribute to the progression of tumors in two aspects: the stimulation of cell growth and differentiation and the inhibition of apoptosis of altered cells at the inflammatory site (43, 44).

The immune response to tumors is constituted by cytokines produced by tumor cells as well as host stromal cells. Tumor-derived cytokines, such as Fas ligand, VEGF, and transforming growth factor- $\beta$ , may facilitate the suppression of immune response to tumors (58). Moreover, inflammatory cytokines have also been reported to facilitate the spectrum of tumor development (2, 58, 100). For example, there is accumulating evidence linking IL-6 to colon cancers (99, 101, 102). IL-6 was first found to play a regulatory role on the proliferation of intestinal epithelial cells (103). In colon cancer patients, IL-6

**Table 1. Key Molecular Players Linking Cancer to Inflammation**

Potential linkers	Functions in linking inflammation to cancer	Refs.
Cytokines		
IL-6	Promote tumor growth	(99, 102)
TNF- $\alpha$	Induce DNA damage and inhibit DNA repair	(108)
	Promote tumor growth	(2, 104, 109)
	Induce angiogenic factors	(110)
Chemokines	Promote tumor cell growth	(3)
	Facilitate invasion and metastasis by directing tumor cell migration and promoting basement membrane degradation	(19, 115-119)
NF- $\kappa$ B	Mediate inflammation progress, promoting chronic inflammation	(134, 135, 142)
	Promote the production of mutagenic reactive oxygen species	(128)
	Protect transformed cells from apoptosis	(157, 158)
	Promote tumor invasion and metastasis	(5, 160)
	Feedback loop between proinflammatory cytokines	(135, 136)
iNOS	Downstream of NF- $\kappa$ B and proinflammatory cytokines	(130, 163)
	Induce DNA damage and disrupt DNA damage response	(108, 166)
	Regulate angiogenesis and metastasis	(167)
COX-2	Produce inflammation mediator prostaglandins	(6, 171)
	Promote cell proliferation, antiapoptotic activity, angiogenesis, and metastasis	(175-181)
HIF-1 $\alpha$	Promote chronic inflammation	(194, 195)
	Induced by proinflammatory cytokines through NF- $\kappa$ B	(83, 196, 197)
	Enhance the glycolytic activity of cancer cells	(198)
	Contribute to angiogenesis, tumor invasion, and metastasis by transactivating VEGF	(198)
STAT3	Activated by proinflammatory cytokines	(199, 200)
	Promote proliferation, apoptosis resistance, and immune tolerance	(201, 202)
Nrf2	Anti-inflammatory activity	(204-206)
	Protect against DNA damage	(208, 210, 211)
NFAT	Regulate proinflammatory cytokine expression	(212-215)
	Required in cell transformation	Yan and Huang, unpublished data

serum levels were found to be strongly elevated and positively correlated to tumor load (101). An *in vitro* study also showed that IL-6 enhanced colony formation of human colon carcinoma cells in a dose-dependent manner, indicating its potential role in promoting cancer growth (102). The role of IL-6 in colon cancer progression has been confirmed *in vivo* by a recent study (99). Within this study, it was found that the IL-6 signaling was mediated by soluble IL-6 receptor derived by tumor cells rather than membrane-bound IL-6 receptor. The inhibition of IL-6 production and IL-6 signaling suppresses the growth of colon cancer (99).

A large number of studies suggest that TNF and chemokines are candidate linking molecules between inflammation and cancer (104-106). TNF, produced mainly by activated macrophages but also by tumor cells, binds to membrane-bound homotrimeric receptors TNFRI and TNFRII (107). In inflammation, TNF plays a critical role in both tissue destruction and damage recovery, maintaining the reversibility of microenvironments, stimulating cellular change, and tissue remodeling (104). TNF may initiate an inflammatory cascade consisting of other inflammatory cytokines, chemokines, growth factors, and endothelial adhesion factors, recruiting a variety of activated cells at the site of tissue damage (104). TNF has both anticancer and procancer actions (104). High-dose administration of TNF might destruct tumor vasculature and have necrotic effects in tumors (104). In contrast, TNF has been found to be required in chemical carcinogen-elicited skin carcinogenesis (105) and also is a major inducer for nuclear factor- $\kappa$ B (NF- $\kappa$ B) activation, which shows antiapoptotic activity (2). The contradictory roles of TNF in regulating cell death might be attributed to the diverse modifications of TNF receptor complexes triggering opposite pathways (106). In addition, TNF can induce DNA damage (108), inhibit DNA repair (108), and act as a growth

factor for tumor cells (109). The ability of TNF to remodel tissues in inflammatory responses may also enable it to regulate the interactions of tumor cells with stroma as well as extracellular matrix. Furthermore, TNF- $\alpha$  may promote angiogenesis and tumor growth by inducing a range of angiogenic factors, thymidine phosphorylase, and MMPs (1, 104, 110, 111). Another role of TNF in linking inflammation to cancer might be its regulation of a network of chemokines (104).

Chemokines include the largest family of cytokines. They can be categorized into CC, CXC, XC, and CX<sub>3</sub>C based on their relative positions of conserved cysteine residues. In the processes of inflammation, chemokines, usually induced by cytokines, are major soluble regulators that control the directional migration of leukocytes to the inflammatory site (96). It is well established that chemokines are involved in the promotion of cancer (96, 112, 113). Previous studies have shown that the expression of CXC receptor 2 might promote preneoplastic cell transformation under certain circumstances (114). It has also been reported that some specific chemokines can promote tumor cell growth (3). Moreover, chemokines also facilitate tumor invasion and metastasis in various cancer types (96, 112, 113) and the balance between chemokines with proangiogenic and angiostatic activities is critical in regulating angiogenesis (19). Mechanistically, chemokines may contribute to tumor invasion and metastasis by mediating the directional migration of tumor cells to specific distal organs via circulation in a similar manner to its control of leukocyte migration (19). They may also facilitate the metastasis of tumor cells by inducing the expression of MMPs and collagenases, which degrade the basement membrane (115-118). A recent study on CXCL-8 showed its role in the interaction between tumor cells and the host environment (119). Ras-transfected human cancer cells are able to produce CXCL-8, a chemokine encoded only in



the human genome. After xenografted into nude mice, those cells secrete CXCL-8 in a paracrine manner, which can recruit host (mice)–produced inflammatory cells to initiate tumor inflammation and angiogenesis, thus facilitating cancer progression (119).

**Nuclear Factor- $\kappa$ B.** NF- $\kappa$ B is a collective term referring to dimeric transcription factors of the Rel family (120). In the cytoplasm, NF- $\kappa$ B exists in the form of an inactive NF- $\kappa$ B-I $\kappa$ B complex in which I $\kappa$ B inhibits NF- $\kappa$ B (121). In response to extracellular stimuli, such as cytokines, I $\kappa$ B is subjected to phosphorylation, ubiquitination, and proteolytic degradation via a canonical I $\kappa$ B kinase (IKK) complex-dependent pathway or a noncanonical NF- $\kappa$ B-inducing kinase pathway (120, 121). I $\kappa$ B degradation may be also through the phosphorylation by casein kinase 2 (122, 123). After I $\kappa$ B degradation, NF- $\kappa$ B is released and translocates to the nucleus (124), where it binds to the promoter regions of its target genes (125). The optimal NF- $\kappa$ B activation also involves the phosphorylation of NF- $\kappa$ B itself. For instance, recent studies showed that the phosphorylation of NF- $\kappa$ B/p65 on Ser<sup>536</sup> is required for the poly-ubiquitination and degradation of I $\kappa$ B $\alpha$ , the predominant I $\kappa$ B protein (126). Suppression of this certain phosphorylation reduced the activation and nucleus translocation of NF- $\kappa$ B and functionally led to the resistance of JB6 cells to TNF- $\alpha$ -induced transformation (127).

Targets of transcription factor NF- $\kappa$ B include immune-mediating genes and inflammatory genes, antiapoptotic genes, cell proliferation regulation genes, and genes encoding negative regulators of NF- $\kappa$ B (128). Within the immune system, NF- $\kappa$ B is involved in the maturation of dendritic cells (129) and the development of lymphocytes (130-133). NF- $\kappa$ B acts as a critical mediator of inflammation progress, regulating the expression of a wide range of inflammatory molecules, such as cytokines and adhesion factors (134, 135). As an important regulator, NF- $\kappa$ B is subjected to tight control by several proteins, such as the zinc finger protein A20 (136). Aberrant and constitutive NF- $\kappa$ B activation plays a role in a variety of inflammatory diseases, including rheumatoid arthritis, atherosclerosis, asthma, inflammatory bowel disease, and *H. pylori*–associated gastritis (135, 136). The lack of IKK $\beta$  in keratinocytes enhances TNF- $\alpha$ -dependent inflammation (137). However, the inhibition of NF- $\kappa$ B in both enterocytes and a murine model of Crohn's disease results in a reduction of inflammatory response (138, 139). Recently, in a murine asthma model, the adenoviral delivery of a NF- $\kappa$ B inhibitory protein to lung epithelium results in the decrease of allergic airway inflammation (140). NF- $\kappa$ B is also required for neutrophil chemotaxis in intraepidermal inflammation (141). It has also been reported that NF- $\kappa$ B regulates inflammatory cell apoptosis and phagocytosis (7, 142). These results indicate that NF- $\kappa$ B may play a critical role in the establishment of chronic inflammation. In regard to carcinogenesis, NF- $\kappa$ B suppresses apoptosis by various mechanisms (128, 138, 143, 144). For instance, it has been shown that the proapoptotic activity of antineoplastic cyclopentenone prostaglandin involves NF- $\kappa$ B inhibition as well as the decrease of various NF- $\kappa$ B-dependent antiapoptotic proteins in malignant B cells (145). In mucosa-associated lymphoid tissue lymphoma, the inhibition of p53-mediated apoptosis was found through the

activation of NF- $\kappa$ B pathway (146). In addition, NF- $\kappa$ B contributes to tumor development by stimulating cell proliferation, because it activates the expression of growth factor genes, proto-oncogene c-Myc, and cell cycle regulator cyclin D1 (128, 147, 148). NF- $\kappa$ B may also play an essential role in late-stage cancer development. It was found that NF- $\kappa$ B is required for the metastasis of injected cultured mammary epithelial cells transformed by Ras oncogene (149).

NF- $\kappa$ B is activated by inflammatory stimuli and its constitutive activation is found in cancer (128); as a result, it has long been suspected to be a critical promoter facilitating the development from inflammation into cancer (128). In squamous epithelium, bacterial lipopolysaccharide-induced human keratinocyte proliferation was found to be dependent on NF- $\kappa$ B activation and subsequent cyclin D1 up-regulation (150). NF- $\kappa$ B may also contribute to genomic instability in two aspects. It promotes the production of reactive oxygen species, which have a potential to cause mutations (128). On the other hand, the antiapoptotic activity of NF- $\kappa$ B prevents mutated precancerous cells from being eliminated (128). NF- $\kappa$ B might be involved in linking inflammation to cancer because of the association between NF- $\kappa$ B and the induction of proinflammatory cytokines, such as IL-6 and TNF- $\alpha$ , and chemokines, such as IL-8, adhesion molecules, MMPs, COX-2, and iNOS (130). Adhesion molecules, such as E-selectin, vascular cell adhesion molecule-1, and intercellular adhesion molecule-1, are responsible for the requirement of NF- $\kappa$ B in leukocyte adhesion and migration, which are important in both inflammation and the inflammatory microenvironment of cancer (151). The increase of adhesion molecules has also been found in certain types of cancer in clinical studies (152-155). This may be because those adhesion molecules are used by tumor cells to facilitate migration and positioning in the process of metastasis (3). Recently, it has been found that decoy receptor 3, a molecule associated with tumorigenesis as well as monocyte differentiation and function, up-regulates intercellular adhesion molecule-1, vascular cell adhesion molecule-1, and IL-8, all of which promote monocyte adhesion (5). NF- $\kappa$ B activation is required for this up-regulation (5). MMPs, as mentioned before, facilitate tumor invasion by their proteolytic activity (128). Noticeably, it is suggested that certain feedback loops exist between proinflammatory cytokines (TNF- $\alpha$ , for instance) and NF- $\kappa$ B activation. In this putative feedback loop, NF- $\kappa$ B is activated by and induces the expression of proinflammatory cytokines (135, 136). This feedback loop may also be responsible for the constitutive activation and geographic spread of NF- $\kappa$ B from cell to cell found in inflammatory diseases and essential for the proposed role of NF- $\kappa$ B in linking inflammation with cancers. For instance, an autocrine system involving IL-1 $\alpha$  has also been found to result in the constitutive expression of NF- $\kappa$ B within a metastatic human cancer model (156).

Recent studies using different animal models provided direct illustrations for the role of NF- $\kappa$ B at the tumor promotion stage in the development of cancers from chronic inflammation (157, 158). One study using two strains of genetically altered mice from a mouse model of colitis-associated cancer showed that the blockade of the IKK $\beta$  gene in the two different cells (myeloid lineage and epithelial cells) may both result in tumor

regression but via different mechanisms (158). NF- $\kappa$ B activation is through IKK complex in inflammatory settings; therefore, NF- $\kappa$ B could not be activated after the knockout of IKK $\beta$  (159). NF- $\kappa$ B pathway inactivation, in the myeloid lineage that macrophages derive from, results in the reduction of both the tumor incidence and the sizes of occurred tumors, which is found to be due to the decrease of several proinflammatory factors facilitating tumor growth (158). The knockout of IKK $\beta$  in epithelial cells leads to the decrease of tumor incidence more often, but no effect on tumor sizes, which indicates that the lack of NF- $\kappa$ B pathway in epithelial cells increases epithelial apoptosis during very early tumor promotion (158). Consistently, NF- $\kappa$ B was also found to protect epithelial cells against inflammation-induced apoptosis in a cell-autonomous role (138). The similar conclusion was also derived from another inflammation-associated cancer model (157). In a Mdr2-knockout mouse strain that spontaneously develops hepatitis and hepatocellular carcinoma, NF- $\kappa$ B in hepatocytes is activated by the inflammatory process via the increase of TNF- $\alpha$  in endothelial and inflammatory cells within the microenvironment (157). After introducing a second knockout of NF- $\kappa$ B, it was found that the blockade of NF- $\kappa$ B does not affect the accumulation of premalignant hepatocytes but leads to the apoptosis of transformed hepatocytes, thus reducing the tumor formation to a great extent (157). Another recent study using a colon cancer cell line indicated a potential role of NF- $\kappa$ B in inflammation-induced metastasis (160). In a murine cancer metastasis model, the introduction of colon adenocarcinoma cell line caused lung metastases. The growth of such lung metastases was found to be stimulated by the injection of bacterial lipopolysaccharide. The knockout of NF- $\kappa$ B in the colon adenocarcinoma cells resulted in the regression of lipopolysaccharide-induced tumor metastases (160).

**Inducible Nitric Oxide Synthase.** iNOS, an enzyme-catalyzing NO production, was found to be overexpressed in chronic inflammatory diseases and various types of cancer (161). Recently, it was found in an animal model study that a selective NOS inhibitor prevents the progression of rat esophageal tumorigenesis, which is induced by the carcinogen *N*-nitrosomethylbenzylamine (162). NO is an important regulatory molecule in both inflammation response (163) and cancer development (164). Because iNOS is subjected to the induction by proinflammatory cytokines, such as TNF- $\alpha$  and IL-1 $\beta$  (163), and the transactivation by NF- $\kappa$ B (130), it may be a downstream effector of cytokines and NF- $\kappa$ B in linking inflammation to cancer. In an experimental model of colitis, TNF- $\alpha$  resulted in inflammation partially via the expression of iNOS triggered by TNF- $\alpha$  (165). Moreover, the blockade of lipopolysaccharide-induced NF- $\kappa$ B activation led to the inhibition of iNOS expression and NO generation and the inhibition of inflammation (161). Under the circumstances of chronic inflammation, the continuous generation of NO may lead to DNA damage, disruption of DNA repair, and cancer-prone post-translational modification (108, 166, 167). Increased NO production might result in p53 activation but also carcinogenic p53 mutations (163, 166, 168). Once the inflammation-associated tumors are formed, iNOS expression may be persistently stimulated by

cytokines and NF- $\kappa$ B that are prevalent within the tumor inflammatory microenvironment (130). NO may also regulate angiogenesis, leukocyte adhesion and infiltration, and metastasis (167). A recent population-based study found that specific polymorphisms in the promoter region of iNOS gene led to higher promoter activities correlated with a higher incidence of gastric cancer in nonsmoking Japanese women (169). It was also suggested that, due to the higher promoter activities of iNOS, excess NO may be produced and cause chronic inflammation, which contributes to the *H. pylori*-induced gastric cancer (169). Noticeably, studies using a wide range of *in vitro* and *in vivo* models show that iNOS/NO signaling can also induce COX-2, which itself is a promising link between inflammation and cancer (167).

**Cyclooxygenase-2.** COX-2 expression may be induced by a wide range of stimuli, including lipopolysaccharide, proinflammatory cytokines, such as IL-1 and TNF, and growth factors, such as epidermal growth factor (128, 170). The products of COX-2 enzyme are prostaglandins, which are key mediators of inflammation (6, 171). Various nonsteroidal anti-inflammatory drugs affect COX-2 activity by covalent modification or competition for a substrate binding site, and the long-term use of nonsteroidal anti-inflammatory drugs was shown by population-based studies to reduce the risk of several cancers (170, 172-174). COX-2 is also overexpressed in various types of cancer and involved in cellular proliferation, antiapoptotic activity, angiogenesis, and an increase of metastasis (175-181). COX-2 pathway is induced by cigarette smoke in human lung fibroblasts, indicating a possible mechanism for the cigarette smoking-triggered cancer-prone inflammatory lung diseases (182). The functions of COX-2 in linking inflammation to cancer are now becoming the target of intense investigation. A recent study using an esophageal model of rats indirectly supports that the COX-2 induction might contribute to the progression of cancers from inflammation (183). In this study, COX-2 inhibitor celecoxib inhibits the COX-2 pathway and delays the developing progress from esophageal inflammation, metaplasia, to adenocarcinoma. However, inconsistency still exists in regard to the exact role of COX-2 in the development from inflammation to cancer. Another recent study in the context of Barrett's esophageal epithelium showed that COX-2 expression is independent of the degree of inflammation but related to the premalignant cells that already existed in the tissue (184). These results indicate that COX-2 expression may not be the driving force for the development from inflammation to cancer but rather play a role in enhancing cancer development in the scenario of chronic inflammation. This notion is supported by a recent immunohistochemical study of human prostate cancers in which the local chronic inflammation has been found to be able to up-regulate COX-2 expression in adjacent tumor cells and induced angiogenesis (185). However, inconsistent results were obtained in another recent study, which indicates that COX-2 may be sufficient to cause inflammation and the subsequent cancerous aberrations. Within this study, it was found that the forced expression of COX-2 transgene under the control of a keratin-5 promoter causes spontaneous inflammation-associated transitional cell

hyperplasia and transitional cell carcinoma in urinary bladders of transgenic mice (186). Future research is needed to address the specific role of COX-2 in linking inflammation and cancer and to extend those findings to other types of inflammation-associated cancers. Arachidonic acids are the substrate for COX-2 to produce prostaglandins. Interestingly, arachidonic acids can be converted by another enzyme lipoxygenases to leukotrienes, which were suggested to be another missing link between inflammation and cancer (187).

**Hypoxia-Inducible Factor-1 $\alpha$ .** HIF-1, widely accepted as a mediator of oxygen homeostasis, is a heterodimeric transcription factor (188). HIF-1 $\alpha$  is oxygen sensitive at the protein level, whereas HIF-1 $\beta$  is constitutively expressed (189). In response to hypoxia, HIF-1 activates a wide range of hypoxia-responsive molecules, such as erythropoietin, iNOS, VEGF, glucose transporter-1, and other glycolytic enzymes (190). At sites of inflammatory lesions, hypoxia is a common feature resulting from metabolic shifts during inflammation (191). Therefore, HIF-1 may be implicated in inflammation. The role of HIF-1 in driving the progression of inflammation may be tissue specific. In a murine model of experimental colitis, HIF-1 in hypoxic epithelium attenuates clinical manifestations of inflammatory disease as an anti-inflammatory factor (192). However, HIF-1 plays an essential role in other models of inflammation, inducing leukocyte adhesion (193) and maintaining normal functions of myeloid cells recruited to sites of inflammation (194). HIF-1 also might promote chronic inflammation by preventing the hypoxic apoptosis of neutrophils and T lymphocytes (194, 195). The relationship between regulation of NF- $\kappa$ B and HIF-1 pathways is dependent on the status of cells (83, 194, 196, 197). The induction of NF- $\kappa$ B by hypoxia is dependent on the existence of HIF-1 $\alpha$  (190), whereas in several normoxic cell lines, including cancer cells, HIF-1 $\alpha$  is activated by proinflammatory cytokines, such as TNF- $\alpha$  and IL-1 $\beta$ , in a NF- $\kappa$ B-dependent manner (83, 196, 197). It is also found that COX-2 mediates IL-1 $\beta$ -induced HIF-1 $\alpha$  by its product prostaglandin E<sub>2</sub> (83). HIF-1 $\alpha$  plays an essential role in tumor development because it facilitates the phenotype of cancer cells with an enhanced glycolytic activity and increases the transcription of VEGF, a potent angiogenic factor that is important in tumor growth and metastasis (198). Although further studies are required to elucidate the interactions among proinflammatory cytokines, NF- $\kappa$ B, COX-2, and HIF-1, we hypothesize that HIF-1 is a candidate in linking inflammation and cancer. In the context of inflammation, HIF-1 induced by hypoxia may contribute to the establishment of cancer-prone chronic inflammation. Its persistent presence may result from NF- $\kappa$ B activation and/or COX-2-mediated induction by proinflammatory cytokines prevalent within the microenvironment. On the other hand, HIF-1 may also act as a promoter facilitating the development of inflammation-associated cancers.

#### *Other Promising Links between Inflammation and Cancer.*

To add even more complexity to the whole picture of inflammation and cancer, there may be still some missing parts of the long-lasting puzzle. The transcription factor signal

transducers and activators of transcription (STAT) might be one missing link. It is well known that cytokines can activate STAT family transcription factors by the signaling of Janus-activated kinases (199). The conformational change of cytokine receptor induced by the binding of their ligands causes the displacement of Janus-activated kinases, which subsequently phosphorylate and activate STAT transcription factors (200). The aforementioned IL-6, which was found to be involved in the progression of colon cancer, is a well-established inducer of STAT3. Because STAT3 has been found constitutively activated in various types of cancer (199), we speculate that the IL-6/Janus-activated kinase/STAT3 pathway might play a role in linking the inflammatory microenvironment and cancer development. Another member of the STAT family STAT5 is also activated by a wide range of cytokines, including IL-2, IL-3, IL-5, IL-7, IL-9, IL-15, and granulocyte-macrophage colony-stimulating factor, in a similar manner to that of STAT3. STAT5 is not associated with so many types of cancer as STAT3 is, and its activation seems to be specifically found in several types of leukemia (199). The function of STATs in cancer development is still under intense investigation, but it has been found that they can promote proliferation and apoptotic resistance in human myeloma cell lines (201) and contribute to the immune tolerance of tumor cells (202). Therefore, we suggest that STAT3 activation occurs after the occurrence of primary malignant cells and plays a role in promoting their development in an inflammatory microenvironment. Other than the STAT family transcription factors, there are still other possible linking molecules. The transcription factor Nrf2, which regulates a wide range of detoxifying and antioxidant genes, was identified as a critical system responding to cellular stresses (203). Interestingly, several *in vivo* studies showed that Nrf2 might play an anti-inflammatory role in inflammation (204-206). The regulatory role of Nrf2 in the resolution of inflammation was through the induction by a prostaglandin product of COX-2 (207). Nrf2 may also be induced by NO (208) and lead to the reduced susceptibility to apoptotic signals, such as TNF- $\alpha$  (209). As for carcinogenesis, Nrf2 was well established to protect against DNA damage and carcinogenesis (208, 210, 211). Moreover, nuclear factor of activated T cells (NFAT), which is expressed in both immune and nonimmune cells, plays an essential role in inflammatory responses by regulating the expression of a wide range of proinflammatory cytokines, such as IL-2, IL-3, IL-4, IL-5, IL-13, granulocyte-macrophage colony-stimulating factor, and TNF- $\alpha$  (212-215). The inhibition of NFAT in T cells resulted in a reduction of allergic pulmonary inflammation (216). In both T cells and colon carcinoma cells, NFAT was involved in the expression of COX-2, which has extensive functions in both inflammation and cancer development (217, 218). Recently, our group found that NFAT was required for TNF- $\alpha$ -induced COX-2 expression and cell transformation in mouse epidermal Cl 41 cells.<sup>1</sup> To date, the extensive roles of Nrf2 and NFAT suggest that they might be promising targets for addressing the puzzling link between inflammation and cancers.

<sup>1</sup> Yan and Huang, unpublished data.

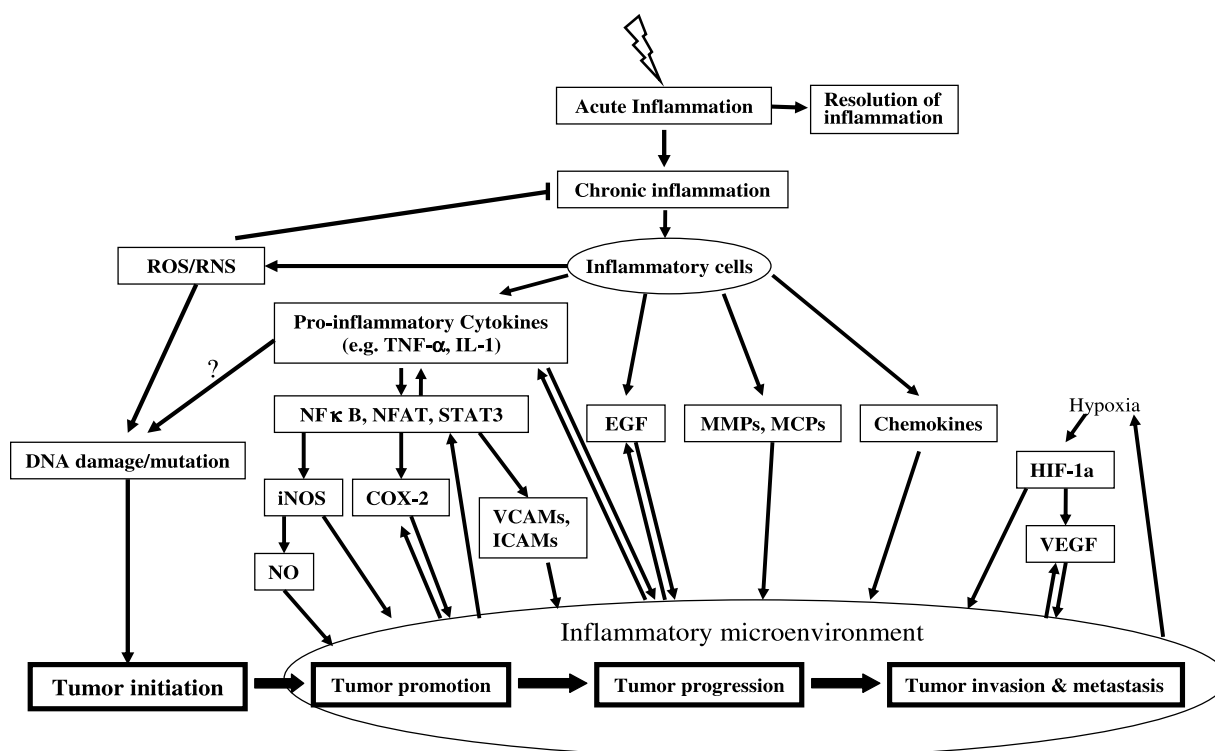


## Concluding Remarks

Inflammation and cancer are both complicated pathologic processes under the control of many driving forces rather than a single one (3, 6, 19). In Fig. 1, we summarize the mechanisms underlying the involvement of inflammation in cancer development discussed in this review, although the mechanisms underlying their association still remain unraveled. The initial inflammation involves the recruitment of a wide range of immune cells to inflamed sites (16) as well as the release of various proinflammatory cytokines and other agents. These molecules function in a coordinative manner to commence an inflammatory cascade (3). The inflammation is precisely timed; however, the aberrations in the apoptosis and phagocytosis of *in situ* inflammatory cells may lead to an unresolved chronic inflammation (219, 220). In a setting of chronic inflammation, the persistent tissue damage and cell proliferation as well as the enrichment of reactive oxygen and nitrogen species contribute to a cancer-prone microenvironment (221). Cytokines, such as migration inhibitory factor, may also protect transformed cells from being arrested by tumor suppressor gene p53 (43, 44). The sinister role of inflammatory cells and molecules still persists after the tumors have been formed, when inflammatory cells are infiltrated into tumor sites. They are involved in the circumvention of tumor cells from host immune response or play an even more direct role to facilitate angiogenesis, tumor growth, invasion, and metastasis by themselves or by inducing other effector molecules, such as MMPs (3). Tumor cells may also release cytokines and chemokines to further enhance the tumor promotion of such a subverted host immune response (3, 5, 47).

Within such a generalized model, several transcription factors, enzymes, besides cytokines and chemokines, should be taken into extensive consideration for their critical regulatory functions during this complicated process. Recently, light has been shed on NF- $\kappa$ B. NF- $\kappa$ B mediates the development of cells participating in inflammatory responses (129, 130) and, more importantly, also drives the development of chronic inflammation by regulating apoptosis of inflammatory cells (7, 142). The proposed positive feedback loop that exist between NF- $\kappa$ B and cytokines, such as TNF- $\alpha$ , may imply the role of NF- $\kappa$ B as an essential regulator in the whole network (134, 135). In addition, this putative feedback loop may partially be the reason for the persistent and prevalent existence of all these signaling molecules in inflammatory tissues and results in the enhancement of their effects in cancer development. Recently, several studies have made a great progress in delineating the role of NF- $\kappa$ B in linking inflammation and cancer. These studies illustrated that NF- $\kappa$ B at the tumor promotion stage of inflammation-associated cancer is an antiapoptotic factor, which protects transformed cells against the elimination by several endogenous apoptotic factors (157, 158). The regulatory role of NF- $\kappa$ B in its downstream molecules, such as iNOS, COX-2, and HIF-1 $\alpha$ , is also shown. These molecules themselves are pleiotropic in inflammation and cancers and thus are potential targets of the links between inflammation and cancers.

The whole story between inflammation and cancer is still far from being completely understood. For instance, the question regarding the intriguing feedback loop between cytokines and



**FIGURE 1.** Microbial infection, chemical irritation, and tissue wounding. Summary of mechanisms for the involvement of inflammation in cancer development. Tumor promotion indicates the process during which initiated cells develop into benign lesions. Tumor progression defines the process during which benign tumors progress to malignant carcinomas.

NF- $\kappa$ B is which activation is the initial event. In addition, animal models for inflammation-derived cancers and combination to molecular approaches, such as specific gene knockout mouse, will be helpful and necessary to address the questions in this field. Besides the delineation of an existing picture, another focus of our future study is to look for the missing links of this intriguing puzzle. We believe that the better clarification of mechanisms linking inflammation and cancers will be beneficial to the development of efficacious prevention and therapies of inflammation-associated cancers.

## Acknowledgments

We thank Juliana Powell for her editorial support.

## References

- Balkwill F, Mantovani A. Inflammation and cancer: back to Virchow? *Lancet* 2001;357:539–45.
- Philip M, Rowley DA, Schreiber H. Inflammation as a tumor promoter in cancer induction. *Semin Cancer Biol* 2004;14:433–9.
- Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002;420:860–7.
- Okada F. Inflammation and free radicals in tumor development and progression. *Redox Rep* 2002;7:357–68.
- Yang CR, Hsieh SL, Ho FM, Lin WW. Decoy receptor 3 increases monocyte adhesion to endothelial cells via NF- $\kappa$ B-dependent up-regulation of intercellular adhesion molecule-1, VCAM-1, and IL-8 expression. *J Immunol* 2005;174:1647–56.
- Nathan C. Points of control in inflammation. *Nature* 2002;420:846–52.
- Maiuri MC, Tajana G, Iuvone T, et al. Nuclear factor- $\kappa$ B regulates inflammatory cell apoptosis and phagocytosis in rat carrageenin-sponge implant model. *Am J Pathol* 2004;165:115–26.
- Levy BD, Clish CB, Schmidt B, Gronert K, Serhan CN. Lipid mediator class switching during acute inflammation: signals in resolution. *Nat Immunol* 2001;2:612–9.
- Hodge-Dufour J, Marino MW, Horton MR, et al. Inhibition of interferon  $\gamma$  induced interleukin 12 production: a potential mechanism for the anti-inflammatory activities of tumor necrosis factor. *Proc Natl Acad Sci U S A* 1998;95:13806–11.
- Savill J, Wyllie AH, Henson JE, Walport MJ, Henson PM, Haslett C. Macrophage phagocytosis of aging neutrophils in inflammation. Programmed cell death in the neutrophil leads to its recognition by macrophages. *J Clin Invest* 1989;83:865–75.
- Savill J, Fadok VA. Corpse clearance defines the meaning of cell death. *Nature* 2000;407:784–8.
- Savill J, Dransfield I, Gregory C, Haslett C. A blast from the past: clearance of apoptotic cells regulates immune responses. *Nat Rev Immunol* 2002;2:965–75.
- Fadok VA, Bratton DL, Konowal A, Freed PW, Westcott JY, Henson PM. Macrophages that have ingested apoptotic cells *in vitro* inhibit proinflammatory cytokine production through autocrine/paracrine mechanisms involving TGF- $\beta$ , PGE<sub>2</sub> and PAF. *J Clin Invest* 1998;101:890–8.
- McDonald PP, Fadok VA, Bratton D, Henson PM. Transcriptional and translational regulation of inflammatory mediator production by endogenous TGF- $\beta$  in macrophages that have ingested apoptotic cells. *J Immunol* 1999;163:6164–72.
- Huynh M-LN, Fadok VA, Henson PM. Phosphatidylserine-dependent ingestion of apoptotic cells promotes TGF- $\beta$  secretion and the resolution of inflammation. *J Clin Invest* 2002;109:41–50.
- Macarthur M, Hold GL, El-Omar EM. Inflammation and cancer. II. Role of chronic inflammation and cytokine polymorphisms in the pathogenesis of gastrointestinal malignancy. *Am J Physiol Gastrointest Liver Physiol* 2004;286:G515–20.
- Berenblum I. The cocarcinogenic action of croton resin. *Cancer Res* 1941;1:44–8.
- Trosko JE. Commentary: is the concept of “tumor promotion” a useful paradigm? *Mol Carcinog* 2001;30:131–7.
- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000;100:57–70.
- Schreiber H, Rowley DA. Inflammation and cancer. In: Gallin JI, Snyderman R, editors. *Inflammation: basic principles and clinical correlates*. 3rd ed. Philadelphia: Lippincott Williams & Wilkins; 1999. p. 1117–29.
- Itzkowitz SH, Yio X. Inflammation and cancer. IV. Colorectal cancer in inflammatory bowel disease: the role of inflammation. *Am J Physiol Gastrointest Liver Physiol* 2004;287:G7–17.
- Seril DN, Lia J, Yang GY, Yang CS. Oxidative stress and ulcerative colitis-associated carcinogenesis: studies in humans and animal models. *Carcinogenesis* 2003;24:353–62.
- Moody GA, Jayanthi V, Probert CS, Mac Kay H, Mayberry JF. Long term therapy with sulphasalazine protects against colorectal cancer in ulcerative colitis: a retrospective study of colorectal cancer risk and compliance with treatment in Leicestershire. *Eur J Gastroenterol Hepatol* 1996;8:1179–83.
- Eaden J, Abrams K, Ekbom A, Jackson E, Mayberry J. Colorectal cancer prevention in ulcerative colitis: a case control study. *Aliment Pharmacol Ther* 2000;14:145–53.
- Keeley D, Rees J. New guidelines in asthma management. *Br Med J* 1997;314:315–6.
- Borm PJ, Driscoll K. Particles, inflammation and respiratory tract carcinogenesis. *Toxicol Lett* 1996;88:109–13.
- Block TM, Mehta AS, Fimmel CJ, Jordan R. Molecular viral oncology of hepatocellular carcinoma. *Oncogene* 2003;22:5093–107.
- Rosin MR, Anwar WA, Ward AJ. Inflammation, chromosomal instability, and cancer: the schistosomiasis model. *Cancer Res* 1994;54:1929–33S.
- Whitcomb DC. Inflammation and cancer. V. chronic pancreatitis and pancreatic cancer. *Am J Physiol Gastrointest Liver Physiol* 2004;287:G315–9.
- Vesterinen E, Pukkala E, Timonen T, Aromaa A. Cancer incidence among 78,000 asthma patients. *Int J Epidemiol* 1993;22:976–82.
- Alavanja MC, Brownson RC, Boice JD, Hock E. Preexisting lung disease and lung cancer among nonsmoking women. *Am J Epidemiol* 1992;136:623–32.
- Wu AH, Fonthan ET, Reynolds P, et al. Previous lung disease and risk of lung cancer among lifetime nonsmoking women in the United States. *Am J Epidemiol* 1995;141:1023–32.
- Askling J, Grunewald J, Eklund A, Hillerdal G, Ekbom A. Increased risk for cancer following sarcoidosis. *Am J Respir Crit Care Med* 1999;160:1668–72.
- Carlson JA, Ambros R, Malfetano J, et al. Vulvar lichen sclerosus and squamous cell carcinoma: a cohort, case control, and investigational study with historical perspective; implications for chronic inflammation and sclerosis in the development of neoplasia. *Hum Pathol* 1998;29:932–48.
- Mayron R, Grimwood RE, Siegle RJ, Camisa C. Verrucous carcinoma arising in ulcerative lichen planus of the soles. *J Dermatol Surg Oncol* 1988;14:547–51.
- Perky L. Epidemiology of cancer of the penis. *Recent Results Cancer Res* 1977;60:97–109.
- Risch HA, Howe GR. Pelvic inflammatory disease and the risk of epithelial ovarian cancer. *Cancer Epidemiol Biomarkers Prev* 1995;4:447–51.
- Palapattu GS, Sutcliffe S, Bastian PJ, et al. Prostate carcinogenesis and inflammation: emerging insights. *Carcinogenesis* 2004;26:1170–81.
- Cordon-Cardo C, Prives C. Commentary: at the crossroads of inflammation and tumorigenesis. *J Exp Med* 1999;190:1367–70.
- Houghton J, Stoicov C, Nomura S, et al. Gastric cancer originating from bone-marrow-derived cells. *Science* 2004;306:1568–71.
- Maeda H, Akaike H. Nitric oxide and oxygen radicals in infection, inflammation, and cancer. *Biochemistry (Mosc)* 1998;63:854–65.
- Fulton AM, Loveless SE, Heppner GH. Mutagenic activity of tumor-associated macrophages in *Salmonella typhimurium* strains TA98 and TA100. *Cancer Res* 1984;44:4308–11.
- Pollard JW. Tumour-educated macrophages promote tumour progression and metastasis. *Nat Rev Cancer* 2004;4:71–8.
- Hudson JD, Shoaibi MA, Maestro R, Carnero A, Hannon GJ, Beach DH. A proinflammatory cytokine inhibits p53 tumor suppressor activity. *J Exp Med* 1999;190:1375–82.
- Petrenko O, Moll UM. Macrophage migration inhibitory factor MIF interferes with the Rb-E2F pathway. *Mol Cell* 2005;17:225–36.
- Chu FF, Esworthy RS, Chu PG, et al. Bacteria-induced intestinal cancer in mice with disrupted Gpx1 and Gpx2 genes. *Cancer Res* 2004;64:962–8.
- Lin EY, Pollard JW. Role of infiltrated leucocytes in tumor growth and spread. *Br J Cancer* 2004;90:2053–8.
- Coussens LM, Werb Z. Inflammatory cells and cancer: think different! *J Exp Med* 2001;193:F23–6.
- Clark WH, Jr., Elder DE, Guerry D IV, et al. Model predicting survival

- in stage I melanoma based on tumor progression. *J Natl Cancer Inst* 1989;81:1893–904.
50. Clemente CG, Mihm MC, Jr., Bufalino R, Zurrida S, Collini P, Cascinelli N. Prognostic value of tumor infiltrating lymphocytes in the vertical growth phase of primary cutaneous melanoma. *Cancer* 1996;77:1303–10.
  51. Naito Y, Saito K, Shiiba K, et al. CD8<sup>+</sup> T cells infiltrated within cancer cell nests as a prognostic factor in human colorectal cancer. *Cancer Res* 1998;58:3491–4.
  52. Nakano O, Sato M, Naito Y, et al. Proliferative activity of intratumoral CD8(+) T-lymphocytes as a prognostic factor in human renal cell carcinoma: clinicopathologic demonstration of antitumor immunity. *Cancer Res* 2001;61:5132–6.
  53. Zhang L, Conejo-Garcia JR, Katsaros D, et al. Intratumoral T cells, recurrence, and survival in epithelial ovarian cancer. *N Eng J Med* 2003;348:203–13.
  54. Dunn G, Bruce A, Ikeda H, Old L, Schreiber R. Cancer immunoediting: from immunosurveillance to tumor escape. *Nat Immunol* 2002;3:991–8.
  55. Brigati C, Noonan DM, Albini A, Benelli R. Tumors and inflammatory infiltrates: friends or foes? *Clin Exp Metastasis* 2002;19:247–58.
  56. Tsung K, Dolan JP, Tsung YL, Norton JA. Macrophages as effector cells in interleukin 12-induced T cell-dependent tumor rejection. *Cancer Res* 2002;62:5069–75.
  57. Mihm M, Clemente C, Cascinelli N. Tumor infiltrating lymphocytes in lymph node melanoma metastases—a histopathologic prognostic indicator and an expression of local immune response. *Lab Invest* 1996;74:43–7.
  58. Smyth MJ, Cretney E, Kershaw MH, Hayakawa Y. Cytokines in cancer immunity and immunotherapy. *Immunol Rev* 2004;202:275–93.
  59. Khong HT, Restifo NP. Natural selection of tumor variants in the generation of “tumor escape” phenotypes. *Nat Immunol* 2002;3:999–1005.
  60. Saji H, Koike M, Yamori T, et al. Significant correlation of monocyte chemoattractant protein-1 expression with neovascularization and progression of breast carcinoma. *Cancer* 2001;92:3282–9.
  61. Leek RD, Harris AL. Tumor-associated macrophages in breast cancer. *J Mammary Gland Biol Neoplasia* 2002;7:177–89.
  62. Lin EY, Gouon-Evans V, Nguyen AV, Pollard JW. The macrophage growth factor, CSF-1, in mammary gland development and cancer. *J Mammary Gland Biol Neoplasia* 2002;7:147–62.
  63. Kacinski BM. CSF-1 and its receptor in breast carcinomas and neoplasms of the female reproductive tract. *Mol Reprod Dev* 1997;46:71–4.
  64. Amann B, Perabo FG, Wirger A, Hugenschmidt H, Schultze-Seemann W. Urinary levels of monocyte chemo-attractant protein-1 correlate with tumour stage and grade in patients with bladder cancer. *Br J Urol* 1998;82:118–21.
  65. Valkovic T, Lucin K, Krstulja M, Dobi-Babic R, Jonjic N. Expression of monocyte chemotactic protein-1 in human invasive ductal breast cancer. *Pathol Res Pract* 1998;194:335–40.
  66. Tsujitani S, Kakeji Y, Watanabe A, Kohnoe S, Maehara Y, Sugimachi K. Infiltration of dendritic cells in relation to tumor invasion and lymph node metastasis in human gastric cancer. *Cancer* 1990;66:2012–6.
  67. Troy A, Davidson P, Atkinson C, Hart D. Phenotypic characterization of the dendritic cell infiltrate in prostate cancer. *J Urol* 1998;160:214–9.
  68. Lespagnard L, Gancberg D, Rouas G, et al. Tumor-infiltrating dendritic cells in adenocarcinomas of the breast: a study of 143 neoplasms with a correlation to usual prognostic factors and to clinical outcome. *Int J Cancer* 1999;84:309–14.
  69. Ribatti D, Vacca A, Nico B, Crivellato E, Roncali L, Dammacco F. The role of mast cells in tumour angiogenesis. *Br J Haematol* 2001;115:514–21.
  70. Nielsen HJ, Hansen U, Christensen IJ, Reimert CM, Brunner N, Moesgaard F. Independent prognostic value of eosinophil and mast cell infiltration in colorectal cancer tissue. *J Pathol* 1999;189:487–95.
  71. Bellocq A, Antoine M, Flahault A, et al. Neutrophil alveolitis in bronchioloalveolar carcinoma: induction by tumor-derived interleukin-8 and relation to clinical outcome. *Am J Pathol* 1998;152:83–92.
  72. Caruso RA, Bellocchio R, Pagano M, Bertoli G, Rigoli L, Infrerra C. Prognostic value of intratumoral neutrophils in advanced gastric carcinoma in a high-risk area in northern Italy. *Mod Pathol* 2002;15:831–7.
  73. Elgert K, Alleva D, Mullins D. Tumor-induced immune dysfunction: the macrophage connection. *J Leukoc Biol* 1998;64:275–90.
  74. Grimschaw MJ, Wilson JL, Balkwill FR. Endothelin-2 is a macrophage chemoattractant: implications for macrophage distribution in tumors. *Eur J Immunol* 2002;32:2393–400.
  75. Foekens JA, Peters HA, Look MP, et al. The urokinase system of plasminogen activation and prognosis in 2780 breast cancer patients. *Cancer Res* 2000;60:636–43.
  76. Hildenbrand R, Glienke W, Magdolen V, Graeff H, Stutte HJ, Schmitt M. Urokinase receptor localization in breast cancer and benign lesions assessed by *in situ* hybridization and immunohistochemistry. *Histochem Cell Biol* 1998;110:27–32.
  77. Fox SB, Taylor M, Grondahl-Hansen J, Kakolyris S, Gatter KC, Harris AL. Plasminogen activator inhibitor-1 as a measure of vascular remodelling in breast cancer. *J Pathol* 2001;195:236–43.
  78. Knoop A, Andreassen PA, Andersen JA, et al. Prognostic significance of urokinase-type plasminogen activator and plasminogen activator inhibitor-1 in primary breast cancer. *Br J Cancer* 1998;77:932–40.
  79. Hildenbrand R, Wolf G, Bohme B, Bleyl U, Steinborn A. Urokinase plasminogen activator receptor (CD87) expression of tumor-associated macrophages in ductal carcinoma *in situ*, breast cancer, and resident macrophages of normal breast tissue. *J Leukoc Biol* 1999;66:40–9.
  80. Bando H, Toi M. Tumor angiogenesis, macrophages, and cytokines. *Adv Exp Med Biol* 2000;476:267–84.
  81. Leek RD, Hunt NC, Landers RJ, Lewis CE, Royds JA, Harris AL. Macrophage infiltration is associated with VEGF and EGFR expression in breast cancer. *J Pathol* 2000;190:430–6.
  82. Barleon B, Sozzani S, Zhou D, Weich HA, Mantovani A, Marme D. Migration of human monocytes in response to vascular endothelial growth factor (VEGF) is mediated via the VEGF receptor flt-1. *Blood* 1996;87:3336–43.
  83. Jung YJ, Isaacs JS, Lee S, Trepel J, Neckers L. IL-1 $\beta$ -mediated up-regulation of HIF-1 $\alpha$  via an NF $\kappa$ B/COX-2 pathway identifies HIF-1 as a critical link between inflammation and oncogenesis. *FASEB J* 2003;17:2115–7.
  84. Coussens LM, Raymond WW, Bergers G, et al. Inflammatory mast cells up-regulate angiogenesis during squamous epithelial carcinogenesis. *Genes Dev* 1999;13:1382–97.
  85. Ogmundsdottir HM, Petursdottir I, Gudmundsdottir I. Interactions between the immune system and breast cancer. *Acta Oncol* 1995;34:647–50.
  86. O’Sullivan C, Lewis CE, Harris AL, McGee JO. Secretion of epidermal growth factor by macrophages associated with breast carcinoma. *Lancet* 1993;342:872–3.
  87. Wyckoff JB, Segall JE, Condeelis JS. The collection of the motile population of cells from a living tumor. *Cancer Res* 2000;60:5401–4.
  88. Wyckoff J, Wang W, Lin EY, et al. A paracrine loop between tumor cells and macrophages is required for tumor cell migration in mammary tumors. *Cancer Res* 2004;64:7022–9.
  89. Fijneman RJ, Vos M, Berkhof J, Demant P, Kraal G. Genetic analysis of macrophage characteristics as a tool to identify tumor susceptibility genes: mapping of three macrophage-associated risk inflammatory factors, Marif1, Marif2, and Marif3. *Cancer Res* 2004;64:3458–64.
  90. Meininger CJ, Zetter BR. Mast cells and angiogenesis. *Semin Cancer Biol* 1992;3:73–9.
  91. Qu Z, Liebler JM, Powers MR, et al. Mast cells are a major source of basic fibroblast growth factor in chronic inflammation and cutaneous hemangioma. *Am J Pathol* 1995;147:564–73.
  92. Hiromatsu Y, Toda S. Mast cells and angiogenesis. *Microsc Res Tech* 2003;60:64–9.
  93. Scapini P, Nesi L, Morini M, et al. Generation of biologically active angiotensin kringles 1–3 by activated human neutrophils. *J Immunol* 2002;168:5798–804.
  94. Schaidt H, Oka M, Bogenrieder T, et al. Differential response of primary and metastatic melanomas to neutrophils attracted by IL-8. *Int J Cancer* 2003;103:335–43.
  95. Haqqani AS, Sandhu JK, Birnboim HC. Expression of interleukin-8 promotes neutrophil infiltration and genetic instability in mutator tumors. *Neoplasia* 2000;2:561–8.
  96. Daniel D, Meyer-Morse N, Bergsland EK, Dehne K, Coussens LM, Hanahan D. Immune enhancement of skin carcinogenesis by CD4<sup>+</sup> T cells. *J Exp Med* 2003;197:1017–28.
  97. Bromwich EJ, McArdle PA, Canna K, et al. The relationship between T-lymphocyte infiltration, stage, tumour grade and survival in patients undergoing curative surgery for renal cell cancer. *Br J Cancer* 2003;89:1906–8.
  98. Canna K, McArdle PA, McMillan DC, et al. The relationship between tumour T-lymphocyte infiltration, the systematic inflammatory response and survival in patients undergoing curative resection for colorectal cancer. *Br J Cancer* 2005;92:651–4.

99. Becker C, Fantini MC, Schramm C, et al. TGF- $\beta$  suppresses tumor progression in colon cancer by inhibition of IL-6 trans-signaling. *Immunity* 2004; 21:491–501.
100. Dranoff G. Cytokines in cancer pathogenesis and cancer therapy. *Nat Rev Cancer* 2004;4:11–22.
101. Chung YC, Chang YF. Serum interleukin-6 levels reflect the disease status of colorectal cancer. *J Surg Oncol* 2003;83:222–6.
102. Schneider MR, Hoefflich A, Fischer JR, Wolf E, Sordat B, Lahm H. Interleukin-6 stimulates colonogenic growth of primary and metastatic human colon carcinoma cells. *Cancer Lett* 2000;151:31–8.
103. Tebbutt NC, Giraud AS, Inglese M, et al. Reciprocal regulation of gastrointestinal homeostasis by SHP2 and STAT-mediated trefoil gene activation in gp130 mutant mice. *Nat Med* 2002;8:1089–97.
104. Balkwill F. Tumor necrosis factor or tumor promoting factor? *Cytokine Growth Factor Rev* 2002;13:135–41.
105. Huang C, Li J, Ma WY, Dong Z. JNK activation is required for JB6 cell transformation induced by tumor necrosis factor- $\alpha$  but not by 12-*O*-tetradecanoylphorbol-13-acetate. *J Biol Chem* 1999;274:29672–6.
106. Micheau O, Tschopp J. Induction of TNF receptor I-mediated apoptosis via two sequential signaling complexes. *Cell* 2003;114:181–90.
107. Locksley RM, Killeen N, Lenardo MJ. The TNF and TNF receptor superfamilies: integrating mammalian biology. *Cell* 2001;104:487–501.
108. Jaiswal M, LaRusso NF, Burgart LJ, Gores GJ. Inflammatory cytokines induce DNA damage and inhibit DNA repair in cholangiocarcinoma cells by a nitric oxide-dependent mechanism. *Cancer Res* 2000;60:184–90.
109. Wu S, Boyer CM, Whitaker RS, et al. Tumor necrosis factor  $\alpha$  as an autocrine and paracrine growth factor for ovarian cancer: monokine induction of tumor cell proliferation and tumor necrosis factor  $\alpha$  expression. *Cancer Res* 1993; 53:1939–44.
110. Leek RD, Landers R, Fox SB, Ng F, Harris AL, Lewis CE. Association of tumour necrosis factor  $\alpha$  and its receptors with thymidine phosphorylase expression in invasive breast carcinoma. *Br J Cancer* 1998;77:2246–51.
111. Aggarwal BB. Signaling pathways of the TNF superfamily: a double-edged sword. *Nat Rev Immunol* 2003;3:745–56.
112. Wilson J, Balkwill F. The role of cytokines in the epithelial cancer microenvironment. *Semin Cancer Biol* 2002;12:113–20.
113. Ardestani SK, Inerra P, Solkoff D, Watson RR. The role of cytokines and chemokines on tumor progression: a review. *Cancer Detect Prev* 1999;23: 215–25.
114. Strieter RM. Chemokines: not just leukocyte chemoattractants in the promotion of cancer. *Nat Immunol* 2001;2:285–6.
115. Luca M, Huang S, Gershenwald JE, Singh RK, Reich R, Bar-Eli M. Expression of interleukin-8 by human melanoma cells up-regulates MMP-2 activity and increases tumor growth and metastasis. *Am J Pathol* 1997;151: 1105–13.
116. Inoue K, Slaton JW, Eve BY, et al. Interleukin-8 expression regulates tumorigenicity and metastases in androgen-independent prostate cancer. *Clin Cancer Res* 2000;6:2104–19.
117. Müller A, Homey B, Soto H, et al. Involvement of chemokine receptors in breast cancer metastasis. *Nature* 2001;410:50–6.
118. Scotton CJ, Wilson JL, Milliken D, Stamp G, Balkwill FR. Epithelial cancer cell migration: a role for chemokine receptors. *Cancer Res* 2001;61: 4961–5.
119. Sparmann A, Bar-Sagi D. Ras-induced interleukin-8 expression plays a critical role in tumor growth and angiogenesis. *Cancer Cell* 2004;6:447–58.
120. Karin M, Ben-Neriah Y. Phosphorylation meets ubiquitination: the control of NF- $\kappa$ B activity. *Annu Rev Immunol* 2000;18:621–63.
121. Viatour P, Merville M-P, Bours V, Chariot A. Phosphorylation of NF- $\kappa$ B and I $\kappa$ B proteins: implications in cancer and inflammation. *Trends Biochem Sci* 2005;30:43–52.
122. Kato T, Jr., Delhase M, Hoffmann A, Karin M. CK2 is a C-terminal I $\kappa$ B kinase responsible for NF- $\kappa$ B activation during the UV response. *Mol Cell* 2003; 12:829–39.
123. Tergaonkar V, Bottero V, Ikawa M, Li Q, Verma IM. I $\kappa$ B kinase-independent I $\kappa$ B $\alpha$  degradation pathway: functional NF- $\kappa$ B activity and implications for cancer therapy. *Mol Cell Biol* 2003;23:8070–83.
124. Baldwin A. The NF- $\kappa$ B and I $\kappa$ B proteins: new discoveries and insights. *Annu Rev Immunol* 1996;14:649–83.
125. Sweeney C, Li L, Shanmugam R, et al. Nuclear factor- $\kappa$ B is constitutively activated in prostate cancer *in vitro* and is overexpressed in prostatic intraepithelial neoplasia and adenocarcinoma of the prostate. *Clin Cancer Res* 2004;10:5501–7.
126. Hu J, Haseebuddin M, Young M, Colburn NH. Suppression of p65 phosphorylation coincides with inhibition of I $\kappa$ B $\alpha$  polyubiquitination and degradation. *Mol Carcinog* 2005;44:274–84.
127. Hu J, Nakano H, Sakurai H, Colburn NH. Insufficient p65 phosphorylation at S536 specifically contributes to the lack of NF- $\kappa$ B activation and transformation in resistant JB6 cells. *Carcinogenesis* 2004;25:1991–2003.
128. Karin M, Cao Y, Greten FR, Li ZW. NF- $\kappa$ B in cancer: from innocent bystander to major culprit. *Nat Rev Cancer* 2002;2:301–10.
129. Caamano J, Hunter C. NF- $\kappa$ B family of transcription factors: central regulators of innate and adaptive immune functions. *Clin Microbiol Rev* 2002;15: 414–29.
130. Li Q, Verma IM. NF- $\kappa$ B regulation in the immune system. *Nat Rev Immunol* 2002;2:725–34.
131. Senfleben U, Li ZW, Baud V, Karin M. IKK $\beta$  is essential for protecting T cells from TNF $\alpha$ -induced apoptosis. *Immunity* 2001;14:217–30.
132. Mora A, Youn J, Keegan A, Boothby MR. NF- $\kappa$ B/Rel participation in the lymphokine-dependent proliferation of T lymphoid cells. *J Immunol* 2001;166: 2218–27.
133. Hettmann T, DiDonato J, Karin M, Leiden JM. An essential role for nuclear factor- $\kappa$ B in promoting double-positive thymocyte apoptosis. *J Exp Med* 1999; 189:145–58.
134. Tak PP, Firestein GS. NF- $\kappa$ B: a key role in inflammatory diseases. *J Clin Invest* 2001;107:7–11.
135. Perkins ND. The Rel/NF- $\kappa$ B family: friend and foe. *Trends Biochem Sci* 2000;25:434–40.
136. Heyninck K, Beyaert R. A20 inhibits NF- $\kappa$ B activation by dual ubiquitin-editing functions. *Trends Biochem Sci* 2005;30:1–4.
137. Pasparakis M, Courtois G, Hafner M, et al. TNF-mediated inflammatory skin disease in mice with epidermis-specific deletion of IKK2. *Nature* 2002;417:861–6.
138. Chen LW, Egan L, Li ZW, Greten FR, Kagnoff MF, Karin M. The two faces of IKK and NF- $\kappa$ B inhibition: prevention of systematic inflammation but increased local injury following intestinal ischemia-reperfusion. *Nat Med* 2003;9:575–81.
139. Hollenbach E, Vieth M, Roessner A, Neumann M, Malfetheriner P, Naumann M. Inhibition of RICK/nuclear factor- $\kappa$ B and p38 signaling attenuates the inflammatory response in a murine model of Crohn disease. *J Biol Chem* 2005;280:14981–8.
140. Bakkouri KE, Wullaert A, Haegman M, Heyninck K, Beyaert R. Adenoviral gene transfer of the NF- $\kappa$ B inhibitory protein ABIN-1 decreases allergic airway inflammation in a murine asthma model. *J Biol Chem* 2005; 280:17938–44.
141. Cataisson C, Pearson AJ, Torgerson S, Nedospasov SA, Yuspa SH. Protein kinase C $\alpha$ -mediated chemotaxis of neutrophils requires NF- $\kappa$ B activity but is independent of TNF $\alpha$  signaling in mouse skin *in vivo*. *J Immunol* 2005;174: 1686–92.
142. Francois S, Benna JE, Dang PMC, Pedruzzi E, Gougerot-Pocidallo M-A, Elbim C. Inhibition of neutrophil apoptosis by TLR agonists in whole blood: involvement of the phosphoinositide 3-kinase/Akt and NF- $\kappa$ B signaling pathways, leading to increased levels of MCL-1, A1, and phosphorylated bad. *J Immunol* 2005;174:3633–42.
143. Lin A, Karin M. NF- $\kappa$ B in cancer: a marked target. *Semin Cancer Biol* 2003;13:107–14.
144. Karin M, Lin A. NF- $\kappa$ B at the crossroads of life and death. *Nat Immunol* 2002;3:221–7.
145. Piva R, Gianferretti P, Ciucci A, Taulli R, Belardo G, Santoro MG. 15-Deoxy- $\Delta^{12,14}$ -prostaglandin J<sub>2</sub> induces apoptosis in human malignant B cells: an effect associated with inhibition of NF- $\kappa$ B activity and down-regulation of antiapoptotic proteins. *Blood* 2005;105:1750–8.
146. Stoffel A, Chaurushiya M, Singh B, Levine AJ. Activation of NF- $\kappa$ B and inhibition of p53-mediated apoptosis by AP12/mucosa-associated lymphoid tissue 1 fusions promote oncogenesis. *Proc Natl Acad Sci U S A* 2004;101:9079–84.
147. Guttridge DC, Albanese C, Reuther JY, Pestell RG, Baldwin AS, Jr. NF- $\kappa$ B controls cell growth and differentiation through transcriptional regulation of cyclin D1. *Mol Cell Biol* 1999;19:5785–99.
148. Hinz M, Krappmann D, Eichten A, Heder A, Scheidereit C, Strauss M. NF- $\kappa$ B function in growth control: regulation of cyclin D1 expression and G<sub>0</sub>-G<sub>1</sub>-to-S-phase transition. *Mol Cell Biol* 1999;19:2690–8.
149. Huber MA, Azoitei N, Baumann B, et al. NF- $\kappa$ B is essential for epithelial-mesenchymal transition and metastasis in a model of breast cancer progression. *J Clin Invest* 2004;114:569–81.



150. Preciado D, Caicedo E, Jhanjee R, et al. *Pseudomonas aeruginosa* lipopolysaccharide induction of keratinocyte proliferation, NF- $\kappa$ B, and cyclin D1 is inhibited by indomethacin. *J Immunol* 2005;174:2964–73.
151. Chen CC, Rosenbloom CL, Anderson DC, Manning AM. Selective inhibition of E-selectin, vascular cell adhesion molecule-1, and intercellular adhesion molecule-1 expression by inhibitors of I $\kappa$ B $\alpha$  phosphorylation. *J Immunol* 1995;155:3538–45.
152. Furbert-Harris PM, Parish-Gause D, Hunter KA, et al. Activated eosinophils upregulate the metastasis suppressor molecule E-cadherin on prostate tumor cells. *Cell Mol Biol* 2003;49:1009–16.
153. Alexiou D, Karayiannakis AJ, Syrigos KN, et al. Clinical significance of serum levels of E-selectin, intercellular adhesion molecule-1, and vascular cell adhesion molecule-1 in gastric cancer patients. *Am J Gastroenterol* 2003;98:478–85.
154. Esposito V, Groeger AM, De Luca L, et al. Expression of surface protein receptors in lung cancer. *Anticancer Res* 2002;22:4039–43.
155. O'Hanlon DM, Fitzsimons H, Lynch J, Tormey S, Malone C, Given HF. Soluble adhesion molecules (E-selectin, ICAM-1, and VCAM-1) in breast carcinoma. *Eur J Cancer* 2002;38:2252–7.
156. Niu J, Li Z, Peng B, Chiao PJ. Identification of an autoregulatory feedback pathway involving interleukin-1 $\alpha$  in induction of constitutive NF- $\kappa$ B activation in pancreatic cancer cells. *J Biol Chem* 2004;279:16452–62.
157. Pikarsky E, Porat RM, Stein I, et al. NF- $\kappa$ B functions as a tumor promoter in inflammation-associated cancer. *Nature* 2004;431:461–6.
158. Greten FR, Eckmann L, Greten TF, et al. IKK $\beta$  links inflammation and tumorigenesis in a mouse model of colitis-associated cancer. *Cell* 2004;118:285–96.
159. Ghosh S, Karin M. Missing pieces in the NF- $\kappa$ B puzzle. *Cell* 2002;109: S81–96.
160. Luo JL, Maeda S, Hsu LC, Yagita H, Karin M. Inhibition of NF- $\kappa$ B in cancer cells converts inflammation-induced tumor growth mediated by TNF $\alpha$  to TRAIL-mediated tumor regression. *Cancer Cell* 2004;6:297–305.
161. Kim YH, Woo KJ, Lim JH, et al. 8-Hydroxyquinoline inhibits iNOS expression and nitric oxide production by down-regulating LPS-induced activity of NF- $\kappa$ B and C/EBP $\beta$  in Raw 264.7 cells. *Biochem Biophys Res Commun* 2005;329:591–7.
162. Chen T, Nines RG, Peschke SM, Kresty LA, Stoner GD. Chemopreventive effects of a selective nitric oxide synthase inhibitor on carcinogen-induced rat esophageal tumorigenesis. *Cancer Res* 2004;64:3714–7.
163. Hussain SP, Trivers GE, Hofseth LJ, et al. Nitric oxide, a mediator of inflammation, suppresses tumorigenesis. *Cancer Res* 2004;64:6849–53.
164. Hofseth LJ, Hussain SP, Wogan GN, Harris CC. Nitric oxide in cancer and chemoprevention. *Free Radic Biol Med* 2003;34:955–68.
165. Colón AL, Menchén LA, Hurtado O, et al. Implication of TNF- $\alpha$  convertase (TACE/ADAM17) in inducible nitric oxide synthase expression and inflammation in an experimental model of colitis. *Cytokine* 2001;16:220–6.
166. Goodman JE, Hofseth LJ, Hussain SP, Harris CC. Nitric oxide and p53 in cancer-prone chronic inflammation and oxyradical overload disease. *Environ Mol Mutagen* 2004;44:3–9.
167. Rao CV. Nitric oxide signaling in colon cancer chemoprevention. *Mutat Res* 2004;555:107–19.
168. Hofseth LJ, Saito S, Hussain SP, et al. Nitric oxide-induced cellular stress and p53 activation in chronic inflammation. *Proc Natl Acad Sci U S A* 2003;100: 143–8.
169. Tatemichi M, Sawa T, Gilbert I, Tazawa H, Katoh T, Ohshima H. Increased risk of internal type of gastric adenocarcinoma in Japanese women associated with long forms of CCTTT pentanucleotide repeat in the inducible nitric oxide synthase promoter. *Cancer Lett* 2005;217:197–202.
170. Williams CS, Mann M, DuBois RN. The role of cyclooxygenases in inflammation, cancer, and development. *Oncogene* 1999;18:7908–16.
171. Steele VE, Hawk ET, Viner JL, Lubert RA. Mechanisms and applications of non-steroidal anti-inflammatory drugs in the chemoprevention of cancer. *Mutat Res* 2003;523–524:137–44.
172. Buskens CJ, Van Rees BP, Sivula A, et al. Prognostic significance of elevated cyclooxygenase-2 expression in patients with adenocarcinoma of the esophagus. *Gastroenterology* 2002;122:1800–7.
173. Farrow DC, Vaughan TL, Hansten PD, et al. Use of aspirin and other nonsteroidal anti-inflammatory drugs and risk of esophageal and gastric cancer. *Cancer Epidemiol Biomarkers Prev* 1998;7:97–102.
174. Giardiello FM, Offerhaus GJ, DuBois RN. The role of nonsteroidal anti-inflammatory drugs in colorectal cancer prevention. *Eur J Cancer* 1995;31A: 1071–6.
175. Tsujii M, Kawano S, Dubois RN. Cyclooxygenase-2 expression in human colon cancer cells increases metastatic potential. *Proc Natl Acad Sci U S A* 1997; 94:3336–40.
176. Prescott SM, Fitzpatrick FA. Cyclooxygenase-2 and carcinogenesis. *Biochim Biophys Acta* 2000;1470:M69–78.
177. Eberhart CE, Coffey RJ, Radhika A, Giardiello FM, Ferrenbach S, Dubois RN. Upregulation of cyclooxygenase-2 gene expression in human colorectal adenomas and adenocarcinomas. *Gastroenterology* 1994;107:1183–8.
178. Ristimäki A, Honkanen N, Jankala H, Sipponen P, Harkonen M. Expression of cyclooxygenase 2 in human gastric cancer. *Cancer Res* 1997;57:1276–80.
179. Hwang D, Scollard D, Byrne J, Levine E. Expression of cyclooxygenase-1 and cyclooxygenase-2 in human breast cancer. *J Natl Cancer Inst* 1998;90: 455–60.
180. Okami J, Yamamoto H, Fujiwara Y, et al. Overexpression of cyclooxygenase-2 in carcinoma of the pancreas. *Clin Cancer Res* 1999;5:2018–24.
181. Hida T, Yatabe Y, Achiwa H, et al. Increased expression of cyclooxygenase 2 occurs frequently in human lung cancers, specifically in adenocarcinomas. *Cancer Res* 1998;58:3761–4.
182. Martey CA, Pollock SJ, Turner CK, et al. Cigarette smoke induces cyclooxygenase-2 and microsomal prostaglandin E<sub>2</sub> synthase in human lung fibroblasts: implications for lung inflammation and cancer. *Am J Physiol Lung Cell Mol Physiol* 2004;287:L981–91.
183. Oyama K, Fujimura T, Ninomiya I, et al. A COX-2 inhibitor prevents esophageal inflammation-metaplasia-adenocarcinoma sequence in rats. *Carcinogenesis* 2004;26:565–70.
184. Abdalla SI, Sanderson IR, Fitzgerald RC. Effect of inflammation on cyclooxygenase-2 expression in benign and malignant oesophageal cells. *Carcinogenesis* 2005;26:1627–33.
185. Wang W, Bergh A, Damber JE. Cyclooxygenase-2 expression correlates with local chronic inflammation and tumor neovascularization in human prostate cancer. *Clin Cancer Res* 2005;11:3250–6.
186. Klein RD, Van Pelt CS, Sabichi AL, et al. Transitional cell hyperplasia and carcinomas in urinary bladders of transgenic mice with keratin 5 promoter-driven cyclooxygenase-2 overexpression. *Cancer Res* 2005;65:1808–13.
187. Dubois RN. Leukotriene A<sub>4</sub> signaling, inflammation, and cancer. *J Natl Cancer Inst* 2003;95:1028–9.
188. Wang GL, Jiang BH, Rue EA, Semenza GL. Hypoxia-inducible factor 1 is a basic-helix-loop-helix-PAS heterodimer regulated by cellular O<sub>2</sub> tension. *Proc Natl Acad Sci U S A* 1995;92:5510–4.
189. Salceda S, Caro J. Hypoxia-inducible factor 1 $\alpha$  (HIF-1 $\alpha$ ) protein is rapidly degraded by the ubiquitin-proteasome system under normoxic conditions. Its stabilization by hypoxia depends on redox-induced changes. *J Biol Chem* 1997; 272:22642–7.
190. Semenza GL. Regulation of mammalian O<sub>2</sub> homeostasis by hypoxia-inducible factor 1. *Annu Rev Cell Dev Biol* 1999;15:551–78.
191. Kong T, Eltzschig HK, Karhausen J, Colgan SP, Shelley CS. Leukocyte adhesion during hypoxia is mediated by HIF-1-dependent induction of  $\beta_2$  integrin gene expression. *Proc Natl Acad Sci U S A* 2004;101:10440–5.
192. Karhausen J, Furuta GT, Tomaszewski JE, Johnson RS, Colgan SP, Haase VH. Epithelial hypoxia-inducible factor-1 is protective in murine experimental colitis. *J Clin Invest* 2004;114:1098–106.
193. Cramer T, Yamanishi Y, Clausen BE, et al. HIF-1 $\alpha$  is essential for myeloid cell-mediated inflammation. *Cell* 2003;112:645–57.
194. Walmsley SR, Print C, Farahi N, et al. Hypoxia-induced neutrophil survival is mediated by HIF-1 $\alpha$ -dependent NF- $\kappa$ B activity. *J Exp Med* 2005; 201:105–15.
195. Makino Y, Nakamura H, Ikeda E, et al. Hypoxia-inducible factor regulates survival of antigen receptor-driven T cells. *J Immunol* 2003;171:6534–40.
196. Zhou J, Schmid T, Brüne B. Tumor necrosis factor- $\alpha$  causes accumulation of a ubiquitinated form of hypoxia inducible factor-1 $\alpha$  through a nuclear factor- $\kappa$ B-dependent pathway. *Mol Biol Cell* 2003;14:2216–25.
197. Jung Y, Isaacs JS, Lee S, Trepel J, Liu ZG, Neckers L. Hypoxia-inducible factor induction by tumor necrosis factor in normoxic cells requires receptor-interacting protein-dependent nuclear factor  $\kappa$ B activation. *Biochem J* 2003;370:1011–7.
198. Jain RK. Tumor angiogenesis and accessibility: role of vascular endothelial growth factor. *Semin Oncol* 2002;29:3–9.
199. Hodge DR, Hurt EM, Farrar WL. The role of IL-6 and STAT3 in inflammation and cancer. *Eur J Cancer* 2005;41:2502–12.
200. Yang J, Chatterjee-Kishore M, Staugaitis SM, et al. Novel roles of unphosphorylated STAT3 in oncogenesis and transcriptional regulation. *Cancer Res* 2005;65:939–47.

201. Hodge DR, Xiao W, Wang LH, Li D, Farrar WL. Activating mutations in STAT3 and STAT5 differentially affect cellular proliferation and apoptotic resistance in multiple myeloma cells. *Cancer Biol Ther* 2004;3:483–9.
202. Wang T, Niu G, Kortylewski M, et al. Regulation of the innate and adaptive immune responses by Stat-3 signaling in tumor cells. *Nat Med* 2004;10:48–54.
203. Motohashi H, Yamamoto M. Nrf2-Keap1 defines a physiologically important stress response mechanism. *Trends Mol Med* 2004;10:549–57.
204. Cho HY, Reddy SPM, Yamamoto M, Kleeberger SR. The transcription factor NRF2 protects pulmonary fibrosis. *FASEB J* 2004;18:1258–60.
205. Rangasamy T, Cho CY, Thimmulappa RK, et al. Genetic ablation of Nrf2 enhances susceptibility to cigarette smoke-induced emphysema in mice. *J Clin Invest* 2004;114:1248–59.
206. Braun S, Hanselmann C, Gassmann MG, et al. Nrf2 transcription factor, a novel target of keratinocyte growth factor action which regulates gene expression and inflammation in the healing skin wound. *Mol Cell Biol* 2002;22:5492–505.
207. Itoh K, Mochizuki M, Ishii Y, et al. Transcription factor Nrf2 regulates inflammation by mediating the effect of 15-deoxy- $\Delta^{12,14}$ -prostaglandin  $J_2$ . *Mol Cell Biol* 2004;24:36–45.
208. Buckley BJ, Marshall ZM, Whorton AR. Nitric oxide stimulates Nrf2 nuclear translocation in vascular endothelium. *Biochem Biophys Res Commun* 2003;307:973–9.
209. Morito N, Yoh K, Itoh K, et al. Nrf2 regulates the sensitivity of death receptor signals by affecting intracellular glutathione levels. *Oncogene* 2003;22:9275–81.
210. Bae I, Fan S, Meng Q, et al. BRCA1 induces antioxidant gene expression and resistance to oxidative stress. *Cancer Res* 2004;64:7893–909.
211. Kwak MK, Wakabayashi N, Itoh K, Motohashi H, Yamamoto M, Kensler TW. Modulation of gene expression by cancer chemopreventive dithiolethiones through the Keap1-Nrf2 pathway. *J Biol Chem* 2003;278:8135–45.
212. Rao A, Luo C, Hogan PG. Transcription factors of the NFAT family: regulation and function. *Annu Rev Immunol* 1997;15:707–47.
213. Crabtree GR. Generic signals and specific outcomes: signaling through  $Ca^{2+}$ , calcineurin and NFAT. *Cell* 1999;96:611–4.
214. Kiani A, Rao A, Aramburu J. Manipulating immune responses with immunosuppressive agents that target NFAT. *Immunity* 2000;12:359–72.
215. Chen J, Amasaki Y, Kamogawa Y, et al. Role of NFATx (NFAT4/NFATc3) in expression of immunoregulatory genes in murine peripheral  $CD4^{+}$  T cells. *J Immunol* 2003;170:3109–17.
216. Diehl S, Krah T, Rinaldi L, Norton R, Irvin CG, Rincón M. Inhibition of NFAT specifically in T cells prevents allergic pulmonary inflammation. *J Immunol* 2004;172:3597–603.
217. Duque J, Fresno M, Iniguez MA. Expression and function of the nuclear factor of activated T cells in colon carcinoma cells. *J Biol Chem* 2005;289:8686–93.
218. Jimenez JL, Iniguez MA, Munoz-Fernández MA, Fresno M. Effect of phosphodiesterase 4 inhibitors on NFAT-dependent cyclooxygenase-2 expression in human T lymphocytes. *Cell Signal* 2004;16:1363–73.
219. Maderna P, Godson C. Phagocytosis of apoptotic cells and the resolution of inflammation. *Biochim Biophys Acta* 2003;1639:141–51.
220. Buckley CD, Pilling D, Lord JM, Akbar AN, Scheel-Toellner D, Salmon M. Fibroblasts regulate the switch from acute resolving to chronic persistent inflammation. *Trends Immunol* 2001;22:199–204.
221. Fitzpatrick FA. Inflammation, carcinogenesis and cancer. *Int Immunopharmacol* 2001;1:1651–67.