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Light-driven micro-tool equipped with a syringe function

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Abstract

Leveraging developments in microfabrication opens new possibilities for optical manipulation. With the structural design freedom from three-dimensional printing capabilities of two-photon polymerization, we are starting to see the emergence of cleverly shaped "light robots" or optically actuated micro-tools that closely resemble their macroscopic counterparts in function and sometimes even in form. In this work, we have fabricated a new type of light robot that is capable of loading and unloading cargo using photothermally induced convection currents within the body of the tool. We have demonstrated this using silica and polystyrene beads as cargo. The flow speeds of the cargo during loading and unloading are significantly larger than when using optical forces alone. This new type of light robot presents a mode of material transport that may have a significant impact on targeted drug delivery and nanofluidics injection.

Keywords: optical manipulation; two-photon fabrication; thermal convection; material transport

Introduction

It is a great challenge to build, power and control tiny machines to perform specific tasks¹. One of the earliest demonstrations of tiny machines powered by chemical fuel is the catalytic nanorods. These are composed of platinum and gold at defined zones that catalyze the decomposition of hydrogen peroxide (H_2O_2) to propel them. Nickel is added to the nanorods so that they can be guided externally by magnets. The linear speed can reach up to 20 μ m s⁻¹ depending on the concentration of the H₂O₂^{2,3}. An improvement on this design uses a rolled-up thin film of InGaAs/GaAs/Cr/Pt and can reach speeds of up to 110 µm s⁻¹. The rolled-up nanotools can be designed to perform corkscrew motions and have been demonstrated to be capable of penetrating a fixed HeLa cell⁴. 3D-printed "microfish" using the same propulsion mechanisms have been demonstrated and applied in detoxification⁵. Recently, an approach for attaching living bacterial flagella motors in a synthetic structure has been presented⁶. The purpose was to construct a bio-hybrid micro-robot that can move autonomously or be guided by chemical gradients, and the synthetic structure could be designed to accommodate drug delivery. In these examples, chemicals introduced to the medium are utilized to drive the tiny machines. Chemical "fuel" can provide high propulsion power; however, the challenge with these types of machines is the compatibility issues of the fuel with biological samples. Magnetic control requires the inclusion of nickel, which is inherently toxic to organic systems. Furthermore, magnetic and chemical gradient controls are difficult to realize for 3Dmanipulation, and fine movements are very limited^{7,8}.

Light is an attractive mechanism for powering tiny machines. Advanced control over natural or fabricated micro- or nano-structures using light has been demonstrated throughout the years, starting with the pioneering work of Ashkin⁹. Developments in pulsed lasers, light-curing polymers and complex optical trapping mechanisms all serve to catalyze the advent of more complex light-based machines and thus the emergence of so-called light robotics¹⁰. The fabrication of a microscopic bull with nano-features by Kawata *et al*¹¹ is a popular example of the precision and control that one can achieve with 3D-printing based on two-photon absorption of light sensitive polymers, with the earliest work dating back to the mid 90's¹². From then on, gradually more sophisticated micro- and nano-structures mimicking macroscopic tools have been first light-fabricated and subsequently light-actuated. A functional micro-oscillatory

system has been made and proposed as a means to investigate the mechanical properties of minute objects¹¹. There have been reports on microscopic gears^{13,14}, pumps¹⁵ and even sophisticated light foils^{16,17} that were all first fabricated and subsequently driven by light only.

The hallmark of light robotics is the use of light for fabrication, active actuation and control. One approach, akin to traditional robotics, exploits materials that can exhibit light-activated contraction to work as artificial muscles. A typical example is the recently reported microscopic walkers, which use the contraction of liquid crystal elastomers for locomotion¹⁸. Another interesting modality is made possible through the use of parallel optical trapping, whether holographic¹⁹⁻²¹ or Generalized Phase Contrast (GPC)-based²²⁻²⁴. Progress in optical manipulation has been boosted by novel technologies, such as graphics processing units and advanced spatial light modulators, which can enable the real-time calculation and generation of multi-beam trapping configurations²⁵. Advanced optical traps can be controlled independently or orchestrated to move a plurality of microscopic objects simultaneously within an imaged plane or even in an extended volume. This has been successfully demonstrated in applications for optical assembly²⁶⁻²⁸ and particle sorting²⁹⁻³¹. An early demonstration of the feasibility of light robotics is the real-time 3D manipulation of custom-fabricated micro-tools made from silica³². The micro-tools were optically translated, rotated and tilted, thus demonstrating all six-degrees-of-freedom, which is a crucial requirement for light robotics to perform delicate tasks such as surface imaging and force measurements^{33,34}. Surface imaging has been further improved with two-photon fabricated tools to the level where the achieved lateral resolution is ~200 nm, and the depth resolution is an impressive 10 nm^{35,36}. Our former work on light robotics that provides targeted-light delivery utilized two-photon fabricated waveguide structures that are optically manipulated to confine and redirect weakly focused incident light to illuminate specific targets orthogonal to the original light propagation. The nanometric feature sizes that can be achieved using two-photon fabrication tightly confine the redirected light radiating from the tip of free-floating waveguides, thus achieving targeted focusing that is stronger and tighter than what the objective lenses in the optical setup can normally provide^{37,38}. Other functionalities, such as surface-enhanced Raman spectroscopy³⁹ and fluorescence enhancement⁴⁰, have been demonstrated using metal-coated micro-tools. The flexibility in microfabrication even allows the possibility to optimize the shape of micro-tools to maximize momentum transfer^{16,41-43} or force clamping^{36,44}. Similar optimizations can be made for trapping light by using position clamping⁴⁵, efficient illumination^{46,47} or adaptive structured illumination⁴⁸.

Here, we present a new generation of light-driven micro-robots with a novel and disruptive functionality embedded inside the micro-structure. Aside from using light to impart momentum to the trapped microtool, we use light to generate and control secondary hydrodynamic effects by heating an embedded thin metal layer within the body of the tool itself. Thermal convection currents occur inside the microstructure, which, in turn, draws fluid together with potential fluid-borne cargo in or out of its body. This loading/unloading functionality and maneuverability makes it suitable as a mode of material transport that can present new and important mechanisms in nanofluidics and drug delivery. This new functionality has the advantages of optical manipulation, such as the ability to work in sealed environments such as microfluidic channels, and being able to work around optical constraints, such as refractive index contrast requirements and absorption effects in the particles being manipulated. Although thermal heating of

metal surfaces⁴⁹ and particles⁵⁰, the use of bubbles as valves⁴⁹ and fluid mixing using convection currents⁵¹ have already been demonstrated, our work is, to the best of our knowledge, the first attempt to integrate the aforementioned functionalities in a self-contained light-printed and light-driven microrobot.

Materials and methods

Fabrication of micro-tools

The fabrication process of the micro-tools is summarized in Figure 1. The shape of each micro-tool is designed to work as a tiny vessel for material transport. Cargo may be loaded and unloaded through an opening at the anterior part of the structure. Spherical handles are added to the structure for optical trapping and manipulation. The micro-tools are fabricated via two-photon direct laser writing using a commercial fabrication setup (Nanoscribe Photonic Professional, Nanoscribe GmbH, Germany). Photoresist (IP-L 780) is prepared over a microscope cover slip by simple drop casting. Normally, structures for two-photon fabrication are designed using CAD or similar 3D rendering software and the volumetric data for the structure is then sliced to identify the layers to be printed. The fabrication proceeds by scanning back and forth, layer by layer, from bottom to top until the structure is fully printed. Although this approach works well for photonic structures, such as woodpile structures, it is very inefficient for our hollow micro-tools because the laser beam will only "write" the outer shell, and there is a possibility of the structure appearing irregular due to this on-off switching of the scanning laser beam. The software for our fabrication setup accepts Cartesian coordinates to define the scanning trajectory, and we find that it is more efficient to use parametric equations to define the shape of our micro-tools⁵². The hollow body of the micro-tool is based on the surface of revolution of a so-called teardrop curve^{53,54}.

The body of the micro-tools has two openings. One serves as the spout of the micro-tool, where cargo can be loaded and ejected, and has a diameter of 6 μ m. The other opening is located on top of each micro-tool body and has a diameter of 8 μ m. The top opening enables subsequent deposition of a thin gold disk on the bottom inner wall of the micro-tool via electron beam vapor deposition. A mask with a matching hole is first fabricated to expose the target region while shielding the rest of the micro-tool during the deposition process. After two-photon exposure, the written structures are developed in a bath of isopropyl alcohol for 15 minutes. A second alcohol bath ensures that no photoresist remains inside the hollow body of each micro-tool.

The developed structures are subjected to electron beam physical vapor deposition to embed a thin metal layer inside the body of each micro-tool. First, a 1 nm layer of titanium is deposited as an adhesion layer, followed by a 5 nm layer of gold. The deposited metal layer is a circular disk with a diameter of 8 μ m corresponding to the hole on the mask and on the micro-tool. More details regarding the fabrication can be found in the accompanying Supplementary Information.

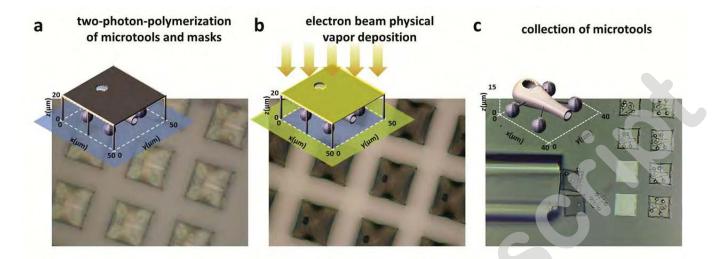


Figure 1| **Fabrication of micro-tools for material transport via enhanced light-induced thermal convection. a)** Each micro-tool is fabricated using the two-photon-polymerization process. The shape of each micro-tool is designed to function as a vessel for material transport. A mask fabricated on top of each micro-tool defines the metal-coated region on the micro-tool. b) After the micro-tools have been developed, a thin adhesion layer of titanium (1 nm) is first deposited, followed by a thin gold layer (5 nm), using electron beam physical vapor deposition. The mask shields the rest of the micro-tool and exposes only the region where the metal layer is going to be deposited **c)** Selected micro-tools are collected by carefully dislodging and then drawing them into a fine glass capillary tube. They are subsequently loaded into a cytometry cuvette for trapping experiments. Each micro-tool has a footprint of 40 µm x 40 µm.

Sample preparation

The fabricated micro-tools are anchored to the glass substrate. A fine glass capillary tube (Vitrocom, 80 μ m × 80 μ m inner cross section) fitted to a microliter syringe (Hamilton, 25 μ L gas tight syringe) is used to remove the protective mask and collect selected micro-tools. The capillary tube-syringe assembly that we made is controlled by motorized actuators (Thorlabs, 6 mm DC actuators). Another motorized actuator controls the microliter syringe plunger to load microtools inside the capillary tube and, subsequently, unload them into a cytometry cuvette (Hellma, 250 μ m × 250 μ m inner channel cross section). The microtools are transferred into a cytometry cuvette containing a solution of deionized water, 0.5% Tween 80 surfactant and 10% ethanol. The process is performed under a microscope, and the transfer efficiency can be as high as 100% due to the selective and interactive picking approach. Video micro-tools remain safely anchored to the substrate for succeeding experiments. Figure 1c shows that the masks have successfully shielded the micro-tools based on the bright square regions on the substrate (unexposed area) where the mask and microtool used to stand. Similar image analysis confirms that a circular disk was deposited inside the microtool.

Optical manipulation and light-induced thermal convection

Optical trapping and manipulation experiments are performed on our BioPhotonics Workstation (see Fig. 2). The BioPhotonics Workstation uses multiple dual-beam optical traps, relayed to the sample through two opposing microscope objectives (Olympus LMPLN 50x IR, WD = 6.0 mm, NA = 0.55), from multiple synchronized beams generated by a proprietary, computer-controlled illumination module. This illumination module uses a spatial light modulator to generate multiple beams out of a CW near-infrared (IPG Photonics, $\lambda = 1070$ nm, 40 W max power). Up to 100 independently controllable dual beam traps can currently be generated. Each dual beam trap consists of two co-axial, counter-propagating beams. A trapped particle can be laterally manipulated by synchronously shifting the counter-propagating beams, whereas axial motion is controlled by changing the intensity ratio of these two beams. We have shown in an earlier work how this optical trapping modality is able to manipulate a plurality of particles in 3D space⁵⁵. In contrast to single beam optical tweezers, which use gradient forces from a tightly focused beam, counter-propagating beams use scattering forces and do not require tight focusing. This means that low numerical aperture objective lenses can be used in our workstation. Low numerical aperture objective lenses and can thus accommodate bigger samples, such as our cytometry cuvette and side imaging³⁷, if needed.

During experiments, a user traps and steers each micro-tool in three dimensions through an interactive computer interface, where an operator can select, trap, move and reorient them in real-time. With the aid of a LabVIEW-based graphical user interface, lateral manipulation is performed by simply dragging the traps, shown as overlay graphics over real-time images acquired from the microscope, whereas axial manipulation is performed by sliding a graphical control object.

For the optical manipulation of our micro-tools, we use four pairs of counter-propagating beams located at the four spherical handles. We can move the micro-tools at approximately 10 μ m s⁻¹ and rotate them at 17° s⁻¹. The BioPhotonics Workstation also generates an additional trapping beam, which is "repurposed" to illuminate the thin metal layer inside each structure, which then serves as a light-activated heating element for the fluid inside the micro-tool. This additional beam controls the loading and unloading, which we will discuss in the next section. Convection becomes observable when the laser power at the sample is approximately 17 mW. Videos of the experiments are obtained from the top-view (e.g., see Supplementary materials), and selected snapshots are presented in the manuscript.

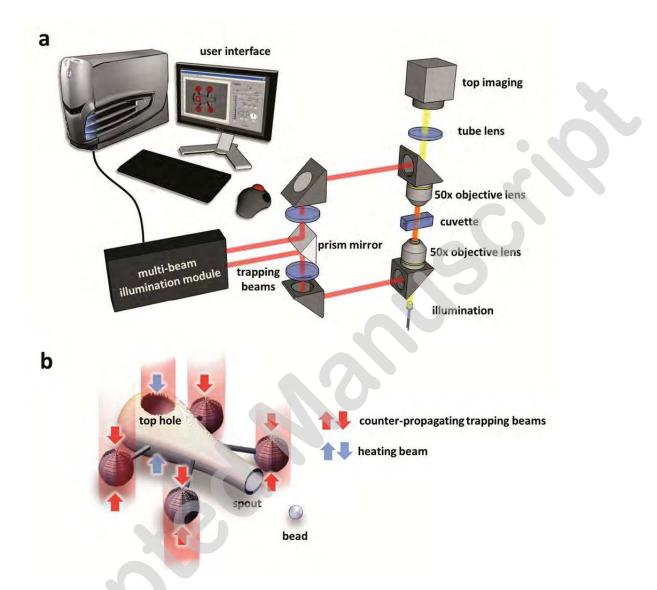


Figure 2 Schematic showing the BioPhotonics Workstation for optical trapping, manipulation and actuation of a micro-tool. a) The BioPhotonics Workstation generates counter-propagating beam traps using a multi-beam illumination module. The top and bottom set of counter-propagating beams are imaged in the cuvette through opposing 50x objective lenses. The top imaging is fed real-time to the user-interface for intuitive optical manipulation. b) Once loaded in the cuvette, optical manipulation of the micro-tools is done using real-time configured counter-propagating beams for each sphere handle. The use of multiple trapping beams allows tool movements with full six-degrees-of-freedom actuation and is controlled using a LabVIEW-based user interface. An extra beam aimed at the micro-tool's top hole is used for heating the thin metallic layer.

Results and discussion

Each of our micro-tools was designed to function as a vessel that can be moved with optical traps in realtime. Proof-of-principle experiments demonstrate that each of them can be used to load and unload cargo

using laser-induced thermal convection. When the thin metal layer is heated by one of the available trapping beams, we observe that the heat generated is enough to create a microbubble and produce strong convection currents that can pull silica ($\emptyset = 2 \ \mu m$) and polystyrene ($\emptyset = 1 \ \mu m$) beads toward the spout and into the tool. We show in Fig. 3 an illustrative flow speed measurement for a silica bead, starting from outside the micro-tool until it enters the body. We observed flow speeds of approximately 10 $\mu m \ s^{-1}$ near the opening, which decrease as it moves toward the wider cross section, in accordance with the continuity equation. Near the heating element, the flow speed reaches more than 25 $\mu m \ s^{-1}$. This is greater than previously reported flow speeds generated by two-photon fabricated rotors acting as micropumps¹⁵.

Thermal convection due to photothermal heating involves both photonic and fluidic phenomena. Multiphysics computer modeling of plasmonic heating elements having dimensions of less than 200 nm predicts flow speeds of approximately 10 nm s⁻¹, and it has been suggested that heating elements should be greater than 1 μ m for microfluidic applications⁵⁶ (we used an 8 μ m diameter). The convection current that draws particles into our micro-tools can be the combined result of natural and Marangoni convection, but it can also create a surface tension gradient along a microbubble surface. The surface tension gradient due to the temperature difference between the top and bottom surfaces of a bubble leads to Marangoni convection, which can be very strong⁵⁸. Once the particle touches the microbubble, surface tension force essentially traps the particle and thus prevents it from coming out. A study of particle assembly on a sandwiched colloidal suspension using Marangoni convection reports a maximum flow velocity at the gas/liquid interface as high as ~0.3 m s⁻¹. Away from the bubble, there is a significant decrease in the velocity⁵⁷. The trend in our flow speed measurement of a tracer particle while it is being dragged by the convective flow (Fig. 3) is consistent with this observation. An increase of 1-2 orders of magnitude of the mass transfer has also been observed for dissolved molecules⁵⁹.

Photothermal particles and thin metal films have been previously used in microfluidics for heat-induced flow control, sorting and mixing. However, the precise spatial control of particles' motion and locations, such as placing them where and when they are needed, can be challenging for smaller particles⁶⁰. Moreover, thin metal films deposited on fixed regions within microfluidic channels completely lack the maneuverability that we show here. In short, our approach demonstrates a solution to this challenge by integrating a thin metal film within each light-controlled micro-tool that can readily function to transport cargo.

In our experiments, laser-induced heating of the metallic layer is able to form a microbubble inside the body of each micro-tool. Such microbubble formation is known to occur at temperatures between 220 and 240 °C for an array of nanoparticles and is more or less invariant with the size of the illuminated area and incident laser power⁶¹. Moreover, we have observed that some of the polystyrene beads captured by a micro-tool can be melted by continuously heating the fluid. No damage has been observed on either micro-tools or metal layers due to the photothermal heating.

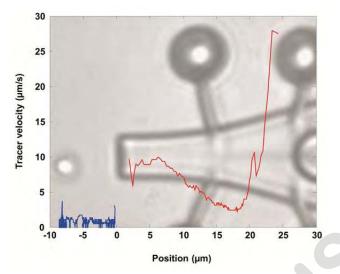


Figure 3| **Flow speed measurement.** The thin metal layer inside the body of each micro-tool is heated using a laser beam (1070 nm), which, in turn, creates a microbubble and generates strong thermal convection currents that gradually draw the cargo toward the spout of the micro-tool. We use a feature tracking algorithm to monitor the movement of the beads. The zero position is set at the spout of the micro-tool. The clear gap observed in the velocity plot is due to the limitation of the tracking algorithm to identify the bead when it crosses the dark outline of the micro-tool. The blue and red plots represent the velocity of the bead while it is outside and inside of the micro-tool, respectively. An image of a micro-tool is added to give a scale-indication of the horizontal axis. See Supplementary material.

Continuous illumination generates strong thermal convection currents that pull surrounding particles toward a laser-induced microbubble. Others have shown the feasibility of exploiting this for material transport, e.g., direct-writing of patterned particle assemblies. It has been reported that dragging a microbubble with a heating CW laser beam can result in the collection and deposition of particles along its path to accomplish direct-writing of patterned particle assemblies⁶². Our tool utilizes the same principle, with the advantage of greater selectivity and control over the particle collection because the design of the tool limits the direction of convective flow. In Fig. 4, we demonstrate spatial control of the micro-tool by picking up scattered silica beads. The particle velocities we have measured for our micro-tool are significantly larger than when using an optical trap alone. We have therefore exploited the conversion of optical energy to heat and, consequently, to kinetic energy via hydrodynamic effects to realize a new light-based micro-tool capable of performing controlled mechanical interactions with its surrounding micro-world in a way that goes beyond the limitations of conventional optical trapping. Whereas conventional optical trapping requires sufficient refractive index contrast between the captured particles and their surrounding medium, our method does not suffer from this inherent limitation. Potentially unwanted radiation effects to the sample can also be minimized or prevented.

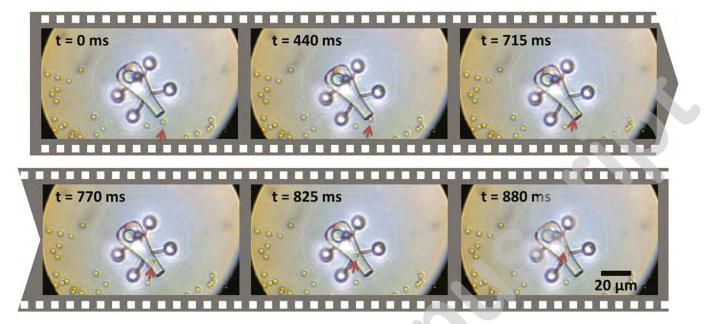


Figure 4 | **Loading of cargo inside the micro-tool.** Due to the spatial control provided by optical manipulation, the light robot can pick up cargo at different locations. See Supplementary material.

Upon loading cargo inside the tool and conveniently moving the tool to another location via optical micromanipulation, our experiments show that we can also exploit light-induced processes to eject captured particles. Figure 5 shows experiment results demonstrating that the cargo can be ejected by slightly moving the heating beam across the body of the micro-tool to perturb the microbubble to pump fluids with the particles out of the structure. This functionality mimics the familiar action of pumping a syringe. It has been observed that bubbles get attracted to regions of higher temperature, a phenomenon called thermocapillary bubble migration⁵⁸. The attraction of a bubble to a heat source is very strong (i.e., up to an order of magnitude stronger than optical forces); thus, it is feasible to use thermocapillary bubble migration as a control for pumping. Trapping of bubbles has also been observed in more viscous molten glass media, where deformation of the bubble is the proposed trapping mechanism⁶³. Some simulations have shown that the Marangoni convection can reverse when there are many particles adhering to the bubble⁵⁷. This reversal of the Marangoni convection may also be present during the unloading of cargo inside our micro-tools. Most studies have been done on unconstrained Marangoni convection, where there is a thin fluid film, and the boundary is only at the bottom or both top and bottom surfaces. For our microtool, the fluid is practically constrained in all directions, allowing only a small opening for the cargo and for the continuity of fluid flow to hold (i.e., top hole). Thus, we expect a nontrivial flow phenomenon that warrants further investigation. At this point, it suffices to say that the pumping action of the micro-tool is light-activated and, to our knowledge, this functionality has never been demonstrated before in a lightactuated micro-tool. Future work will explore new structure designs to optimize control over the convection processes constrained within them and even avoid ejecting particles out through the top hole, which occasionally happens in the current design (e.g., covering the top hole with mesh-like features after metal deposition).

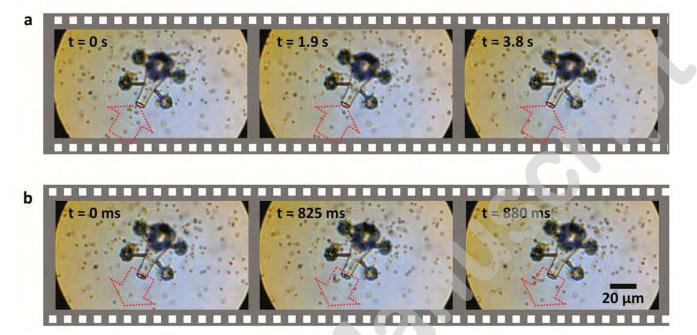


Figure 5| **The micro-tool can be used as a tiny pump. a)** A relatively large number of polystyrene beads (1 µm diameter) are dispersed in the trapping medium, and the micro-tool is used to collect them. **b)** By changing the location of the heating beam, the micro-tool can be used to eject the captured particles by using the microbubble as a light-controlled piston. See Supplementary material.

Conclusion and outlook

Various methods and technologies can be integrated to build and control optically-actuated micro-tools or light robots that can perform specific tasks. Microfluidics, plasmonics, optical manipulation and fabrication have already found successful applications in their respective areas. However, combining them presents not only new challenges but also new and exciting ways to enable disruptive functionalities that would otherwise be difficult to realize by each sub-discipline in isolation. In this work, we have presented an integration of optical manipulation, two-photon fabrication and metal deposition to create a new category in the toolbox of light robotics. We have embedded thin metal layers inside a plurality of lightdriven micro-tools that enable the conversion of incident optical energy into heat and eventually hydrodynamic effects. Heating the metal layers generates thermal convection currents that can be used to load and unload cargo. We have demonstrated that light-controlled pumping makes each micro-tool suitable for material transport. A potential application that can fully utilize the capability of our new micro-tools is drug delivery. Micro-machined devices with modified surface chemistry and morphology have already been successfully used in drug delivery⁶⁴. Such devices are fabricated using a standard lithography process and have a planar geometry. However, using optically manipulated micro-sources can provide much better spatial and temporal selectivity, as shown by experiments on cell stimulation via chemotaxis⁶⁵. Such examples motivate the idea of a structure-mediated approach in biological studies that use light-controlled, steered and actuated microstructures to mediate access to the sub-micron domain.

Light robotics is thus an excellent candidate for realizing these new functionalities in a fully flexible and dynamic context. The structural design freedom in two-photon fabrication can even adopt micro-needle structures, which are commonly used for transdermal drug delivery⁶⁶, as an approach to advanced intracellular drug delivery. There has been interest in understanding diseases such as circulating tumor cells, which are very rare in blood samples. Because of their rarity, bulk measurement will average out the unique signature of this type of cell⁶⁷. Our micro-tools can work in plurality and even with other micro-tools of different functionality to probe this single cell to investigate cellular responses to spatially or temporally correlated mechanical or chemical stimulation.

Because the operation of our first batch of novel, internally functionalized micro-tools is based on photothermal heating, resonant plasmonic structures can be readily integrated and used in future light robotic tools for more efficient heating and wavelength selectivity. The metal coating can be added on the spout of the micro-tool and can also be heated up once in contact with a cell of interest. It has been proposed in transfection experiments that heating of the cell membrane induces phase changes in the lipid layer and thus allows the entry of foreign material. Our micro-tools do not preclude the possibility of being loaded prior to introduction to the trapping medium; thus, it is also possible to have micro-tools with different chemicals that can perform precise chemical stimulation that is not possible in a standard cell culture. We envision these micro-tools to be an important addition to the current tools for understanding biology at the micro-scale.

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Supplementary information accompanies the manuscript on the Light: Science & Applications website (http://www.nature.com/lsa/)

References

- 1 Ozin GA, Manners I, Fournier-Bidoz S, Arsenault A. Dream nanomachines. *Adv Mater* 2005; **17**: 3011-3018.
- 2 Paxton WF, Kistler KC, Olmeda CC, Sen A, St Angelo SK *et al.* Catalytic nanomotors: Autonomous movement of striped nanorods. *J Am Chem Soc* 2004; **126**: 13424-13431.
- Paxton WF, Sundararajan S, Mallouk TE, Sen A. Chemical locomotion. *Angew Chemie Int Ed* 2006;
 45: 5420-5429.
- 4 Solovev AA, Xi W, Gracias DH, Harazim SM, Deneke C *et al.* Self-propelled nanotools. *ACS Nano* 2012; **6**: 1751-1756.
- 5 Zhu W, Li JX, Leong YJ, Rozen I, Qu X *et al.* 3D-printed artificial microfish. *Adv Mater* 2015; **27**: 4411-4417.

- 6 Barroso Á, Landwerth S, Woerdemann M, Alpmann C, Buscher T *et al.* Optical assembly of biohybrid micro-robots. *Biomed Microdevices* 2015; **17**: 26.
- 7 Campuzano S, Kagan D, Orozco J, Wang J. Motion-driven sensing and biosensing using electrochemically propelled nanomotors. *Analyst* 2011; **136**: 4621-4630.
- 8 Kim K, Guo JH, Xu XB, Fan DL. Recent progress on man-made inorganic nanomachines. *Small* 2015; **11**: 4037-4057.
- 9 Ashkin A, Dziedzic JM, Bjorkholm JE, Chu S. Observation of a single-beam gradient force optical trap for dielectric particles. *Opt Lett* 1986; **11**: 288.
- 10 Palima D, Glückstad J. Gearing up for optical microrobotics: micromanipulation and actuation of synthetic microstructures by optical forces. *Laser Photon Rev* 2013; 7: 478-494.
- Kawata S, Sun HB, Tanaka T, Takada K. Finer features for functional microdevices. *Nature* 2001;
 412: 697-698.
- 12 Maruo S, Nakamura O, Kawata S. Three-dimensional microfabrication with two-photon-absorbed photopolymerization. *Opt Lett* 1997; **22**: 132-134.
- Galajda P, Ormos P. Complex micromachines produced and driven by light. *Appl Phys Lett* 2001; 78: 249-251.
- 14 Kelemen L, Valkai S, Ormos P. Integrated optical motor. *Appl Opt* 2006; **45**: 2777-2780.
- 15 Maruo S, Inoue H. Optically driven micropump produced by three-dimensional two-photon microfabrication. *Appl Phys Lett* 2006; **89**: 144101.
- 16 Swartzlander GA, Peterson TJ, Artusio-Glimpse AB, Raisanen AD. Stable optical lift. *Nat Photonics* 2011; **5**: 48-51.
- 17 Glückstad J. Optical manipulation: Sculpting the object. *Nat Photonics* 2011; **5**: 7-8.
- 18 Zeng H, Wasylczyk P, Parmeggiani C, Martella D, Burresi M *et al.* Light-fueled microscopic walkers. *Adv Mater* 2015 ; **27**: 3883-3887; doi:10.1002/adma.201501446.
- 19 Liesener J, Reicherter M, Haist T, Tiziani HJ. Multi-functional optical tweezers using computergenerated holograms. *Opt Commun* 2000; **185**: 77-82.
- 20 Dufresne ER, Spalding GC, Dearing MT, Sheets SA, Grier DG. Computer-generated holographic optical tweezer arrays. *Rev Sci Instrum* 2001; **72**: 1810-1816.

- 21 Curtis JE, Koss BA, Grier DG. Dynamic holographic optical tweezers. *Opt Commun* 2002; **207**: 169-175.
- 22 Eriksen RL, Daria VR, Glückstad J. Fully dynamic multiple-beam optical tweezers. *Opt Express* 2002; **10**: 597-602.
- 23 Rodrigo PJ, Daria VR, Glückstad J. Four-dimensional optical manipulation of colloidal particles. *Appl Phys Lett* 2005; 86: 074103.
- Ulriksen HU, Thøgersen J, Keiding S, Perch-Nielsen I, Dam J *et al.* Independent trapping,
 manipulation and characterization by an all-optical biophotonics workstation. *J Eur Opt Soc Rapid Publ* 2008; **3**: 08034.
- 25 Persson M, Engström D, Goksör M. Real-time generation of fully optimized holograms for optical trapping applications. In: Dholakia K, Spalding GC, editors. Proceedings of SPIE Optical Trapping and Optical Micromanipulation VIII. SPIE: San Diego, CA, USA, 2011; 8097: 80971H.
- 26 Chapin SC, Germain V, Dufresne ER. Automated trapping, assembly, and sorting with holographic optical tweezers. *Opt Express* 2006; **14**: 13095-13100.
- 27 Rodrigo PJ, Kelemen L, Alonzo CA, Perch-Nielsen IR, Dam JS *et al.* 2D optical manipulation and assembly of shape-complementary planar microstructures. *Opt Express* 2007; **15**: 9009-9014.
- 28 Rodrigo PJ, Kelemen L, Palima D, Alonzo CA, Ormos P *et al.* Optical microassembly platform for constructing reconfigurable microenvironments for biomedical studies. *Opt Express* 2009; **17**: 6578-6583.
- 29 Buican TN, Smyth MJ, Crissman HA, Salzman GC, Stewart CC *et al.* Automated single-cell manipulation and sorting by light trapping. *Appl Opt* 1987; 26: 5311-5316.
- 30 Wang MM, Tu E, Raymond DE, Yang JM, Zhang HC *et al.* Microfluidic sorting of mammalian cells by optical force switching. *Nat Biotechnol* 2005; 23: 83-87.
- 31 Perch-Nielsen I, Palima D, Dam JS, Glückstad J. Parallel particle identification and separation for active optical sorting. *J Opt A Pure Appl Opt* 2009; **11**: 34013-34018.
- 32 Rodrigo PJ, Gammelgaard L, Bøggild P, Perch-Nielsen IR, Glückstad J. Actuation of microfabricated tools using multiple GPC-based counterpropagating-beam traps. *Opt Express* 2005; **13**: 6899-6904.
- 33 Phillips DB, Grieve JA, Olof SN, Kocher SJ, Bowman R *et al.* Surface imaging using holographic optical tweezers. *Nanotechnology* 2011; **22**: 285503.

- 34 Ikin L, Carberry DM, Gibson GM, Padgett MJ, Miles MJ. Assembly and force measurement with SPM-like probes in holographic optical tweezers. *New J Phys* 2009; **11**: 023012.
- 35 Phillips DB, Gibson GM, Bowman R, Padgett MJ, Hanna S *et al.* An optically actuated surface scanning probe. *Opt Express* 2012; **20**: 29679-29693.
- 36 Phillips DB, Padgett MJ, Hanna S, Ho YLD, Carberry DM *et al.* Shape-induced force fields in optical trapping. *Nat Photonics* 2014; 8: 400-405.
- 37 Palima D, Bañas AR, Vizsnyiczai G, Kelemen L, Ormos P *et al.* Wave-guided optical waveguides. Opt Express 2012; 20: 2004-2014.
- 38 Villangca M, Bañas A, Palima D, Glückstad J. Dynamic diffraction-limited light-coupling of 3Dmaneuvered wave-guided optical waveguides. *Opt Express* 2014; 22: 17880-17889.
- Vizsnyiczai G, Lestyán T, Joniova J, Aekbote BL, Strejčková A *et al.* Optically trapped surfaceenhanced Raman probes prepared by silver photo-reduction to 3D microstructures. *Langmuir* 2015;
 31: 10087-10093; doi:10.1021/acs.langmuir.5b01210.
- 40 Aekbote BL, Schubert F, Ormos P, Kelemen L. Gold nanoparticle-mediated fluorescence enhancement by two-photon polymerized 3D microstructures. *Opt Mater* 2014; **38**: 301-309.
- 41 Metzger NK, Mazilu M, Kelemen L, Ormos P, Dholakia K. Observation and simulation of an optically driven micromotor. *J Opt* 2011; **13**: 044018.
- 42 Palima D, Bañas AR, Vizsnyiczai G, Kelemen L, Aabo T *et al.* Optical forces through guided light deflections. *Opt Express* 2013; **21**: 581-593.
- 43 Neale SL, MacDonald MP, Dholakia K, Krauss TF. All-optical control of microfluidic components using form birefringence. *Nat Mater* 2005; **4**: 530-533.
- 44 Simpson SH, Phillips DB, Carberry DM, Hanna S. Bespoke optical springs and passive force clamps from shaped dielectric particles. *J Quant Spectrosc Radiat Transf* 2013; **126**: 91-98.
- 45 Preece D, Bowman R, Linnenberger A, Gibson G, Serati S *et al.* Increasing trap stiffness with position clamping in holographic optical tweezers. *Opt Express* 2009; **17**: 22718-22725.
- 46 Bañas A, Kopylov O, Villangca M, Palima D, Glückstad J. GPC light shaper: static and dynamic experimental demonstrations. *Opt Express* 2014; **22**: 23759-23769.
- 47 Villangca M, Bañas A, Palima D, Glückstad J. GPC-enhanced read-out of holograms. *Opt Commun* 2015; **351**: 121-127.

- 48 Taylor MA, Waleed M, Stilgoe AB, Rubinsztein-Dunlop H, Bowen WP. Enhanced optical trapping via structured scattering. *Nat Photonics* 2015; **9**: 669-674.
- 49 Zhang K, Jian AQ, Zhang XM, Wang Y, Li ZH *et al.* Laser-induced thermal bubbles for microfluidic applications. *Lab Chip* 2011; **11**: 1389-1395.
- Liu GL, Kim J, Lu Y, Lee LP. Optofluidic control using photothermal nanoparticles. *Nat Mater* 2006;
 5: 27-32.
- 51 Miao XY, Wilson BK, Lin LY. Localized surface plasmon assisted microfluidic mixing. *Appl Phys Lett* 2008; **92**: 124108.
- 52 Villangca M, Bañas A, Palima D, Glückstad J. Generalized phase contrast-enhanced diffractive coupling to light-driven microtools. *Opt Eng* 2015; **54**: 111308.
- 53 Weisstein EW. Teardrop curve. Mathematica Notebook. [updated 22 March 2016; cited 29 March 2016]. Available from: http://mathworld.wolfram.com/TeardropCurve.html
- 54 Weisstein EW. Surface of revolution. Mathematica Notebook. [updated 22 March 2016; cited 29 March 2016]. Available from: http://mathworld.wolfram.com/SurfaceofRevolution.html
- 55 Rodrigo PJ, Perch-Nielsen IR, Alonzo CA, Glückstad J. GPC-based optical micromanipulation in 3D real-time using a single spatial light modulator. *Opt Express* 2006; **14**: 13107-13112.
- 56 Donner JS, Baffou G, McCloskey D, Quidant R. Plasmon-assisted optofluidics. *ACS Nano* 2011; **5**: 5457-5462.
- 57 Lin LH, Peng XL, Mao ZM, Li W, Yogeesh MN *et al.* Bubble-pen lithography. *Nano Lett* 2016; **16**: 701-708; doi:10.1021/acs.nanolett.5b04524.
- 58 Berry DW, Heckenberg NR, Rubinszteindunlop H. Effects associated with bubble formation in optical trapping. *J Mod Opt* 2000; **47**: 1575-1585.
- 59 Louchev OA, Juodkazis S, Murazawa N, Wada S, Misawa H. Coupled laser molecular trapping, cluster assembly, and deposition fed by laser-induced Marangoni convection. *Opt Express* 2008; 16: 5673-5680.
- 60 Kayani AA, Khoshmanesh K, Ward SA, Mitchell A, Kalantar-zadeh K. Optofluidics incorporating actively controlled micro- and nano-particles. *Biomicrofluidics* 2012; **6**: 031501.
- 61 Baffou G, Polleux J, Rigneault H, Monneret S. Super-heating and micro-bubble generation around plasmonic nanoparticles under cw illumination. *J Phys Chem C* 2014; **118**: 4890-4898.

- 62 Zheng YJ, Liu H, Wang Y, Zhu C, Wang SM *et al.* Accumulating microparticles and direct-writing micropatterns using a continuous-wave laser-induced vapor bubble. *Lab Chip* 2011; **11**: 3816-3820.
- 63 Murazawa N, Juodkazis S, Misawa H, Wakatsuki H. Laser trapping of deformable objects. *Opt Express* 2007; **15**: 13310-13317.
- 64 Tao SL, Desai TA. Micromachined devices: The impact of controlled geometry from cell-targeting to bioavailability. *J Control Release* 2005; **109**: 127-138.
- 65 Kress H, Park JG, Mejean CO, Forster JD, Park J *et al.* Cell stimulation with optically manipulated microsources. *Nat Methods* 2009; **6**: 905-909.
- 66 Gittard SD, Ovsianikov A, Chichkov BN, Doraiswamy A, Narayan RJ. Two-photon polymerization of microneedles for transdermal drug delivery. *Expert Opin Drug Deliv* 2010; **7**: 513-533.
- 67 Villangca M, Casey D, Glückstad J. Optically-controlled platforms for transfection and single-and sub-cellular surgery. *Biophys Rev* 2015; **7**: 379-390.

