



Review

Melatonin Can Modulate Neurodegenerative Diseases by Regulating Endoplasmic Reticulum Stress

Yeong-Min Yoo ^{1,*} and Seong Soo Joo ^{2,*}

¹ East Coast Life Sciences Institute, College of Life Science, Gangneung-Wonju National University, Gangneung 25457, Republic of Korea

² Department of Marine Bioscience, College of Life Science, Gangneung-Wonju National University, Gangneung 25457, Republic of Korea

* Correspondence: yooym@gwnu.ac.kr (Y.-M.Y.); ssj66@gwnu.ac.kr (S.S.J.); Tel.: +82-10-2494-5309 (Y.-M.Y.); +82-33-640-2856 (S.S.J.); Fax: +82-33-640-2849 (Y.-M.Y. & S.S.J.)

Abstract: As people age, their risks of developing degenerative diseases such as cancer, diabetes, Parkinson's Disease (PD), Alzheimer's Disease (AD), rheumatoid arthritis, and osteoporosis are generally increasing. Millions of people worldwide suffer from these diseases as they age. In most countries, neurodegenerative diseases are generally recognized as the number one cause afflicting the elderly. Endoplasmic reticulum (ER) stress has been suggested to be associated with some human neurological diseases, such as PD and AD. Melatonin, a neuroendocrine hormone mainly synthesized in the pineal gland, is involved in pleiotropically biological functions, including the control of the circadian rhythm, immune enhancement, and antioxidant, anti-aging, and anti-tumor effects. Although there are many papers on the prevention or suppression of diseases by melatonin, there are very few papers about the effects of melatonin on ER stress in neurons and neurodegenerative diseases. This paper aims to summarize and present the effects of melatonin reported so far, focusing on its effects on neurons and neurodegenerative diseases related to ER stress. Studies have shown that the primary target molecule of ER stress for melatonin is CHOP, and PERK and GRP78/BiP are the secondary target molecules. Therefore, melatonin is crucial in protecting neurons and treating neurodegeneration against ER stress.

Keywords: melatonin; neurons; neurodegenerative diseases; endoplasmic reticulum stress



Citation: Yoo, Y.-M.; Joo, S.S. Melatonin Can Modulate Neurodegenerative Diseases by Regulating Endoplasmic Reticulum Stress. *Int. J. Mol. Sci.* **2023**, *24*, 2381. <https://doi.org/10.3390/ijms24032381>

Academic Editors: Beatriz Caballero and Yaiza Potes

Received: 26 December 2022

Revised: 17 January 2023

Accepted: 21 January 2023

Published: 25 January 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The endoplasmic reticulum (ER) is critical for normal cellular function. ER stress due to dysfunction or loss of integrity caused by the accumulation of unfolded proteins or changes in calcium homeostasis within the ER can lead to apoptosis and cell signals associated with apoptosis [1–3]. Common causes of neurodegenerative diseases include the accumulation and deposition of misfolded proteins that affect neuronal connectivity, cell death, and faulty cell signaling [4,5]. Unfolded protein response (UPR) activity, typically dysfunctional due to erroneous protein aggregation or oxidative stress in cells, can cause more protein accumulation in the cells, leading to ER stress and disease exacerbation [6,7]. Melatonin, a neuroendocrine hormone mainly synthesized in the pineal gland, is involved in pleiotropically biological functions, including the control of the circadian rhythm, immune enhancement, antioxidant, anti-aging, and anti-tumor effects [8,9].

This paper aimed to collect and analyze data on the association between neurodegenerative diseases and ER stress in PubMed published up to 23 November 2022, focusing on molecular mechanisms involved in such association according to the year of publication and how melatonin might contribute to the relationship between ER stress and human neurological disorders.

2. Relationship between Endoplasmic Reticulum Stress and Neurodegenerative Diseases

As of 23 November 2022, the total number of published articles related to neurodegenerative diseases in PubMed was 400,813, of which 17,869 included oxidative stress among the cell mechanisms related to neurodegenerative diseases. There were 13,097 articles on apoptosis, 11,477 on mitochondria, 6481 on autophagy, and 1627 on ER stress. Among the neurodegenerative disease studies, ER stress-related research started in 1990 had fewer studies than the other studies. According to PubMed, there were 514 review articles. In 2005, there were only about 20 papers. However, the research has been quite explosive recently (Figure 1).

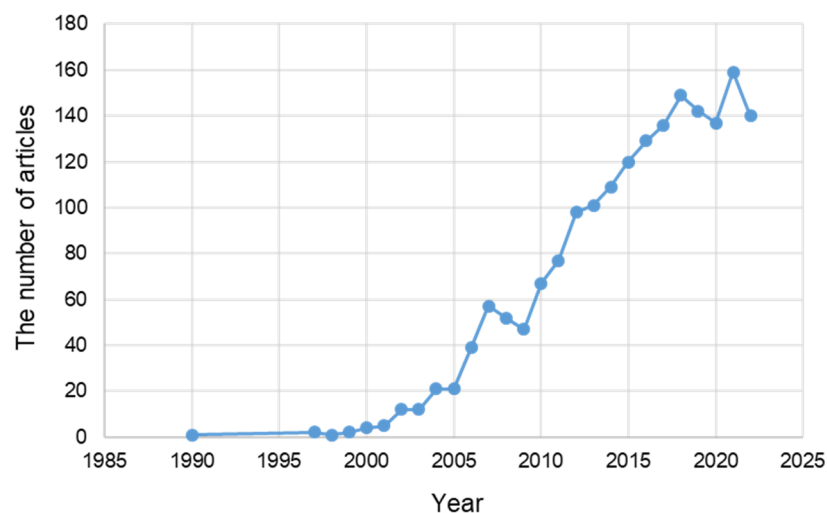


Figure 1. The number of articles related to ER stress among articles on neurodegenerative diseases published in PubMed from 1990 to 23 November 2022.

According to PubMed, Parkinson's Disease (PD) was the most studied neurodegenerative disease, with a focus on ER stress, followed by Alzheimer's Disease (AD), Amyotrophic Lateral Sclerosis (ALS), Transmissible Spongiform Encephalopathy (TSE), polyglutamine diseases, acute neurodegeneration, and neuronal storage diseases (Figure 2). This suggests that PD, AD, and ALS are the most relevant neurodegenerative diseases in the elderly. It also suggests that the research has been focused on ER stress for treating these diseases.

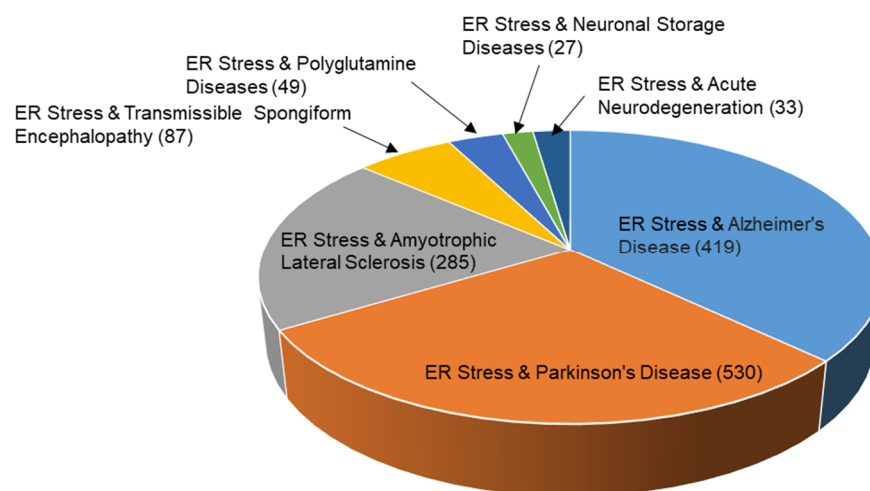


Figure 2. The number of articles related to neurodegenerative diseases and ER stress investigated by PubMed by 23 November 2022.

In the following, we will describe how ER stress in neurodegenerative diseases may contribute to the disease process of human neurological disorders.

2.1. ER Stress and Parkinson's Disease (PD)

PD is an age-related, chronic, progressive, degenerative disease caused by the decline of dopaminergic neurons in a specific brain area, i.e., the substantia nigra pars compacta. Typically, PD is characterized by intraneuronal cytoplasmic inclusion bodies, known as Lewy bodies, caused by the abnormal deposition of a protein called alpha-synuclein in the brain [10]. However, the exact cellular mechanisms that cause selective neuronal death in PD are still not fully understood yet [11].

According to PubMed, research on ER stress in PD began in 2000 and gradually increased. Since 2016, more than 46 papers have been intensively published every year (Figure 3).

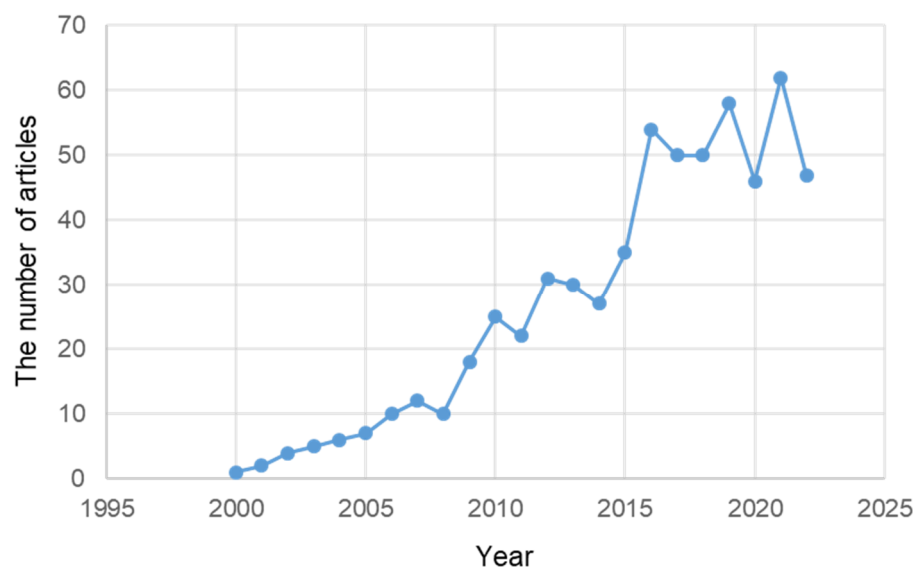


Figure 3. The number of articles related to ER stress in PD published in PubMed from 2000 to 23 November 2022.

The following papers are organized by year of publication in PubMed: 6-Hydroxydopamine (6-OHDA) and 1-methyl-4-phenyl-pyridinium (MPP+) are widely used to induce dopaminergic neuron death in vitro and in vivo PD models. An association between ER stress and the UPR has been reported in neuronal PC12 cells [12], sympathetic neurons from PERK null mice [13], and the MN9D cells' dopaminergic cell line [13]. ER-localized molecular chaperones (GRP78/BiP and GRP94) have been revealed in SH-SY5Y cells [14]. The relationship between Parkin protein and ER stress in PD has also been proven [15]. IRE1alpha, an ER stress molecule, participates in unfolded protein responses through its interaction with the Jun activation domain-binding protein-1 (JAB1) [16]. Alpha-synuclein cytotoxicity is related to ER stress in PD [17–21]. The ER stress molecules CHOP/Gadd153 and GRP78/BiP are increased in human neuroblastoma SH-SY5Y cells treated with 6-OHDA, indicating that ER dysfunction is involved in the mechanisms induced by 6-OHDA in SH-SY5Y cells [22]. The relationship between alpha-synuclein and PD concerning ER stress has been reviewed [23–25]. As ER stress markers, the phosphorylated pancreatic ER kinase (p-PERK) and phosphorylated eukaryotic initiation factor 2alpha (p-eIF2alpha) have been detected in the substantia nigra of PD patients, suggesting that the UPR's activation is closely related to the accumulation of alpha-synuclein aggregates [26]. The emerging mechanisms of ER stress in PD have been reviewed [27,28] and demonstrated [29].

Parkin has a role in the crosstalk between ER and mitochondrial stress, suggesting that Parkin has a role in the crosstalk between ER and mitochondrial stress, suggesting that both ER and mitochondrial stress can contribute to the pathogenesis of PD [30]. GRP78/BiP can bind to α -synuclein and increase the aggregated α -synuclein's accumulation in in vitro and in vivo models, indicating that the activation of the UPR pathway in the PD brain is associated with α -synuclein's accumulation occurring, in part, within the ER [31]. Cali et al. [32] have reviewed the interplay between the mitochondria, ER, and proteasome

systems in PD-associated neurodegeneration. When molecular events occurring during ER stress and the unfolded protein response are evaluated, it has been found that the ER stress response plays a role in neurodegenerative disorders, including AD, PD, ALS, and prion diseases [33–37]. Alpha-synuclein can inhibit ATF6, a protective branch of the UPR, suggesting a link between ER stress and the role of the UPR in PD [38]. Tsujii et al. have reviewed the involvement of ER stress with in vitro and in vivo PD models [39]. GRP78/BiP and p-PERK are increased in the Lewy bodies of patients with dementia [40]. An increase in activating transcription factor 4 (ATF4) as a member of the PERK signaling pathway has been identified in experimental animal models and the postmortem melanin-containing neurons of PD patients [41,42]. The IRE1 pathway can drive PD's progression by coupling the ER stress [43]. The significant role of ER stress in PD pathogenesis has been highlighted [44]. eIF2 α phosphorylation mediated by PERK can protect DA neurons against chronic heat stress in *Drosophila* [45]. The neurodegenerative disease can be activated or inhibited through the PERK pathway [46].

Therefore, the target molecules of ER stress for PD are GRP78/BiP, CHOP, PERK, IRE1, ATF6, and ATF4, suggesting that these ER stress molecules can act mainly as pathological factors in PD.

2.2. ER Stress and Alzheimer's Disease (AD)

AD is a neurodegenerative disease that can lead to progressive impairments in memory, behavioral, and thinking functions. AD causes the loss of neurons in several brain regions within the prefrontal cortex, hippocampus, and basal forebrain area [47]. Pathological features of AD patients' brain tissues include extracellular senile plaques with amyloid β -peptide ($A\beta$) deposits and intracellular accumulation of the tau protein with neurofibrillary tangles (NFTs). AD includes early-onset familial AD (FAD) and later-onset sporadic AD. Mutations in presenilin PS1 and PS2 are associated with FAD [47].

According to PubMed, research on ER stress in AD was started in 1997. The number of such research studies then gradually increased. Since 2016, more than 50 papers on ER stress in AD have been published intensively annually (Figure 4).

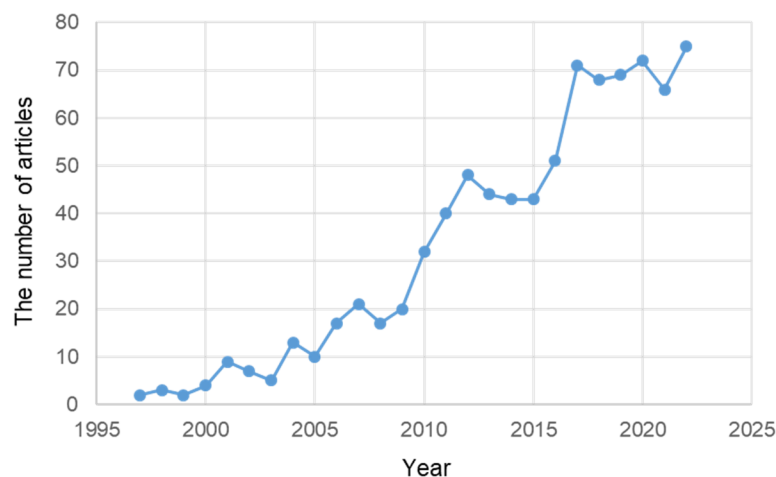


Figure 4. The number of articles related to ER stress in AD published in PubMed from 1997 to 23 November 2022.

The following papers are organized according to the year presented in PubMed: For the pathogenesis of neurodegeneration in AD, presenilin has been suggested to be related to ER [48,49]. PS1 mutations or loss of the PS1 function can affect the UPR, such as the expression of GRP78/BiP or CHOP, suggesting that PS1 can increase vulnerability to ER stress in association with the UPR [50,51]. Activation of the UPR in AD and increased GRP78/BiP and p-PERK in normal neurons suggest that the UPR is involved in AD

neurodegeneration [52]. ER stress can be a critical trigger in the neuronal response to inflammatory activity. This has been reviewed in terms of the pathology of AD [53].

A β s can activate the ER stress response factor X-box binding protein 1 (XBP-1) in transgenic flies and mammalian-cultured neurons [54]. AD pathogenesis involves ER stress and hyperphosphorylation of the tau protein [55]. Although the oligomeric form of A β s has been thought to play a critical role in AD, the mechanisms involved remain unclear in the pathogenesis of AD. A β s can induce ER stress in AD, suggesting that targeting this toxic component can enable the treatment of various neurodegenerative diseases [56]. Differential manifestations of ER stress and docosahexaenoic acid's (DHA's) reactivity may help explain the variable clinical results obtained with DHA treatment, suggesting that DHA might be adequate for a subset of AD patients [57]. The IgG Fc γ receptor II-b (Fc γ RIIb) can mediate A β neurotoxicity and neurodegeneration. Soluble A β oligomers can interact with Fc γ RIIb in vitro and in AD brains to activate ER stress and caspase-12 [58]. The pharmacological inhibition of brain inflammation and ER stress in mice can prevent glucose intolerance, demonstrating that AD-associated A β oligomers can affect the central nervous system [59,60]. This result suggests a novel molecular mechanism between metabolic disease and hypothalamic dysfunction in AD. The complex roles of ER stress, including XBP1 and PERK, during A β pathogenesis, indicate the status of disease progression [61]. Inflammasome-mediated peripheral inflammation can contribute to AD pathology [62]. ER stress and the UPR are critical to AD development and onset pathogenesis [63,64]. ER stress and the UPR in cellular aging and neuroinflammatory processes can lead to memory impairment and synapse dysfunction in AD [65].

Therefore, the target molecules of ER stress for AD are GRP78/BiP, CHOP, PERK, and XBP-1, suggesting that these ER stress molecules can act mainly as pathological factors in AD.

2.3. ER Stress and Amyotrophic Lateral Sclerosis (ALS)

ALS is a progressive neurodegenerative disease of neurons in the spinal cord and the brain, resulting in a loss of muscle control. Motor neuron loss continues until the person loses the ability to eat, speak, move, and breathe. ALS eventually causes paralysis and premature death due to respiratory failure [66–68]. ALS is most common in people aged 40 to 70 years, regardless of racial or ethnic group, although it can occur at younger ages. ALS has two types: sporadic ALS and familial ALS. Sporadic ALS is the most common form in the United States, accounting for approximately 90 to 95% of all cases. It occurs randomly without a known cause or family history. Familial ALS affects 5 to 10% of people and is inherited [69,70]. Although the disease usually progresses rapidly, the underlying causes of neuronal death are not fully known.

According to PubMed, research on ER stress in ALS started in 2003 and gradually increased. Since 2013, more than 20 papers have been published yearly (Figure 5).

The following papers are organized according to the year presented in PubMed: Motor neurons in the spinal cords of transgenic mice with the mutant SOD1 linked to familial amyotrophic lateral sclerosis (FALS) show a significant increase in GRP78/BiP just before motor symptoms begin, suggesting that ER stress is significantly related to the pathogenesis of FALS with SOD1 mutation [71]. When caspase-12 activation occurs in the spinal cord of a transformed ALS mouse, oxidation and ER-induced stress are induced, resulting in neuronal cell death and disease progression [72]. This suggests that caspase-12 and the ER stress pathways for neuronal apoptosis might be potential new targets for ALS treatment. The balance between the anti- and pro-apoptotic proteins associated with ER stress can induce damage by affecting the presymptomatic phase in an ALS mouse model, suggesting that this imbalance might be involved in the pathological cause of motor neuron degeneration in ALS [73]. Alterations in the fatty acid composition, mitochondrial function, and proteasome activity can be induced by excitotoxicity in the neurons, thereby inducing oxidative stress and, finally, endoplasmic reticulum stress in sporadic ALS [74]. The BH3-only protein, Puma, associated with ER stress, is critically involved in neurodegenerative

disease progression in SOD1G93A mice, indicating that Puma is an essential factor in controlling chronic neurodegeneration in ALS and other neurodegenerative disorders, including defects in the protein quality control [75]. SOD1-induced protein misfolding associated with ALS mutations can induce ER stress, suggesting that it might contribute to motor neuron degeneration in ALS pathogenesis [76].

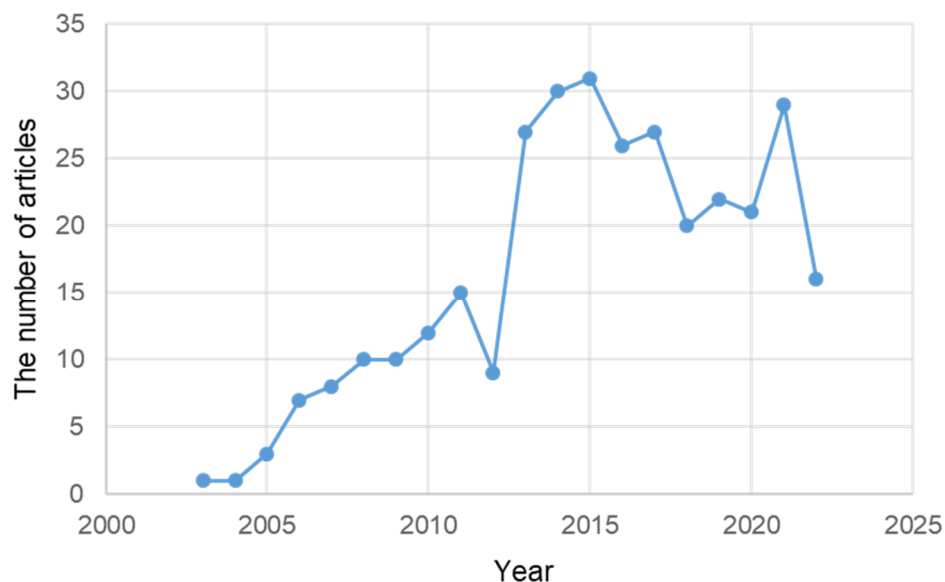


Figure 5. The number of articles related to ER stress in ALS published in PubMed from 2003 to 23 November 2022.

The interaction between Derlin-1 and SOD1 mutation can induce ER stress. Subsequent ASK1 activation is critical for disease progression in familial ALS [77]. Vesicle-associated membrane protein-associated protein B (VAPB), a novel ALS causative gene, plays a pivotal role in the UPR to ER stress. The ALS-linked P56S mutation in VAPB can abolish the function of VAPB, resulting in motor neuron vulnerability to ER stress [78]. A review article has presented ER stress and UPR dysfunction concerning ALS pathological mechanisms [79]. Although all motoneurons in ALS can be preferentially affected, the ER stress response in the specific motoneuron subtypes of ALS can affect the gradual onset of weakness and paralysis [80]. A new function of XBP-1 in autophagy indicates the critical correlation between the increased autophagy in the motoneurons and a reduced accumulation of mutant SOD1 aggregates, which can protect the neurons against neurodegeneration in ALS [81,82]. The upregulation of CHOP in the motor neurons and glial cells might play a pivotal role in the pathogenesis of ALS [83]. ER stress may act as a possible risk factor for the development of ALS by increasing the susceptibility of the wild-type SOD1 to aggregation during aging [84].

Therefore, the target molecules of ER stress for ALS are GRP78/BiP, CHOP, and XBP-1, suggesting that these ER stress molecules can act mainly as pathological factors in ALS.

2.4. ER Stress and Transmissible Spongiform Encephalopathies (TSEs)

TSEs or prion diseases are rare progressive neurodegenerative brain disorders affecting humans and animals with a spongy form in the brain. TSEs include Creutzfeldt–Jakob disease, bovine spongiform encephalopathy (BSE), and scrapie. The pathological hallmark of TSEs is the accumulation of a misfolded protease-resistant form of prion protein (PrP) in the cerebrum, leading to extensive neuronal cell death as the disease progresses [85–87].

According to PubMed, ER stress in TSE was not studied much until recently. However, in 2000, papers began to be published gradually. In 2015, 12 papers were published. The number of published papers gradually decreased until recently (Figure 6).

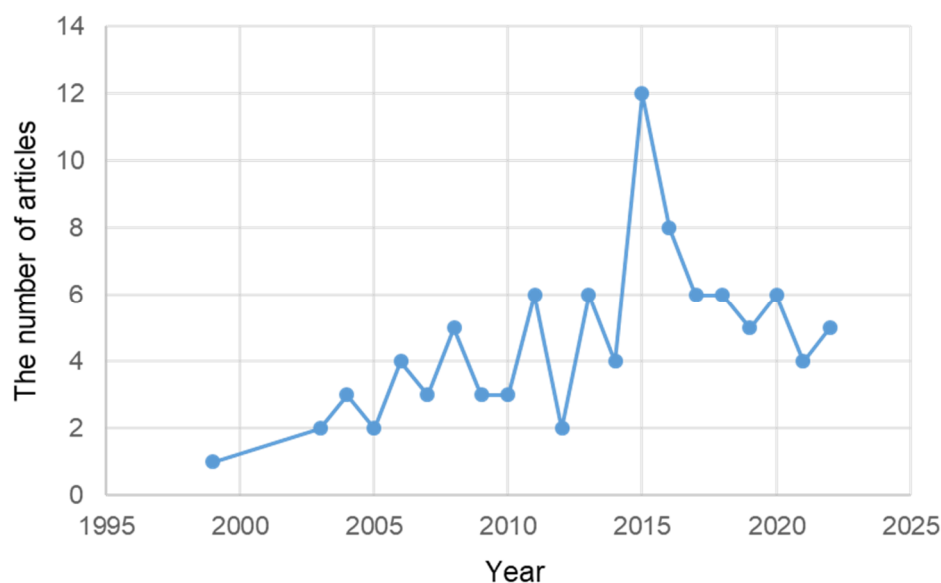


Figure 6. The number of articles related to ER stress in TSE published in PubMed from 1999 to 23 November 2022.

The following articles are organized by year of publication in PubMed: Caspase-12 is increased in cells showing misfolded prion protein (PrP^{Sc}). It is also increased in the brain tissues of PrP^{Sc}-infected mice and Creutzfeldt–Jakob disease patients [88]. This is the first study to present that ER stress and the activation of caspase-12 can cause neuronal cell death by accumulating mutant prion protein in the neurons. The overexpression of GRP58 can protect the cells from PrP^{Sc} toxicity and reduce caspase-12, suggesting that the expression of GRP58 can act as a neuroprotective factor against prion neurotoxicity [89]. Studies on GRP58 have suggested that it can be used to develop new targets for treating prion diseases [89]. The induction of ER stress in transgenic mice expressing PrP variants is accompanied by reduced translocation, a finding that could link age-dependent clinical and histological signs to PrP-mediated neurodegeneration [90]. One mechanism of prion-mediated neurodegeneration involves indirect ER stress-dependent effects on early PrP biosynthesis and metabolism [90]. The role of GRP78/BiP has been demonstrated to be important in prion diseases through *in vivo* and *in vitro* approaches [91]. The upregulation of PERK, GRP78/BiP, the ER protein, disulfide isomerase, and ubiquitin in prion disease indicates that ER stress and proteasome damage can initiate the early stages of spontaneous prion disease [92].

Therefore, the target molecules of ER stress for TSEs are GRP58, GRP78/BiP, and PERK, suggesting that these ER stress molecules can act mainly as pathological factors in TSEs.

2.5. ER Stress and Polyglutamine Diseases

Polyglutamine diseases are manifested by progressive neurodegeneration that can lead to behavioral and physical impairments. Polyglutamine diseases include Huntington's disease (HD), dentatorubral-pallidoluysian atrophy, spinal and bulbar muscular atrophy, and spinocerebellar ataxias 1, 2, 3, 6, 7, and 17 [93]. The disease features selective neuronal cell death and the accumulation of intracellular protein aggregates in cultured cells, transgenic animals, and human postmortem brain tissues. The pathophysiology of polyglutamine disorders with polyglutamine expansion and protein deposits remains unclear [94]. ASK1 is essential for neuropathological changes in polyglutamine diseases. It is crucial for killing neurons by inducing ER stress [95]. In HD, the N-terminal huntingtin protein can increase ER stress-related GRP78/BiP, CHOP, c-Jun-N-terminal kinase (JNK) phosphorylation, and caspase-12 activation [96]. Polyglutamine-expanded huntingtin fragments expressing yeast cells and neuron-like PC12 cells can impair the ER-associated degradation (ERAD) proteins

Npl4, Ufd1, and p97 [97]. Soluble oligomeric polyglutamine-expanded huntingtin can induce ER stress before its aggregation through p97/VCP [98].

Therefore, the target molecules of ER stress for polyglutamine diseases are ASK1, GRP78/BiP, CHOP, and ERAD, suggesting that these ER stress molecules can act mainly as pathological factors in polyglutamine diseases.

2.6. ER Stress and Neuronal Storage Diseases or Lysosomal Storage Disease (LSD)

Lysosomal storage disease (LSD) is a hereditary metabolic disorder with a defective lysosomal function. LSD includes more than 70 diseases with an abnormal lysosomal function, most of which are inherited as autosomal recessive traits. Although these disorders are rare individually, collectively, they occur in one in five thousand births [99]. Most LSDs show clinically progressive neurodegeneration, although symptoms of other organ systems are also frequently present. LSD has been classified and described in considerable detail previously [99].

Pelizaeus–Merzbacher disease (PMD) is an X-linked recessive pediatric and neurodegenerative disorder characterized by diffuse hypomyelination of the central nervous system. Mutations, duplications, or deletions of the phospholipid protein (PLP) gene can cause PMD by accumulating the PLP and/or DM20 proteins in the ER. CHOP in oligodendrocytes is directly involved in mice with the gene deletion causing PMD [100]. The accumulation of GM2-gangliosidosis in a mouse model of Sandhoff disease is caused by reduced calcium uptake into the ER due to decreased sarcoplasmic/ER calcium ATPase (SERCA), resulting in neuronal death [101]. GM1-ganglioside (GM1) is a significant sialoglycolipid in nerve cell membranes that can regulate calcium homeostasis. In a mouse model with GM1-gangliosidosis, the upregulation of GRP78/BiP and CHOP and the activation of JNK2 and caspase-12 can induce neuronal cell death, suggesting that the UPR can induce the accumulation of the sialoglylipid GM1 in the neuronal cells and then cause neuronal apoptosis [102]. CLN6 is a non-glycosylated ER-resident membrane protein with an unknown function. Mutant Cln6 can prevent the accumulation of the misfolded Cln6 protein by proteasomal degradation, impair constitutive autophagy by lysosomal dysfunction, and promote neurodegeneration [103]. A deficiency in the hexosaminidase activity can disrupt the normal metabolism of GM2-ganglioside, leading to progressive neurodegenerative diseases [104]. In cultured neurons, GM2-ganglioside accumulation in the ER can induce luminal Ca^{2+} depletion, activating PERK. Therefore, it can induce neurite atrophy and apoptosis [104]. Pelizaeus–Merzbacher-like disease type 1 (PMLD1) is a hypomyelination disorder in patients with mutations in the GJC2 coding for Connexin47 (Cx47). PMLD1 can cause nystagmus, cerebellar ataxia, convulsions, and changes in the CNS's white matter. In three PMLD1-related mutants (p.P87S, p.Y269D, and p.M283T), Cx47 can induce ER stress, including the UPR, and activate the apoptotic pathway [105].

Therefore, the target molecules of ER stress for LSD are GRP78/BiP, CHOP, PERK, IRE1, ATF6, and ATF4, suggesting that these ER stress molecules can act mainly as pathological factors in LSD.

2.7. ER Stress and Acute Neurodegeneration

ER stress can induce acute brain disorders such as cerebral ischemia and traumatic brain injury, apart from chronic neurodegenerative diseases. Oxygen-regulated protein 150 kDa (ORP150) and 94 kDa glucose-regulated protein (GRP94) as ER chaperones can be directly involved in the ER response in brain injury. ORP150 is elevated in the neurons of a human brain with ischemic stress [106]. In human neuroblastoma cells exposed to hypoxia/reoxygenation, GRP94 is increased [107]. Caspase-12-mediated ER apoptosis might be involved in rat traumatic brain injury pathology [108]. Primary hippocampal neurons in CHOP-deficient mice are resistant to hypoxia-reoxygenation-induced apoptosis, indicating that ischemia-induced hippocampal neuronal death is caused by the ER stress pathway associated with CHOP induction [109]. Global cerebral ischemia in transgenic

rats can increase the expression of ATF4 and CHOP, indicating that superoxide is involved in ER stress-induced cell death [110].

Therefore, the target molecules of ER stress for acute neurodegeneration are GRP94, CHOP, and ATF4, suggesting that these ER stress molecules can act mainly as pathological factors in acute neurodegeneration.

3. Melatonin

Melatonin is a potent antioxidant and free radical scavenger [111,112] and protects against inflammation, apoptosis, or autophagy in physiological and pathophysiological conditions [113–116]. In addition, melatonin modulates ER stress and UPR dysfunction in cancers, liver diseases, and other pathologies [115–117]. In particular, review articles introduce that melatonin can suppress various diseases related to ER stress: chronic intestinal inflammation and colon cancer [118], breast cancer [119], osteoarthritis [120], hepatocellular carcinoma [121], delaying ovarian aging [122], acute myocardial infarction [123], and diabetic cardiomyopathy [124].

Although there are many papers on the prevention or suppression of diseases by melatonin, there are very few papers about the effects of melatonin on ER stress in neurons and neurodegenerative diseases. The following describes the effects of melatonin reported so far, focusing on its effects on neurons and neurodegenerative diseases related to ER stress.

3.1. Melatonin and ER Stress in Neurons

Melatonin can significantly reduce the neuron splicing of the XBP-1 mRNA, increase eIF2 α phosphorylation, and elevate the expression of the chaperone proteins GRP78/BiP and Hsp70 in hypoxia-ischemia of a rat brain [125]. In addition, melatonin can significantly reduce CHOP expression. Melatonin can reduce ER stress induced by neonatal hypoxia-ischemia in newborn rats through SIRT-1 as the modulation and neuroprotection [125]. Melatonin can protect neuroblastoma cells against methamphetamine-induced ER stress and apoptosis by modulating CHOP, spliced XBP1, caspase-12, and caspase-3 [126]. Methamphetamine can induce ER stress in glial cells by stimulating the UPR to increase PERK phosphorylation, ATF6 expression, and the phosphorylated inositol-requiring enzyme 1 (p-IRE1). Moreover, the expression levels of GRP78/BiP and CHOP, caspase-12 activation, eIF2 α phosphorylation, and XBP-1 mRNA expression are increased. However, melatonin can reduce ER stress through methamphetamine toxicity by reducing the expression of the ER stress response genes and proteins [127]. Ischemia/reperfusion in acute neuronal injury after an ischemic stroke can induce neuronal apoptosis by increasing ER stress, including the phosphorylation of PERK and IRE1 and the expression of ATF6 and CHOP. Melatonin pretreatment can attenuate ischemia/reperfusion-induced ER stress [128]. Melatonin treatment can attenuate the insulin-induced mRNA or protein expression levels of ER stress markers such as p-eIF2 α , ATF4, CHOP, sXBP1, p-IRE1, and p-ASK1. Therefore, melatonin can regulate neuronal cell death induced by ER stress under insulin resistance by reducing the ER stress in SH-SY5Y neuroblastoma cells [129].

Melatonin can effectively downregulate the levels of ER stress molecules GRP78/BiP, CHOP, and caspase-12 proteins in kainic acid-induced N2a cells [130]. Melatonin can also inhibit tau hyperphosphorylation, the phosphorylation of PERK and eIF2 α , and the expression levels of ATF4 and GRP78/BiP in the kainic acid-treated mouse hippocampus [131]. Melatonin can attenuate ER stress, such as the phosphorylation of PERK and eIF2 α and the expression levels of ATF4 and CHOP in neurons exposed to oxygen and glucose deprivation and in rats subjected to transient focal cerebral ischemia [132]. These results indicate that melatonin suppresses post-ischemic ER stress after an ischemic stroke. The administration of melatonin can significantly decrease the mRNA and protein levels of ATF6 and CHOP in intracerebral hemorrhage rats, indicating that melatonin can protect neurons against apoptosis by suppressing ATF6 and CHOP [133]. Melatonin can decrease ER stress, such as the p-IRE1, p-PERK, GRP78/BiP, and CHOP, in the chronic cerebral hypoperfusion of rats, suggesting that melatonin can improve cognitive impairment following the induction

of bilateral typical carotid artery occlusion by attenuating AD markers and reducing ER stress [134].

Therefore, these studies show that the first target molecule of ER stress for melatonin is CHOP, and the second target molecules are PERK and GRP78/BiP. Therefore, melatonin is a crucial strategy to protect and treat neurons against ER stress.

3.2. Melatonin as ER Stress Modulator against Neurodegenerative Diseases

Direct oxidative damage and organelle lesions in rats can increase the expression of ER stress-related proteins, including GRP78/BiP and CHOP. The addition of melatonin can partially suppress ER stress-related behavioral and molecular disturbances. These results suggest that melatonin is directly related to AD-like behavioral and molecular pathologies with ER stress-related mechanisms [135]. One review article has presented the effects of melatonin on the fundamental ER stress mechanisms, focusing on its ability to modulate autophagy and apoptotic processes in the development of cancer cells, neurodegeneration, liver disease, and other pathologies [117].

Melatonin can protect mesenchymal stem cells from ischemic damage by inhibiting ER stress (such as decreasing the phosphorylation of PERK/eIF2 α /IRE1 α and the expression levels of ATF4 and CHOP proteins) and autophagy both in vivo and in vitro. In addition, melatonin can increase the cellular prion protein (PrPC) expression to prevent apoptotic cell death induced by ER stress and autophagy [136]. Melatonin is an endogenously produced molecule that can act as a copper chelator, a potent antioxidant, and an inhibitor of ER stress and the UPR in the liver and brain. Therefore, melatonin can potentially lower copper levels and limit the progression of Wilson's disease [137]. Melatonin can reduce A β peptide accumulation and ER stress markers such as the protein levels of GRP78/BiP, CHOP, and caspase-12 in an AlCl₃-treated AD model. Therefore, adding melatonin might be an alternative way to alleviate the onset of Alzheimer's disease [138].

Therefore, an in vivo study of melatonin has also revealed that the molecules of ER stress are CHOP, GRP78/BiP, and PERK. Therefore, melatonin has a significant strategic value in inhibiting and treating neurodegeneration caused by ER stress.

4. Conclusions

ER participates in various cellular processes, from which proteins are synthesized and transported to their final destinations after correct processing and modification. In neurodegenerative diseases, misprocessing occurs in many cellular processes related to ER, which can act as ER stress and finally cause neuronal cell death. However, two questions remain in this process: (1) how much cellular degeneration will occur due to ER stress? and (2) how much would another pathway induce cellular degeneration? The proteins that regulate or participate in the ER-mitochondrial interaction that causes cell death are not fully understood yet. ER stress molecules are involved in apoptosis, leading to a loss of ER function and the activation of the apoptosis cascade. Likewise, neuronal death occurs when proteins associated with neurodegenerative diseases accumulate to form cellular aggregates. This probably depends on the nature and function of the protein in question. Therefore, we think it is necessary to clearly understand and solve the mutual relationship between the molecules involved in ER stress and neuronal cell death.

Taken together, the reviewed findings suggest that we can target specific stages of the disease by focusing on the ER stress target molecules of melatonin to prevent or treat neurodegenerative diseases. This review suggests that CHOP, GRP78/BiP, and PERK are the ER stress target molecules of melatonin. Of course, more research studies are needed to test this possibility.

Author Contributions: Y.-M.Y. and S.S.J. conceived the manuscript's concept, wrote the paper, performed scientific and language editing, and revised the paper. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported by the Basic Science Research Program through the National Research Foundation of Korea (NRF), funded by the Ministry of Education (2020R111A1A01060627).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Paschen, W.; Frandsen, A. Endoplasmic reticulum dysfunction—A common denominator for cell injury in acute and degenerative diseases of the brain? *J. Neurochem.* **2001**, *79*, 719–725. [[CrossRef](#)]
2. Breckenridge, D.G.; Germain, M.; Mathai, J.P.; Nguyen, M.; Shore, G.C. Regulation of apoptosis by endoplasmic reticulum pathways. *Oncogene* **2003**, *22*, 8608–8618. [[CrossRef](#)]
3. Rao, R.V.; Ellerby, H.M.; Bredesen, D.E. Coupling endoplasmic reticulum stress to the cell death program. *Cell Death Differ.* **2004**, *11*, 372–380. [[CrossRef](#)]
4. Soto, C. Unfolding the role of protein misfolding in neurodegenerative diseases. *Nat. Rev. Neurosci.* **2003**, *4*, 49–60. [[CrossRef](#)]
5. Bence, N.F.; Sampat, R.M.; Kopito, R.R. Impairment of the ubiquitin-proteasome system by protein aggregation. *Science* **2001**, *292*, 1552–1555. [[CrossRef](#)]
6. Ciechanover, A.; Brundin, P. The ubiquitin proteasome system in neurodegenerative diseases: Sometimes the chicken, sometimes the egg. *Neuron* **2003**, *40*, 427–446. [[CrossRef](#)] [[PubMed](#)]
7. Imai, Y.; Soda, M.; Inoue, H.; Hattori, N.; Mizuno, Y.; Takahashi, R. An unfolded putative transmembrane polypeptide, which can lead to endoplasmic reticulum stress, is a substrate of Parkin. *Cell* **2001**, *105*, 891–902. [[CrossRef](#)]
8. Di Bella, G.; Mascia, F.; Gualano, L.; Di Bella, L. Melatonin anticancer effects: Review. *Int. J. Mol. Sci.* **2013**, *14*, 2410–2440. [[CrossRef](#)]
9. Lee, Y. Roles of circadian clocks in cancer pathogenesis and treatment. *Exp. Mol. Med.* **2021**, *53*, 1529–1538. [[CrossRef](#)]
10. Healy, D.G.; Abou-Sleiman, P.M.; Wood, N.W. PINK, PANK, or PARK? A clinicians' guide to familial parkinsonism. *Lancet Neurol.* **2004**, *3*, 652–662. [[CrossRef](#)]
11. Greenamyre, J.T.; Hastings, T.G. Biomedicine. Parkinson's—divergent causes, convergent mechanisms. *Science* **2004**, *304*, 1120–1122. [[CrossRef](#)]
12. Ryu, E.J.; Harding, H.P.; Angelastro, J.M.; Vitolo, O.V.; Ron, D.; Greene, L.A. Endoplasmic reticulum stress and the unfolded protein response in cellular models of Parkinson's disease. *J. Neurosci.* **2002**, *22*, 10690–10698. [[CrossRef](#)]
13. Holtz, W.A.; O'Malley, K.L. Parkinsonian mimetics induce aspects of unfolded protein response in death of dopaminergic neurons. *J. Biol. Chem.* **2003**, *278*, 19367–19377. [[CrossRef](#)]
14. Ishibashi, T.; Matsuo, N.; Ogawa, S.; Tohyama, M. RA410/Sly1 suppresses MPP+ and 6-hydroxydopamine-induced cell death in SH-SY5Y cells. *Neurobiol. Dis.* **2005**, *18*, 143–151. [[CrossRef](#)]
15. Takahashi, R.; Imai, Y.; Hattori, N.; Mizuno, Y. Parkin and endoplasmic reticulum stress. *Ann. N. Y. Acad. Sci.* **2003**, *991*, 101–106. [[CrossRef](#)] [[PubMed](#)]
16. Yamagishi, S.; Matsuda, S.; Hitomi, J.; Miyata, S.; Mizuno, T.; Imaizumi, K.; Katayama, T.; Tohyama, M. JAB1 participates in unfolded protein responses by association and dissociation with IRE1. *Neurochem. Int.* **2004**, *45*, 765–772. [[CrossRef](#)]
17. Smith, W.W.; Jiang, H.; Pei, Z.; Tanaka, Y.; Morita, H.; Sawa, A.; Dawson, V.L.; Dawson, T.M.; Ross, C.A. Endoplasmic reticulum stress and mitochondrial cell death pathways mediate A53T mutant alpha-synuclein-induced toxicity. *Hum. Mol. Genet.* **2005**, *14*, 3801–3811. [[CrossRef](#)]
18. Jiang, P.; Gan, M.; Ebrahim, A.S.; Lin, W.L.; Melrose, H.L.; Yen, S.H. ER stress response plays an important role in aggregation of α -synuclein. *Mol. Neurodegener.* **2010**, *5*, 56. [[CrossRef](#)] [[PubMed](#)]
19. Colla, E.; Jensen, P.H.; Pletnikova, O.; Troncoso, J.C.; Glabe, C.; Lee, M.K. Accumulation of toxic α -synuclein oligomer within endoplasmic reticulum occurs in α -synucleinopathy in vivo. *J. Neurosci.* **2012**, *32*, 3301–3305. [[CrossRef](#)] [[PubMed](#)]
20. Heman-Ackah, S.M.; Manzano, R.; Hoozemans, J.J.M.; Scheper, W.; Flynn, R.; Haerty, W.; Cowley, S.A.; Bassett, A.R.; Wood, M.J.A. Alpha-synuclein induces the unfolded protein response in Parkinson's disease SNCA triplication iPSC-derived neurons. *Hum. Mol. Genet.* **2017**, *26*, 4441–4450. [[CrossRef](#)]
21. Stojkowska, I.; Wani, W.Y.; Zunke, F.; Belur, N.R.; Pavlenko, E.A.; Mwenda, N.; Sharma, K.; Francelle, L.; Mazzulli, J.R. Rescue of α -synuclein aggregation in Parkinson's patient neurons by synergistic enhancement of ER proteostasis and protein trafficking. *Neuron* **2022**, *110*, 436–451.e11. [[CrossRef](#)]
22. Yamamuro, A.; Yoshioka, Y.; Ogita, K.; Maeda, S. Involvement of endoplasmic reticulum stress on the cell death induced by 6-hydroxydopamine in human neuroblastoma SH-SY5Y cells. *Neurochem. Res.* **2006**, *31*, 657–664. [[CrossRef](#)]

23. Chua, C.E.; Tang, B.L. Alpha-synuclein and Parkinson's disease: The first roadblock. *J. Cell. Mol. Med.* **2006**, *10*, 837–846. [[CrossRef](#)]
24. Lashuel, H.A.; Hirling, H. Rescuing defective vesicular trafficking protects against alpha-synuclein toxicity in cellular and animal models of Parkinson's disease. *ACS Chem. Biol.* **2006**, *1*, 420–424. [[CrossRef](#)]
25. Olivares, D.; Huang, X.; Branden, L.; Greig, N.H.; Rogers, J.T. Physiological and pathological role of alpha-synuclein in Parkinson's disease through iron mediated oxidative stress; the role of a putative iron-responsive element. *Int. J. Mol. Sci.* **2009**, *10*, 1226–1260. [[CrossRef](#)]
26. Hoozemans, J.J.; van Haastert, E.S.; Eikelenboom, P.; de Vos, R.A.; Rozemuller, J.M.; Scheper, W. Activation of the unfolded protein response in Parkinson's disease. *Biochem. Biophys. Res. Commun.* **2007**, *354*, 707–711. [[CrossRef](#)]
27. Wang, H.Q.; Takahashi, R. Expanding insights on the involvement of endoplasmic reticulum stress in Parkinson's disease. *Antioxid. Redox Signal.* **2007**, *9*, 553–561. [[CrossRef](#)] [[PubMed](#)]
28. Hosoi, T.; Ozawa, K. Endoplasmic reticulum stress in disease: Mechanisms and therapeutic opportunities. *Clin. Sci.* **2009**, *118*, 19–29. [[CrossRef](#)]
29. Su, L.J.; Auluck, P.K.; Outeiro, T.F.; Yeager-Lotem, E.; Kritzer, J.A.; Tardiff, D.F.; Strathearn, K.E.; Liu, F.; Cao, S.; Hamamichi, S.; et al. Compounds from an unbiased chemical screen reverse both ER-to-Golgi trafficking defects and mitochondrial dysfunction in Parkinson's disease models. *Dis. Model. Mech.* **2010**, *3*, 194–208. [[CrossRef](#)]
30. Bouman, L.; Schlierf, A.; Lutz, A.K.; Shan, J.; Deinlein, A.; Kast, J.; Galehdar, Z.; Palmisano, V.; Patenge, N.; Berg, D.; et al. Parkinson is transcriptionally regulated by ATF4: Evidence for an interconnection between mitochondrial stress and ER stress. *Cell Death Differ.* **2011**, *18*, 769–782. [[CrossRef](#)]
31. Bellucci, A.; Navarria, L.; Zaltieri, M.; Falarti, E.; Bodei, S.; Sigala, S.; Battistin, L.; Spillantini, M.; Missale, C.; Spano, P. Induction of the unfolded protein response by α -synuclein in experimental models of Parkinson's disease. *J. Neurochem.* **2011**, *116*, 588–605. [[CrossRef](#)]
32. Cali, T.; Ottolini, D.; Brini, M. Mitochondria, calcium, and endoplasmic reticulum stress in Parkinson's disease. *Biofactors* **2011**, *37*, 228–240. [[CrossRef](#)]
33. Doyle, K.M.; Kennedy, D.; Gorman, A.M.; Gupta, S.; Healy, S.J.; Samali, A. Unfolded proteins and endoplasmic reticulum stress in neurodegenerative disorders. *J. Cell. Mol. Med.* **2011**, *15*, 2025–2039. [[CrossRef](#)]
34. Ghemrawi, R.; Khair, M. Endoplasmic Reticulum Stress and Unfolded Protein Response in Neurodegenerative Diseases. *Int. J. Mol. Sci.* **2020**, *21*, 6127. [[CrossRef](#)] [[PubMed](#)]
35. Costa, C.A.D.; Manaa, W.E.; Duplan, E.; Checler, F. The Endoplasmic Reticulum Stress/Unfolded Protein Response and Their Contributions to Parkinson's Disease Physiopathology. *Cells* **2020**, *9*, 2495. [[CrossRef](#)] [[PubMed](#)]
36. Ren, H.; Zhai, W.; Lu, X.; Wang, G. The Cross-Links of Endoplasmic Reticulum Stress, Autophagy, and Neurodegeneration in Parkinson's Disease. *Front. Aging Neurosci.* **2021**, *13*, 691881. [[CrossRef](#)] [[PubMed](#)]
37. Kim, S.; Kim, D.K.; Jeong, S.; Lee, J. The Common Cellular Events in the Neurodegenerative Diseases and the Associated Role of Endoplasmic Reticulum Stress. *Int. J. Mol. Sci.* **2022**, *23*, 5894. [[CrossRef](#)] [[PubMed](#)]
38. Credle, J.J.; Forcelli, P.A.; Delannoy, M.; Oaks, A.W.; Permaul, E.; Berry, D.L.; Duka, V.; Wills, J.; Sidhu, A. α -Synuclein-mediated inhibition of ATF6 processing into COPII vesicles disrupts UPR signaling in Parkinson's disease. *Neurobiol. Dis.* **2015**, *76*, 112–125. [[CrossRef](#)]
39. Tsujii, S.; Ishisaka, M.; Hara, H. Modulation of endoplasmic reticulum stress in Parkinson's disease. *Eur. J. Pharmacol.* **2015**, *765*, 154–156. [[CrossRef](#)]
40. Baek, J.H.; Whitfield, D.; Howlett, D.; Francis, P.; Berezcki, E.; Ballard, C.; Hortobágyi, T.; Attems, J.; Aarsland, D. Unfolded protein response is activated in Lewy body dementias. *Neuropathol. Appl. Neurobiol.* **2016**, *42*, 352–365. [[CrossRef](#)]
41. Gully, J.C.; Sergeev, V.G.; Bhootada, Y.; Mendez-Gomez, H.; Meyers, C.A.; Zolotukhin, S.; Gorbatyuk, M.S.; Gorbatyuk, O.S. Up-regulation of activating transcription factor 4 induces severe loss of dopamine nigral neurons in a rat model of Parkinson's disease. *Neurosci. Lett.* **2016**, *627*, 36–41. [[CrossRef](#)]
42. Demmings, M.D.; Tennyson, E.C.; Petroff, G.N.; Tarnowski-Garner, H.E.; Cregan, S.P. Activating transcription factor-4 promotes neuronal death induced by Parkinson's disease neurotoxins and α -synuclein aggregates. *Cell Death Differ.* **2021**, *28*, 1627–1643. [[CrossRef](#)]
43. Yan, C.; Liu, J.; Gao, J.; Sun, Y.; Zhang, L.; Song, H.; Xue, L.; Zhan, L.; Gao, G.; Ke, Z.; et al. IRE1 promotes neurodegeneration through autophagy-dependent neuron death in the Drosophila model of Parkinson's disease. *Cell Death Dis.* **2019**, *10*, 800. [[CrossRef](#)]
44. Mou, Z.; Yuan, Y.H.; Zhang, Z.; Song, L.K.; Chen, N.H. Endoplasmic reticulum stress, an important factor in the development of Parkinson's disease. *Toxicol. Lett.* **2020**, *324*, 20–29. [[CrossRef](#)] [[PubMed](#)]
45. Elvira, R.; Cha, S.J.; Noh, G.M.; Kim, K.; Han, J. PERK-Mediated eIF2 α Phosphorylation Contributes to The Protection of Dopaminergic Neurons from Chronic Heat Stress in Drosophila. *Int. J. Mol. Sci.* **2020**, *21*, 845. [[CrossRef](#)] [[PubMed](#)]
46. Shacham, T.; Patel, C.; Lederkremer, G.Z. PERK Pathway and Neurodegenerative Disease: To Inhibit or to Activate? *Biomolecules* **2021**, *11*, 354. [[CrossRef](#)]
47. Knopman, D.S.; Amieva, H.; Petersen, R.C.; Chételat, G.; Holtzman, D.M.; Hyman, B.T.; Nixon, R.A.; Jones, D.T. Alzheimer disease. *Nat. Rev. Dis. Prim.* **2021**, *7*, 33. [[CrossRef](#)]

48. Mattson, M.P.; Guo, Q. Cell and molecular neurobiology of presenilins: A role for the endoplasmic reticulum in the pathogenesis of Alzheimer's disease? *J. Neurosci. Res.* **1997**, *50*, 505–513. [[CrossRef](#)]
49. Mattson, M.P.; Guo, Q.; Furukawa, K.; Pedersen, W.A. Presenilins, the endoplasmic reticulum, and neuronal apoptosis in Alzheimer's disease. *J. Neurochem.* **1998**, *70*, 1–14. [[CrossRef](#)] [[PubMed](#)]
50. Katayama, T.; Imaizumi, K.; Sato, N.; Miyoshi, K.; Kudo, T.; Hitomi, J.; Morihara, T.; Yoneda, T.; Gomi, F.; Mori, Y.; et al. Presenilin-1 mutations downregulate the signalling pathway of the unfolded-protein response. *Nat. Cell Biol.* **1999**, *1*, 479–485. [[CrossRef](#)]
51. Sato, N.; Urano, F.; Yoon Leem, J.; Kim, S.H.; Li, M.; Donoviel, D.; Bernstein, A.; Lee, A.S.; Ron, D.; Veselits, M.L.; et al. Upregulation of BiP and CHOP by the unfolded-protein response is independent of presenilin expression. *Nat. Cell Biol.* **2000**, *2*, 863–870. [[CrossRef](#)]
52. Hoozemans, J.J.; Veerhuis, R.; Van Haastert, E.S.; Rozemuller, J.M.; Baas, F.; Eikelenboom, P.; Scheper, W. The unfolded protein response is activated in Alzheimer's disease. *Acta Neuropathol.* **2005**, *110*, 165–172. [[CrossRef](#)]
53. Salminen, A.; Kauppinen, A.; Suuronen, T.; Kaarniranta, K.; Ojala, J. ER stress in Alzheimer's disease: A novel neuronal trigger for inflammation and Alzheimer's pathology. *J. Neuroinflamm.* **2009**, *6*, 41. [[CrossRef](#)] [[PubMed](#)]
54. Casas-Tinto, S.; Zhang, Y.; Sanchez-Garcia, J.; Gomez-Velazquez, M.; Rincon-Limas, D.E.; Fernandez-Funez, P. The ER stress factor XBP1s prevents amyloid-beta neurotoxicity. *Hum. Mol. Genet.* **2011**, *20*, 2144–2160. [[CrossRef](#)]
55. Ho, Y.S.; Yang, X.; Lau, J.C.; Hung, C.H.; Wuwongse, S.; Zhang, Q.; Wang, J.; Baum, L.; So, K.F.; Chang, R.C. Endoplasmic reticulum stress induces tau pathology and forms a vicious cycle: Implication in Alzheimer's disease pathogenesis. *J. Alzheimers Dis.* **2012**, *28*, 839–854. [[CrossRef](#)] [[PubMed](#)]
56. Roussel, B.D.; Kruppa, A.J.; Miranda, E.; Crowther, D.C.; Lomas, D.A.; Marciniak, S.J. Endoplasmic reticulum dysfunction in neurological disease. *Lancet Neurol.* **2013**, *12*, 105–118. [[CrossRef](#)]
57. Kondo, T.; Asai, M.; Tsukita, K.; Kutoku, Y.; Ohsawa, Y.; Sunada, Y.; Imamura, K.; Egawa, N.; Yahata, N.; Okita, K.; et al. Modeling Alzheimer's disease with iPSCs reveals stress phenotypes associated with intracellular A β and differential drug responsiveness. *Cell Stem Cell* **2013**, *12*, 487–496. [[CrossRef](#)] [[PubMed](#)]
58. Kam, T.I.; Song, S.; Gwon, Y.; Park, H.; Yan, J.J.; Im, I.; Choi, J.W.; Choi, T.Y.; Kim, J.; Song, D.K.; et al. Fc γ RIIb mediates amyloid- β neurotoxicity and memory impairment in Alzheimer's disease. *J. Clin. Investig.* **2013**, *123*, 2791–2802. [[CrossRef](#)] [[PubMed](#)]
59. Clarke, J.R.; Lyra e Silva, N.M.; Figueiredo, C.P.; Frozza, R.L.; Ledo, J.H.; Beckman, D.; Katashima, C.K.; Razolli, D.; Carvalho, B.M.; Frazão, R.; et al. Alzheimer-associated A β oligomers impact the central nervous system to induce peripheral metabolic deregulation. *EMBO Mol. Med.* **2015**, *7*, 190–210. [[CrossRef](#)]
60. Sprengle, N.T.; Sims, S.G.; Sánchez, C.L.; Meares, G.P. Endoplasmic reticulum stress and inflammation in the central nervous system. *Mol. Neurodegener.* **2017**, *12*, 42. [[CrossRef](#)]
61. Cheng, K.C.; Chiang, H.C. XBP1 and PERK Have Distinct Roles in A β -Induced Pathology. *Mol. Neurobiol.* **2018**, *55*, 7523–7532. [[CrossRef](#)]
62. Pereira, C.F.; Santos, A.E.; Moreira, P.I.; Pereira, A.C.; Sousa, F.J.; Cardoso, S.M.; Cruz, M.T. Is Alzheimer's disease an inflammasomopathy? *Ageing Res. Rev.* **2019**, *56*, 100966. [[CrossRef](#)] [[PubMed](#)]
63. Uddin, M.S.; Tewari, D.; Sharma, G.; Kabir, M.T.; Barreto, G.E.; Bin-Jumah, M.N.; Perveen, A.; Abdel-Daim, M.M.; Ashraf, G.M. Molecular Mechanisms of ER Stress and UPR in the Pathogenesis of Alzheimer's Disease. *Mol. Neurobiol.* **2020**, *57*, 2902–2919. [[CrossRef](#)]
64. Ajoolabady, A.; Lindholm, D.; Ren, J.; Pratico, D. ER stress and UPR in Alzheimer's disease: Mechanisms, pathogenesis, treatments. *Cell Death Dis.* **2022**, *13*, 706. [[CrossRef](#)]
65. Uddin, M.S.; Yu, W.S.; Lim, L.W. Exploring ER stress response in cellular aging and neuroinflammation in Alzheimer's disease. *Ageing Res. Rev.* **2021**, *70*, 101417. [[CrossRef](#)]
66. Bruijn, L.I.; Miller, T.M.; Cleveland, D.W. Unraveling the mechanisms involved in motor neuron degeneration in ALS. *Annu. Rev. Neurosci.* **2004**, *27*, 723–749. [[CrossRef](#)]
67. Hilton, J.B.; White, A.R.; Crouch, P.J. Metal-deficient SOD1 in amyotrophic lateral sclerosis. *J. Mol. Med.* **2015**, *93*, 481–487. [[CrossRef](#)]
68. Schweingruber, C.; Hedlund, E. The Cell Autonomous and Non-Cell Autonomous Aspects of Neuronal Vulnerability and Resilience in Amyotrophic Lateral Sclerosis. *Biology* **2022**, *11*, 1191. [[CrossRef](#)]
69. Leblond, C.S.; Kaneb, H.M.; Dion, P.A.; Rouleau, G.A. Dissection of genetic factors associated with amyotrophic lateral sclerosis. *Exp. Neurol.* **2014**, *262*, 91–101. [[CrossRef](#)] [[PubMed](#)]
70. Van Damme, P.; Robberecht, W.; Van Den Bosch, L. Modelling amyotrophic lateral sclerosis: Progress and possibilities. *Dis. Model. Mech.* **2017**, *10*, 537–549. [[CrossRef](#)] [[PubMed](#)]
71. Tobisawa, S.; Hozumi, Y.; Arawaka, S.; Koyama, S.; Wada, M.; Nagai, M.; Aoki, M.; Itoyama, Y.; Goto, K.; Kato, T. Mutant SOD1 linked to familial amyotrophic lateral sclerosis, but not wild-type SOD1, induces ER stress in COS7 cells and transgenic mice. *Biochem. Biophys. Res. Commun.* **2003**, *303*, 496–503. [[CrossRef](#)]
72. Wootz, H.; Hansson, I.; Korhonen, L.; Näpänkangas, U.; Lindholm, D. Caspase-12 cleavage and increased oxidative stress during motoneuron degeneration in transgenic mouse model of ALS. *Biochem. Biophys. Res. Commun.* **2004**, *322*, 281–286. [[CrossRef](#)] [[PubMed](#)]

73. Nagata, T.; Ilieva, H.; Murakami, T.; Shiote, M.; Narai, H.; Ohta, Y.; Hayashi, T.; Shoji, M.; Abe, K. Increased ER stress during motor neuron degeneration in a transgenic mouse model of amyotrophic lateral sclerosis. *Neurol. Res.* **2007**, *29*, 767–771. [[CrossRef](#)] [[PubMed](#)]
74. Ilieva, E.V.; Ayala, V.; Jové, M.; Dalfó, E.; Cacabelos, D.; Povedano, M.; Bellmunt, M.J.; Ferrer, I.; Pamplona, R.; Portero-Otín, M. Oxidative and endoplasmic reticulum stress interplay in sporadic amyotrophic lateral sclerosis. *Brain* **2007**, *130 Pt 12*, 3111–3123. [[CrossRef](#)] [[PubMed](#)]
75. Kieran, D.; Woods, I.; Villunger, A.; Strasser, A.; Prehn, J.H. Deletion of the BH3-only protein puma protects motoneurons from ER stress-induced apoptosis and delays motoneuron loss in ALS mice. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 20606–20611. [[CrossRef](#)]
76. Oh, Y.K.; Shin, K.S.; Yuan, J.; Kang, S.J. Superoxide dismutase 1 mutants related to amyotrophic lateral sclerosis induce endoplasmic stress in neuro2a cells. *J. Neurochem.* **2008**, *104*, 993–1005. [[CrossRef](#)]
77. Nishitoh, H.; Kadowaki, H.; Nagai, A.; Maruyama, T.; Yokota, T.; Fukutomi, H.; Noguchi, T.; Matsuzawa, A.; Takeda, K.; Ichijo, H. ALS-linked mutant SOD1 induces ER stress- and ASK1-dependent motor neuron death by targeting Derlin-1. *Genes Dev.* **2008**, *22*, 1451–1464. [[CrossRef](#)]
78. Suzuki, H.; Kanekura, K.; Levine, T.P.; Kohno, K.; Olkkonen, V.M.; Aiso, S.; Matsuoka, M. ALS-linked P56S-VAPB, an aggregated loss-of-function mutant of VAPB, predisposes motor neurons to ER stress-related death by inducing aggregation of co-expressed wild-type VAPB. *J. Neurochem.* **2009**, *108*, 973–985. [[CrossRef](#)]
79. Kanekura, K.; Suzuki, H.; Aiso, S.; Matsuoka, M. ER stress and unfolded protein response in amyotrophic lateral sclerosis. *Mol. Neurobiol.* **2009**, *39*, 81–89. [[CrossRef](#)]
80. Saxena, S.; Cabuy, E.; Caroni, P. A role for motoneuron subtype-selective ER stress in disease manifestations of FALS mice. *Nat. Neurosci.* **2009**, *12*, 627–636. [[CrossRef](#)]
81. Hetz, C.; Thielen, P.; Matus, S.; Nassif, M.; Court, F.; Kiffin, R.; Martinez, G.; Cuervo, A.M.; Brown, R.H.; Glimcher, L.H. XBP-1 deficiency in the nervous system protects against amyotrophic lateral sclerosis by increasing autophagy. *Genes Dev.* **2009**, *23*, 2294–2306. [[CrossRef](#)] [[PubMed](#)]
82. Matus, S.; Nassif, M.; Glimcher, L.H.; Hetz, C. XBP-1 deficiency in the nervous system reveals a homeostatic switch to activate autophagy. *Autophagy* **2009**, *5*, 1226–1228. [[CrossRef](#)]
83. Ito, Y.; Yamada, M.; Tanaka, H.; Aida, K.; Tsuruma, K.; Shimazawa, M.; Hozumi, I.; Inuzuka, T.; Takahashi, H.; Hara, H. Involvement of CHOP, an ER-stress apoptotic mediator, in both human sporadic ALS and ALS model mice. *Neurobiol. Dis.* **2009**, *36*, 470–476. [[CrossRef](#)]
84. Medinas, D.B.; Rozas, P.; Martínez Traub, F.; Woehlbier, U.; Brown, R.H.; Bosco, D.A.; Hetz, C. Endoplasmic reticulum stress leads to accumulation of wild-type SOD1 aggregates associated with sporadic amyotrophic lateral sclerosis. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, 8209–8214. [[CrossRef](#)] [[PubMed](#)]
85. Prusiner, S.B. Prions. *Proc. Natl. Acad. Sci. USA* **1998**, *95*, 13363–13383. [[CrossRef](#)] [[PubMed](#)]
86. Aguzzi, A.; Heppner, F.L. Pathogenesis of prion diseases: A progress report. *Cell Death Differ.* **2000**, *7*, 889–902. [[CrossRef](#)] [[PubMed](#)]
87. Soto, C.; Castilla, J. The controversial protein-only hypothesis of prion propagation. *Nat. Med.* **2004**, *10*, S63–S67. [[CrossRef](#)] [[PubMed](#)]
88. Hetz, C.; Russelakis-Carneiro, M.; Maundrell, K.; Castilla, J.; Soto, C. Caspase-12 and endoplasmic reticulum stress mediate neurotoxicity of pathological prion protein. *EMBO J.* **2003**, *22*, 5435–5445. [[CrossRef](#)]
89. Hetz, C.; Russelakis-Carneiro, M.; Wälchli, S.; Carboni, S.; Vial-Knecht, E.; Maundrell, K.; Castilla, J.; Soto, C. The disulfide isomerase Grp58 is a protective factor against prion neurotoxicity. *J. Neurosci.* **2005**, *25*, 2793–2802. [[CrossRef](#)] [[PubMed](#)]
90. Rane, N.S.; Kang, S.W.; Chakrabarti, O.; Feigenbaum, L.; Hegde, R.S. Reduced translocation of nascent prion protein during ER stress contributes to neurodegeneration. *Dev. Cell* **2008**, *15*, 359–370. [[CrossRef](#)]
91. Park, K.W.; Eun Kim, G.; Morales, R.; Moda, F.; Moreno-Gonzalez, I.; Concha-Marambio, L.; Lee, A.S.; Hetz, C.; Soto, C. The Endoplasmic Reticulum Chaperone GRP78/BiP Modulates Prion Propagation in vitro and in vivo. *Sci. Rep.* **2017**, *7*, 44723. [[CrossRef](#)]
92. Otero, A.; Betancor, M.; Eraña, H.; Fernández Borges, N.; Lucas, J.J.; Badiola, J.J.; Castilla, J.; Bolea, R. Prion-Associated Neurodegeneration Causes Both Endoplasmic Reticulum Stress and Proteasome Impairment in a Murine Model of Spontaneous Disease. *Int. J. Mol. Sci.* **2021**, *22*, 465. [[CrossRef](#)]
93. Zoghbi, H.Y.; Orr, H.T. Glutamine repeats and neurodegeneration. *Annu. Rev. Neurosci.* **2000**, *23*, 217–247. [[CrossRef](#)] [[PubMed](#)]
94. Lipinski, M.M.; Yuan, J. Mechanisms of cell death in polyglutamine expansion diseases. *Curr. Opin. Pharmacol.* **2004**, *4*, 85–90. [[CrossRef](#)] [[PubMed](#)]
95. Nishitoh, H.; Matsuzawa, A.; Tobiume, K.; Saegusa, K.; Takeda, K.; Inoue, K.; Hori, S.; Kakizuka, A.; Ichijo, H. ASK1 is essential for endoplasmic reticulum stress-induced neuronal cell death triggered by expanded polyglutamine repeats. *Genes Dev.* **2002**, *16*, 1345–1355. [[CrossRef](#)] [[PubMed](#)]
96. Reijonen, S.; Putkonen, N.; Nørremølle, A.; Lindholm, D.; Korhonen, L. Inhibition of endoplasmic reticulum stress counteracts neuronal cell death and protein aggregation caused by N-terminal mutant huntingtin proteins. *Exp. Cell Res.* **2008**, *314*, 950–960. [[CrossRef](#)]
97. Duennwald, M.L.; Lindquist, S. Impaired ERAD and ER stress are early and specific events in polyglutamine toxicity. *Genes Dev.* **2008**, *22*, 3308–3319. [[CrossRef](#)] [[PubMed](#)]

98. Leitman, J.; Ulrich Hartl, F.; Lederkremer, G.Z. Soluble forms of polyQ-expanded huntingtin rather than large aggregates cause endoplasmic reticulum stress. *Nat. Commun.* **2013**, *4*, 2753. [[CrossRef](#)]
99. Platt, F.M.; d'Azzo, A.; Davidson, B.L.; Neufeld, E.F.; Tiffet, C.J. Lysosomal storage diseases. *Nat. Rev. Dis. Primers* **2018**, *4*, 27. [[CrossRef](#)]
100. Southwood, C.M.; Garbern, J.; Jiang, W.; Gow, A. The unfolded protein response modulates disease severity in Pelizaeus-Merzbacher disease. *Neuron* **2002**, *36*, 585–596. [[CrossRef](#)]
101. Pelled, D.; Lloyd-Evans, E.; Riebeling, C.; Jeyakumar, M.; Platt, F.M.; Futerman, A.H. Inhibition of calcium uptake via the sarco/endoplasmic reticulum Ca^{2+} -ATPase in a mouse model of Sandhoff disease and prevention by treatment with N-butyldeoxynojirimycin. *J. Biol. Chem.* **2003**, *278*, 29496–29501. [[CrossRef](#)] [[PubMed](#)]
102. Tessitore, A.; Martin, M.D.; Sano, R.; Ma, Y.; Mann, L.; Ingrassia, A.; Laywell, E.D.; Steindler, D.A.; Hendershot, L.M.; d'Azzo, A. GM1-ganglioside-mediated activation of the unfolded protein response causes neuronal death in a neurodegenerative gangliosidosis. *Mol. Cell* **2004**, *15*, 753–766. [[CrossRef](#)] [[PubMed](#)]
103. Thelen, M.; Damme, M.; Schweizer, M.; Hagel, C.; Wong, A.M.; Cooper, J.D.; Bräulke, T.; Galliciotti, G. Disruption of the autophagy-lysosome pathway is involved in neuropathology of the *ncl* mouse model of neuronal ceroid lipofuscinosis. *PLoS ONE* **2012**, *7*, e35493. [[CrossRef](#)]
104. Virgolini, M.J.; Feliziani, C.; Cambiasso, M.J.; Lopez, P.H.; Bollo, M. Neurite atrophy and apoptosis mediated by PERK signaling after accumulation of GM2-ganglioside. *Biochim. Biophys. Acta Mol. Cell Res.* **2019**, *1866*, 225–239. [[CrossRef](#)] [[PubMed](#)]
105. Flores-Obando, R.E.; Freidin, M.M.; Hernández, A.I.; Abrams, C.K. Activation of the unfolded protein response by Connexin47 mutations associated with Pelizaeus-Merzbacher-like disease. *Mol. Cell. Neurosci.* **2022**, *120*, 103716. [[CrossRef](#)]
106. Tamatani, M.; Matsuyama, T.; Yamaguchi, A.; Mitsuda, N.; Tsukamoto, Y.; Taniguchi, M.; Che, Y.H.; Ozawa, K.; Hori, O.; Nishimura, H.; et al. ORP150 protects against hypoxia/ischemia-induced neuronal death. *Nat. Med.* **2001**, *7*, 317–323. [[CrossRef](#)]
107. Bando, Y.; Katayama, T.; Kasai, K.; Taniguchi, M.; Tamatani, M.; Tohyama, M. GRP94 (94 kDa glucose-regulated protein) suppresses ischemic neuronal cell death against ischemia/reperfusion injury. *Eur. J. Neurosci.* **2003**, *18*, 829–840. [[CrossRef](#)]
108. Larner, S.F.; Hayes, R.L.; McKinsey, D.M.; Pike, B.R.; Wang, K.K. Increased expression and processing of caspase-12 after traumatic brain injury in rats. *J. Neurochem.* **2004**, *88*, 78–90. [[CrossRef](#)]
109. Tajiri, S.; Oyadomari, S.; Yano, S.; Morioka, M.; Gotoh, T.; Hamada, J.I.; Ushio, Y.; Mori, M. Ischemia-induced neuronal cell death is mediated by the endoplasmic reticulum stress pathway involving CHOP. *Cell Death Differ.* **2004**, *11*, 403–415. [[CrossRef](#)]
110. Hayashi, T.; Saito, A.; Okuno, S.; Ferrand-Drake, M.; Dodd, R.L.; Chan, P.H. Damage to the endoplasmic reticulum and activation of apoptotic machinery by oxidative stress in ischemic neurons. *J. Cereb. Blood Flow Metab.* **2005**, *25*, 41–53. [[CrossRef](#)]
111. Bonnefont-Rousselot, D.; Collin, F.; Jore, D.; Gardès-Albert, M. Reaction mechanism of melatonin oxidation by reactive oxygen species in vitro. *J. Pineal Res.* **2011**, *50*, 328–335. [[CrossRef](#)] [[PubMed](#)]
112. Galano, A.; Tan, D.X.; Reiter, R.J. Melatonin as a natural ally against oxidative stress: A physicochemical examination. *J. Pineal Res.* **2011**, *51*, 1–16. [[CrossRef](#)] [[PubMed](#)]
113. Kim, S.H.; Lee, S.M. Cytoprotective effects of melatonin against necrosis and apoptosis induced by ischemia/reperfusion injury in rat liver. *J. Pineal Res.* **2008**, *44*, 165–171. [[CrossRef](#)] [[PubMed](#)]
114. Muñoz-Casares, F.C.; Padillo, F.J.; Briceño, J.; Collado, J.A.; Muñoz-Castañeda, J.R.; Ortega, R.; Cruz, A.; Túnez, I.; Montilla, P.; Pera, C.; et al. Melatonin reduces apoptosis and necrosis induced by ischemia/reperfusion injury of the pancreas. *J. Pineal Res.* **2006**, *40*, 195–203. [[CrossRef](#)] [[PubMed](#)]
115. Tuñón, M.J.; San-Miguel, B.; Crespo, I.; Laliena, A.; Vallejo, D.; Álvarez, M.; Prieto, J.; González-Gallego, J. Melatonin treatment reduces endoplasmic reticulum stress and modulates the unfolded protein response in rabbits with lethal fulminant hepatitis of viral origin. *J. Pineal Res.* **2013**, *55*, 221–228. [[CrossRef](#)] [[PubMed](#)]
116. San-Miguel, B.; Crespo, I.; Vallejo, D.; Álvarez, M.; Prieto, J.; González-Gallego, J.; Tuñón, M.J. Melatonin modulates the autophagic response in acute liver failure induced by the rabbit hemorrhagic disease virus. *J. Pineal Res.* **2014**, *56*, 313–321. [[CrossRef](#)]
117. Fernández, A.; Ordóñez, R.; Reiter, R.J.; González-Gallego, J.; Mauriz, J.L. Melatonin and endoplasmic reticulum stress: Relation to autophagy and apoptosis. *J. Pineal Res.* **2015**, *59*, 292–307. [[CrossRef](#)]
118. Motilva, V.; García-Mauriño, S.; Talero, E.; Illanes, M. New paradigms in chronic intestinal inflammation and colon cancer: Role of melatonin. *J. Pineal Res.* **2011**, *51*, 44–60. [[CrossRef](#)] [[PubMed](#)]
119. Naziroğlu, M.; Tokat, S.; Demirci, S. Role of melatonin on electromagnetic radiation-induced oxidative stress and Ca^{2+} signaling molecular pathways in breast cancer. *J. Recept. Signal Transduct.* **2012**, *32*, 290–297. [[CrossRef](#)]
120. Hosseinzadeh, A.; Kamrava, S.K.; Joghataei, M.T.; Darabi, R.; Shakeri-Zadeh, A.; Shahriari, M.; Reiter, R.J.; Ghaznavi, H.; Mehrzadi, S. Apoptosis signaling pathways in osteoarthritis and possible protective role of melatonin. *J. Pineal Res.* **2016**, *61*, 411–425. [[CrossRef](#)]
121. Mortezaee, K. Human hepatocellular carcinoma: Protection by melatonin. *J. Cell. Physiol.* **2018**, *233*, 6486–6508. [[CrossRef](#)] [[PubMed](#)]
122. Yang, Y.; Cheung, H.H.; Zhang, C.; Wu, J.; Chan, W.Y. Melatonin as Potential Targets for Delaying Ovarian Aging. *Curr. Drug Targets* **2019**, *20*, 16–28. [[CrossRef](#)] [[PubMed](#)]
123. Fu, Z.; Jiao, Y.; Wang, J.; Zhang, Y.; Shen, M.; Reiter, R.J.; Xi, Q.; Chen, Y. Cardioprotective Role of Melatonin in Acute Myocardial Infarction. *Front. Physiol.* **2020**, *11*, 366. [[CrossRef](#)]

124. Huang, K.; Luo, X.; Zhong, Y.; Deng, L.; Feng, J. New insights into the role of melatonin in diabetic cardiomyopathy. *Pharmacol. Res. Perspect.* **2022**, *10*, e00904. [[CrossRef](#)]
125. Carloni, S.; Albertini, M.C.; Galluzzi, L.; Buonocore, G.; Proietti, F.; Balduini, W. Melatonin reduces endoplasmic reticulum stress and preserves sirtuin 1 expression in neuronal cells of newborn rats after hypoxia-ischemia. *J. Pineal Res.* **2014**, *57*, 192–199. [[CrossRef](#)]
126. Wongprayoon, P.; Govitrapong, P. Melatonin Protects SH-SY5Y Neuronal Cells Against Methamphetamine-Induced Endoplasmic Reticulum Stress and Apoptotic Cell Death. *Neurotox. Res.* **2017**, *31*, 1–10. [[CrossRef](#)]
127. Tungkom, W.; Jumnonprakon, P.; Tocharus, C.; Govitrapong, P.; Tocharus, J. Melatonin suppresses methamphetamine-triggered endoplasmic reticulum stress in C6 cells glioma cell lines. *J. Toxicol. Sci.* **2017**, *42*, 63–71. [[CrossRef](#)] [[PubMed](#)]
128. Feng, D.; Wang, B.; Wang, L.; Abraham, N.; Tao, K.; Huang, L.; Shi, W.; Dong, Y.; Qu, Y. Pre-ischemia melatonin treatment alleviated acute neuronal injury after ischemic stroke by inhibiting endoplasmic reticulum stress-dependent autophagy via PERK and IRE1 signalings. *J. Pineal Res.* **2017**, *62*, e12395. [[CrossRef](#)]
129. Song, J.; Kim, O.Y. Melatonin Modulates Neuronal Cell Death Induced by Endoplasmic Reticulum Stress under Insulin Resistance Condition. *Nutrients* **2017**, *9*, 593. [[CrossRef](#)]
130. Xue, F.; Shi, C.; Chen, Q.; Hang, W.; Xia, L.; Wu, Y.; Tao, S.Z.; Zhou, J.; Shi, A.; Chen, J. Melatonin Mediates Protective Effects against Kainic Acid-Induced Neuronal Death through Safeguarding ER Stress and Mitochondrial Disturbance. *Front. Mol. Neurosci.* **2017**, *10*, 49. [[CrossRef](#)] [[PubMed](#)]
131. Shi, C.; Zeng, J.; Li, Z.; Chen, Q.; Hang, W.; Xia, L.; Wu, Y.; Chen, J.; Shi, A. Melatonin Mitigates Kainic Acid-Induced Neuronal Tau Hyperphosphorylation and Memory Deficits through Alleviating ER Stress. *Front. Mol. Neurosci.* **2018**, *11*, 5. [[CrossRef](#)] [[PubMed](#)]
132. Lin, Y.W.; Chen, T.Y.; Hung, C.Y.; Tai, S.H.; Huang, S.Y.; Chang, C.C.; Hung, H.Y.; Lee, E.J. Melatonin protects brain against ischemia/reperfusion injury by attenuating endoplasmic reticulum stress. *Int. J. Mol. Med.* **2018**, *42*, 182–192. [[CrossRef](#)] [[PubMed](#)]
133. Xu, W.; Lu, X.; Zheng, J.; Li, T.; Gao, L.; Lenahan, C.; Shao, A.; Zhang, J.; Yu, J. Melatonin Protects Against Neuronal Apoptosis via Suppression of the ATF6/CHOP Pathway in a Rat Model of Intracerebral Hemorrhage. *Front. Neurosci.* **2018**, *12*, 638. [[CrossRef](#)]
134. Thangwong, P.; Jearjaroen, P.; Govitrapong, P.; Tocharus, C.; Tocharus, J. Melatonin improves cognitive function by suppressing endoplasmic reticulum stress and promoting synaptic plasticity during chronic cerebral hypoperfusion in rats. *Biochem. Pharmacol.* **2022**, *198*, 114980. [[CrossRef](#)] [[PubMed](#)]
135. Ling, Z.Q.; Tian, Q.; Wang, L.; Fu, Z.Q.; Wang, X.C.; Wang, Q.; Wang, J.Z. Constant illumination induces Alzheimer-like damages with endoplasmic reticulum involvement and the protection of melatonin. *J. Alzheimers Dis.* **2009**, *16*, 287–300. [[CrossRef](#)]
136. Lee, J.H.; Yoon, Y.M.; Han, Y.S.; Jung, S.K.; Lee, S.H. Melatonin protects mesenchymal stem cells from autophagy-mediated death under ischaemic ER-stress conditions by increasing prion protein expression. *Cell Prolif.* **2019**, *52*, e12545. [[CrossRef](#)]
137. Sharma, R.; Reiter, R.J.; Ma, Q. Melatonin: A hypothesis regarding its use to treat Wilson disease. *Med. Hypotheses* **2019**, *133*, 109408. [[CrossRef](#)]
138. Promyo, K.; Iqbal, F.; Chaidee, N.; Chetsawang, B. Aluminum chloride-induced amyloid β accumulation and endoplasmic reticulum stress in rat brain are averted by melatonin. *Food Chem. Toxicol.* **2020**, *146*, 111829. [[CrossRef](#)]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.