

Document downloaded from:

<http://hdl.handle.net/10251/70887>

This paper must be cited as:

García Pérez, JV.; Mulet Pons, A.; Carcel Carrión, JA. (2011). Ultrasound-assisted extraction of natural products. *Food Engineering Reviews*. 3:108-120. doi:10.1007/s12393-011-9036-6.



The final publication is available at

<https://dx.doi.org/10.1007/s12393-011-9036-6>

Copyright Springer Verlag (Germany)

Additional Information

# Ultrasonic assisted extraction of natural products

<sup>1</sup>Esclapez, M.D., <sup>2</sup>García-Pérez, J.V., <sup>2</sup>Mulet, A. and <sup>2</sup>Cárcel J.A.

<sup>1</sup>*Departamento de Química Física e Instituto Universitario de Electroquímica, Universidad de Alicante, Ap. Correos 99, 03080 Alicante, Spain*

<sup>2</sup>*Grupo de Análisis y Simulación de Procesos Agroalimentarios, Departamento de Tecnología de Alimentos, Universidad Politécnica de Valencia, Camí de Vera s/n, E46022, Valencia, Spain*

Ultrasound assisted extraction (USAE) is an interesting process to obtain high valuable compounds and could contribute to the increase the value of some food by-products when used as sources of natural compounds. The main benefits will be a more effective extraction, thus saving energy, and also to the use of moderate temperatures, which is beneficial for heat sensitive compounds. For a successful application of the USAE, it is necessary to consider the influence of several process variables, the main ones being the applied ultrasonic power, the frequency, the extraction temperature, the reactor characteristics and the solvent-sample interaction. The highest extraction rate is usually achieved in the first few minutes, which is the most profitable period. To optimize the process, rate equations and unambiguous process characterization are needed, aspects that have often been lacking.

*Key words: ultrasound, mass transfer, food by-product, quality*

Abbreviations:

$\Pi$  : ultrasonic power density

f : applied frequency

USAE: ultrasound assisted extraction

## 1. Introduction.

In the last few years, ultrasound technology has been revealed as valuable tool in food engineering processes (Bhaskaracharya et al. 2009), and this field of research has become a very active one. Two very different approaches to the application of ultrasound have been explored mainly related to the frequency and the power of the ultrasonic vibration. One of them consists of using high frequency-low intensity ultrasound and it has specifically focused on the quality monitoring of products or processes, such as the assessment of the cheese maturation time (Benedito et al. 2000) or compositional analysis (Benedito et al. 2001). The applications are mainly related to the measurement of ultrasound velocity, the attenuation of the signal or the analysis of frequency spectra. In these applications there is no influence on the process or the product.

1 Another approach, the low frequency-high power ultrasound, has been  
2 successfully applied to improve the process and/or the products. This was the case for  
3 brining processes (Cárcel et al. 2007a), maceration (Cárcel et al. 2007b), microbial and  
4 enzyme inactivation (Vercet et al. 2001), freezing processes (Zheng and Sun 2006), the  
5 modification of ingredients (Kardos and Luche 2001), the extraction of natural products  
6 (Vilkhu et al. 2008), food cutting operations (Arnold et al. 2009), improving the quality of  
7 emulsions (Wulff-Pérez et al. 2009) or slurry preparation (Montalbo-Lomboy et al. 2010),  
8 sugar substitution processes (Garcia-Noguera et al. 2010) and fermentations processes  
9 (Riener et al. 2010) among others.

14 Ultrasound is usually applied in solid/fluid media, being the fluid either gas or  
15 liquid. The applications in solid/gas systems are not frequent because the impedance  
16 mismatch and the air absorption make the transmission of ultrasound difficult. However,  
17 advances in transducer technology are allowing a more thorough understanding of power  
18 ultrasound assisted drying. The use of ultrasonic systems both with contact or not contact  
19 between samples and vibrating elements has been reported such as in vibrating  
20 chambers. In these applications, the power ultrasound increases the surface moisture  
21 evaporation rate and causes oscillating velocities and microstreaming at the interfaces  
22 which may affect the diffusion boundary layer and generate rapid series of alternative  
23 contractions and expansions of the material, affecting mass transfer (García-Pérez et al.  
24 2006).

31 In solid/liquid systems, the applications are more common. The enhancement of  
32 the mass transfer brought about by acoustic induced cavitation in a liquid medium is one  
33 of the beneficial effects. When mechanical waves are transmitted through a fluid, the  
34 average distance within molecules is modified, oscillating around their equilibrium  
35 position. During the compression cycle, the intermolecular distance shortens and  
36 lengthens again in the rarefaction cycle. When the pressure decrease in the rarefaction  
37 cycle is enough to exceed the critical distance between molecules, cavities can appear in  
38 the bulk liquid. Those incipient bubbles keep on growing until the system reaches its  
39 minimal pressure and the subsequent compression cycle starts. Then, the cavities can  
40 start another rarefaction cycle or collapse adiabatically resulting in a violent implosion at  
41 the end of a compression cycle, yielding shock waves of several hundred atmospheres  
42 and around 5000 K of temperature (Leighton 2007). Cavitation bubbles exist for few  
43 acoustic cycles before collapsing, giving rise to smaller bubbles which could act as new  
44 cavitation nuclei or simply get dissolved.

53 The mechanical effects induced by cavitation bubbles in the solid-liquid interface  
54 of a heterogeneous medium include micro jet impacts and shockwave induced damages.  
55 Thus, the potential energy of the expanded bubble is converted into kinetic energy of a  
56 liquid jet that extends inside the bubble and penetrates the opposite bubble wall, reaching  
57 velocities of hundreds of meters per second (Suslick 2001). Moreover, sonochemical  
58 processes led chemical reactions induced by the formation of radical species, mainly  $\cdot\text{OH}$ ,  
59  
60  
61  
62

1 from the scission of water when it acts as a solvent (Fischer et al. 1986). The kind of  
2 radicals formed depends on the kind of gas dissolved (Adewuyi 2001). Furthermore,  
3 cavitation modifies the chemical processes in the system, mainly enhancing the reaction  
4 rates of existing processes or starting new reaction mechanisms, by the formation of  
5 reactive radical species (Ince et al. 2001). Those statements could suggest dramatic  
6 changes in the parameters as temperature or the pressure of the bulk surrounding but  
7 this is not the case because the time scale for these micro-reactors is really small to  
8 affect the macroscopic system. However, they could affect cellular structures and  
9 enhance mass transport processes, like extraction.  
10  
11  
12

13  
14 From the effects so far mentioned, it appears that the use of ultrasound could be  
15 a promising way to introduce innovation into extraction processes.  
16

## 17 **2. Extraction processes of natural products.**

18  
19 In food engineering, the extraction of natural compounds appears to be of quite  
20 considerable interest mainly due to the properties of some natural extracts. Moreover, the  
21 extraction of bioactive compounds can increase the value of some food industry by-  
22 products, one example being the extraction of flavonoids from citrus peel that could be  
23 further exploited by both the pharmaceutical and food industries (Sun et al. 2011),  
24 another being the obtainment of natural extracts from marine sources (Ferraro et al.  
25 2010), from the residues of the distillation of essential oils (Velickovic et al. 2008) or from  
26 apple pomace (Virost et al. 2010).  
27  
28  
29  
30  
31  
32  
33

34 The extraction processes have been mainly focused on vegetal compounds.  
35 Plants contain a broad range of bioactive compounds such as lipids, phytochemicals,  
36 flavours, fragrances or pigments that are widely used in the food, pharmaceutical and  
37 cosmetic industries. The interest in those compounds has led to an increased need for  
38 better extraction methods, which could obtain the greatest possible amount of bioactive  
39 constituents in a shorter processing time and at a low cost. The extraction of antioxidants  
40 such as anthocyanins, flavonols, or phenolic acids has been specially addressed. The  
41 properties exhibited by polyphenols are of interest in the production of functional foods  
42 increasing their antioxidant activity and enhancing their biological benefits (Liazid et al.  
43 2010). The extraction has also focused on natural additives, such as flavour from soy  
44 (Yang et al. 2011) or capsaicinoids (Barbero et al. 2008) and colorants (Sivakumar et al.  
45 2009). Furthermore, the extraction can also be used for the elimination of undesirable  
46 components in foods, for example, some aroma compounds (Abad Romero et al. 2010).  
47  
48  
49  
50  
51  
52  
53

54 There are many extraction techniques, such as maceration, digestion, infusion,  
55 decoction, percolation, hot continuous extraction, counter-current extraction, supercritical  
56 fluid extraction, microwave assisted extraction, phytonic processes or ultrasound assisted  
57 extraction (Handa et al. 2008). The main differences among these techniques are related  
58 to the design of the reactors, the solvents used, the time and temperature of the  
59  
60  
61  
62  
63  
64  
65

1 processes or the use of new technologies (microwave, ultrasound, supercritical fluids or  
2 the use of enzymes).

3 The conventional liquid–solid extraction techniques are laborious, time-  
4 consuming and often require large volumes of organic solvents (Wang and Weller 2006).  
5 Nowadays, the tendency is to use more environmentally friendly solvents such as  
6 alcohol-water mixtures, thus increasing the costs and lowering the extraction yield. Then,  
7 there is great interest in the development of efficient extraction methods.  
8  
9

10 The supercritical phase can be attained by increasing the pressure and  
11 temperature, reaching a state of aggregation at which no distinction between the gas and  
12 liquid can be observed (Starmans and Nijhuis 1996). The density of supercritical fluids is  
13 intermediate between gas and liquid and, therefore, its solubilising power is higher than  
14 gases. Their transport properties, as the lower viscosity and the higher diffusivity than  
15 liquid solvents, favor higher extraction yields by penetrating into porous solid materials  
16 more effectively than liquid solvents (Riera et al. 2004). The most common fluid used for  
17 supercritical fluid extractions is carbon dioxide, due to its low supercritical temperature  
18 and pressure and its quasi inert character (Sahena et al. 2009). Indeed, the extracted  
19 organic compounds are easily separated because carbon dioxide is a gas at atmospheric  
20 pressure and can, therefore, be recycled.  
21  
22  
23  
24  
25  
26  
27

28 Ultrasound assisted extraction (USAE) has been used extensively in the last two  
29 decades as an efficient extraction method both in the food and pharmaceutical industries.  
30 The number of papers published in the last three decades related with this topic has  
31 suffered an exponential increase (Figure 1). According to the applied power, different  
32 effects may be sought. At low intensities, the external and, possibly, the internal mass  
33 transfer resistances are affected. Nevertheless, the product structure remains mainly  
34 unaffected. Intermediate intensities may affect the product structure thus increasing the  
35 effects on the internal mass transfer resistance. If ultrasonic power is further increased,  
36 cell disintegration can take place.  
37  
38  
39  
40  
41  
42

43 Nowadays, it is a powerful tool in extraction processes, even being the reference  
44 point for some extractive technologies in development (Chena et al. 2009). USAE has  
45 been used, for example, in the extraction of polyphenols from vegetal tissues (Ma et al.  
46 2009), protein, sugar (Karki et al. 2010) and starch (Naguleswaran and Vasanthan 2010)  
47 from cereals and legumes, oil (Zhang et al. 2009), flavour compounds (Da Porto and  
48 Decorti 2009; Da Porto et al. 2009) from spearmint and lavender. Sonication improves the  
49 extraction efficiency and rate, reduces the temperature needed, allows solvents to be  
50 saved and favours the solubilisation of the targeted compounds (Luque de Castro et al.  
51 2007). The marked increase in the very local temperature enhances the solubility of the  
52 analytes (as in the case of lipids) in the solvent and eases their diffusion from the sample  
53 matrix to the outer region. On the other hand, the local pressure increase facilitates both  
54 the penetration of the solvent into the sample matrix and the transfer of compounds  
55 (Abad Romero et al. 2010). Moreover, the implosion of cavitation bubbles can hit the  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 surface of the solid matrix and disintegrate the cells (Figure 2) (Veillet et al. 2010). As a  
2 consequence of these effects, less extraction time is required than in conventional  
3 extraction processes. Thus, Roldán-Gutiérrez et al. (2008) showed that for the extraction  
4 of valuable compounds from aromatic plants and flowers, the USAE was faster than  
5 conventional steam distillation extraction in a range from 45 to 12 and from 9 to 2 times  
6 faster than supercritical water extraction. One of the main drawbacks of USAE, compared  
7 with carbon dioxide supercritical extraction, is the unavoidable use of organic solvents in  
8 some cases, but the equipment is simpler and the overall cost is lower (Abad Romero et  
9 al. 2010).

### 14 **3. Main variables to be considered in USAE.**

17 Besides innovation, the reliability and reproducibility of results, are among the  
18 main goals in any scalable process. In the case of USAE, it is necessary to consider the  
19 influence of process variables, such as ultrasonic power, frequency, temperature, reactor  
20 design, solvents, solvent-sample ratio or particle size and structure. Those variables may  
21 allow the identification of the scale factors which are necessary to be able to transfer the  
22 experimental conditions found in a laboratory to an industrial context. Moreover, it is  
23 necessary to consider that in extraction processes, the target of the process is not always  
24 to achieve the largest extraction yield (i.e. in maceration processes), but the lowest  
25 consumption of monetary and energy resources. Thus, optimization is an important step  
26 in the USAE processes. This has been extensively addressed in the literature, as can be  
27 seen in Table 1, where a collection of research work dealing with different USAE for  
28 natural compounds is summarized.

31 In addition to the previously mentioned variables, the pre-treatment, conservation  
32 and storage (Romdhan and Gourdon 2002) of the sample influence the extraction yield to  
33 a high extent and should also be carefully controlled. Domínguez et al. (1994) reviewed  
34 the beneficial effects of the enzymatic pre-treatment of fruits and oilseeds prior to  
35 conventional solvent extraction, concluding that shorter times can be achieved and the  
36 capacity of conventional equipment increased. In the same way, García-Pérez et al.  
37 (2010) pointed to the influence of the drying method used on grape stalk before the  
38 extraction of antioxidants. Thus, the extracts obtained from freeze dried samples  
39 presented the highest antioxidant capacity and, the extraction process involved, a higher  
40 effective diffusivity and mass transfer coefficient than conventionally dried samples.  
41 Extracts from hot air drying involved a lower antioxidant concentration and a slower  
42 extraction process.

#### 54 **3.1 Ultrasonic power.**

57 The measurement of the actual acoustic energy applied in a sonochemical  
58 process is quite difficult and is not very often reported. Sometimes, considering the  
59 different power levels of the device, authors show the values of power applied as, for  
60

1 example, “20 % of the total electric power capacity” and this is not an accurate  
2 measurement at all. In fact, in most of the ultrasound devices, the power measured is not  
3 proportional to the power step shown (Figure 3), leading to wrong conclusions or  
4 irreproducible results.  
5

6 Several methods can be found in literature for the estimation of the acoustic  
7 energy transferred by measuring the physical or chemical changes occurring in solution  
8 due to ultrasound. Among the physical methods the direct measurement of the acoustic  
9 pressure using hydrophones (Martin and Law 1983) or optical microscopes, the  
10 aluminium foil method (Chivate and Pandit 1995) or the calorimetric method (Margulis  
11 and Margulis 2003) can be highlighted. The application of chemical methods is based on  
12 the indirect estimation of the  $\cdot\text{OH}$  radical formed by cavitation using measurements of the  
13 sonoluminescence related to cavitation processes (Archley and Crum 1998) or the  
14 chemical dosimeters able to react with the radicals (Makino et al. 1982). The products  
15 formed (or reacting products) can be determined easily (i.e. by UV-Vis, fluorescence), as,  
16 for instance, in the case of luminol (Trabelsi et al. 1996), iodide (Entezari and Kruus  
17 1994), phenolphthaleine (Rong et al. 2008), terephthalate (Mason et al. 1994) or 4-  
18 nitrophenol (Kotronarou et al. 1991), among others. Even knowing the ultrasonic power  
19 actually applied, it is difficult to compare the effects because often, the results are not  
20 only reported on a different basis, but are also influenced by the geometry of the reactor.  
21 For instance, to report data indicating only the power applied is not enough. Indicating the  
22 power density ( $\Pi$ ;  $\text{W} / \text{cm}^3$ ) is more appropriate, especially for the case of ultrasonic  
23 baths, where the whole bath volume should be considered, but the bath volume is usually  
24 a missing data in research papers, making it difficult to achieve reproducible results. In  
25 this paper, the value of  $\Pi$  will be given if it is possible to calculate it from the reported  
26 data, assuming anyway, that there is no homogeneity in the pieces of work found on the  
27 measurement of the applied ultrasonic power.  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39

40 As previously stated, the effects of the applied power are different according to  
41 the applied power density. It is possible that the power density applied will not be enough  
42 to affect the external/internal resistance thus, a threshold will be observed. It should also  
43 be noticed that the power density is not the same for the whole volume, being the power  
44 density distribution in the stationary field an important factor to be considered. A different  
45 power distribution may be a source of disagreement among authors. A power density limit  
46 may also be observed once the cells have been disrupted and the ultrasonic effects will  
47 no longer increase. This could also be the case when it is only the external resistance  
48 that is affected. For instance, Cárcel et al. (2007b) observed an increase in solute gain  
49 and water losses for the osmotic treatment of apple in aqueous sucrose solutions at  $\Pi$ :  
50  $0.076 \text{ W} / \text{cm}^3$  (20 kHz), thus indicating a decrease of internal/external resistance due to  
51 ultrasound. On the other hand, when using  $\Pi$ :  $0.107 \text{ W} / \text{cm}^3$  (20 kHz) in meat brining  
52 processes, Cárcel et al. (2007a) found that the amount of salt in the meat increased, but  
53 so did the moisture. That points to a different influence of the ultrasound in the mass  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 transfer process. In this case, the microinjection of brine inside the meat could explain the  
2 observed increase in both salt and moisture content.

3 The extraction yield usually increases in line with the power intensity applied,  
4 sometimes linearly. This was observed by Zou et al. (2010) working on the extraction of  
5 melanin from *Auricularia auricula* applying ultrasonic power density from 3.33 to 8.33 W /  
6 cm<sup>3</sup> (frequency: 40 kHz; solvent: water pH 12; extraction temperature: 60 °C; extraction  
7 time: 30 min; ratio raw material/solvent: 1g / 30 mL). In the case of the USAE of  
8 geniposide from gardenia fruit (frequency: 18 kHz; water as solvent; extraction  
9 temperature: 20 °C; extraction time: 160 min; ratio raw material/solvent: 1g / 40 mL) Ji et  
10 al. (2006) demonstrated that stirring had no effect on the total extraction, but ultrasound  
11 intensified both the mass transfer in the liquid film around fruit particles and intraparticle  
12 diffusion. The external mass transfer coefficient and the intraparticle diffusion coefficient  
13 increased in line with an increase in ultrasonic intensity. Stirring could intensify the  
14 external mass transfer, but had no effect on the intraparticle diffusion.

15 However, some systems could present a power plateau, as the results obtained  
16 by Zhao et al. (2007) indicate. In this work, it was observed that for the extraction of  
17 saikosaponins from *Radix Bupleuri* (frequency: 20 kHz; solvent composition: 70/30 v/v  
18 ethanol-water; extraction temperature: 80 °C; extraction time: 30 min; ratio raw  
19 material/solvent: 1g / 25 mL), at  $\Pi$  of above 0.28 W / cm<sup>3</sup>, no increase of the yield of  
20 extraction was found. Less usual was the effect found by Ma et al. (2008) for the  
21 extraction yields of hesperidin from Penggan peels (frequency: 100, 20 and 40 kHz;  
22 methanol as solvent; extraction temperature: 40 °C; extraction time: 5 min; ratio raw  
23 material/solvent: 1g / 40 mL) where the applied ultrasound power had only a slight effect  
24 on the yield of hesperidin, achieving similar results for the different acoustic power  
25 intensities applied (3.2, 8, 30 and 56 W). These results point to the existence of  
26 thresholds depending on the material treated.

27 In some cases, the proper use of the pulse mode of ultrasound can replace  
28 continuous irradiation by ultrasound in order to obtain better extraction yields, as in the  
29 case of the extraction of colorants from beetroot (Sivakumar et al, 2009), or to reduce the  
30 electrical energy consumption (Sun et al. 2011).

31 The applied power not only greatly influences the total amount of compounds  
32 extracted, but can also affect the proportion of species extracted because there is an  
33 effect on the extraction rate. In this sense, Wei et al. (2010) (frequency not shown; water  
34 as solvent; extraction temperature: 25 °C; 50 g / 400 mL; 5 min followed re-sonication in  
35 300 mL, 5 min), observed that the percentage of neutral polysaccharides obtained from  
36 tea flower decreased while increasing the applied power, rising from 100 to 300 W, but no  
37 influence was found on the total amount of polysaccharides or the acidic ones.

38 Therefore, considering all the parameters mentioned, it is apparent that the applied  
39 ultrasonic power can have a great influence on the extraction, not only increasing the  
40 extraction yield at shorter times, but also modifying, in some cases, the proportion of final



1 products. Researchers/processors should carefully evaluate the kind of characterization  
2 control that should be carried out in their system in order to achieve reproducible results  
3 in different devices for every single extraction problem, and check the ultrasonic power  
4 applied.  
5

### 6 **3.2 Ultrasonic frequency.**

7  
8  
9 It is known that the lower the frequency, the larger the cavitation bubble. So, the  
10 lower frequencies of high power ultrasound (around 20 kHz) achieve more violent bubble  
11 implosions, and consequently, are more efficient in extraction processes. As far as the  
12 extractive system is a heterogeneous and complex porous media, the effect the bubble  
13 size has on the efficiency of the extraction should be addressed. The frequency effect  
14 may be linked not only to the cavitation bubble size, but also to its influence on the  
15 external and internal resistances to mass transfer. Although the subject has not been  
16 addressed in extraction studies, it has been found in drying. In that process the solid  
17 structure/porosity plays a key role on the effects derived from ultrasonic application.  
18  
19

20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65  
There are few comparative studies related to the frequency effects in extraction  
processes, but Ma et al. (2008) observed that for the same applied power, the best  
frequency for the extraction of hesperidin from peengan peel was 60 kHz, instead of 20 or  
100 kHz. Dong et al. (2010) observed the extraction yields of salvianolic acid B from  
*Salvia miltiorrhiza* root (solvent composition: 60/40 v/v ethanol-water; extraction  
temperature: 30°C; ratio raw material/solvent: 1g / 20 ml; extraction time: 25 min; power  
applied: 100 W) under different ultrasound frequencies (28, 45 and 100 kHz), in a  
cleaning bath. The extraction yield under the highest frequency was lower than those  
under lower frequencies. So, there could exist an optimal frequency for the extraction  
related to the specific textural properties of any complex matrix.

### 66 **2.3. Temperature.**

67  
68  
69  
70  
71  
72  
73  
74  
75  
76  
77  
78  
79  
80  
81  
82  
83  
84  
85  
86  
87  
88  
89  
90  
91  
92  
93  
94  
95  
96  
97  
98  
99  
100  
101  
102  
103  
104  
105  
106  
107  
108  
109  
110  
111  
112  
113  
114  
115  
116  
117  
118  
119  
120  
121  
122  
123  
124  
125  
126  
127  
128  
129  
130  
131  
132  
133  
134  
135  
136  
137  
138  
139  
140  
141  
142  
143  
144  
145  
146  
147  
148  
149  
150  
151  
152  
153  
154  
155  
156  
157  
158  
159  
160  
161  
162  
163  
164  
165  
166  
167  
168  
169  
170  
171  
172  
173  
174  
175  
176  
177  
178  
179  
180  
181  
182  
183  
184  
185  
186  
187  
188  
189  
190  
191  
192  
193  
194  
195  
196  
197  
198  
199  
200  
201  
202  
203  
204  
205  
206  
207  
208  
209  
210  
211  
212  
213  
214  
215  
216  
217  
218  
219  
220  
221  
222  
223  
224  
225  
226  
227  
228  
229  
230  
231  
232  
233  
234  
235  
236  
237  
238  
239  
240  
241  
242  
243  
244  
245  
246  
247  
248  
249  
250  
251  
252  
253  
254  
255  
256  
257  
258  
259  
260  
261  
262  
263  
264  
265  
266  
267  
268  
269  
270  
271  
272  
273  
274  
275  
276  
277  
278  
279  
280  
281  
282  
283  
284  
285  
286  
287  
288  
289  
290  
291  
292  
293  
294  
295  
296  
297  
298  
299  
300  
301  
302  
303  
304  
305  
306  
307  
308  
309  
310  
311  
312  
313  
314  
315  
316  
317  
318  
319  
320  
321  
322  
323  
324  
325  
326  
327  
328  
329  
330  
331  
332  
333  
334  
335  
336  
337  
338  
339  
340  
341  
342  
343  
344  
345  
346  
347  
348  
349  
350  
351  
352  
353  
354  
355  
356  
357  
358  
359  
360  
361  
362  
363  
364  
365  
366  
367  
368  
369  
370  
371  
372  
373  
374  
375  
376  
377  
378  
379  
380  
381  
382  
383  
384  
385  
386  
387  
388  
389  
390  
391  
392  
393  
394  
395  
396  
397  
398  
399  
400  
401  
402  
403  
404  
405  
406  
407  
408  
409  
410  
411  
412  
413  
414  
415  
416  
417  
418  
419  
420  
421  
422  
423  
424  
425  
426  
427  
428  
429  
430  
431  
432  
433  
434  
435  
436  
437  
438  
439  
440  
441  
442  
443  
444  
445  
446  
447  
448  
449  
450  
451  
452  
453  
454  
455  
456  
457  
458  
459  
460  
461  
462  
463  
464  
465  
466  
467  
468  
469  
470  
471  
472  
473  
474  
475  
476  
477  
478  
479  
480  
481  
482  
483  
484  
485  
486  
487  
488  
489  
490  
491  
492  
493  
494  
495  
496  
497  
498  
499  
500  
501  
502  
503  
504  
505  
506  
507  
508  
509  
510  
511  
512  
513  
514  
515  
516  
517  
518  
519  
520  
521  
522  
523  
524  
525  
526  
527  
528  
529  
530  
531  
532  
533  
534  
535  
536  
537  
538  
539  
540  
541  
542  
543  
544  
545  
546  
547  
548  
549  
550  
551  
552  
553  
554  
555  
556  
557  
558  
559  
560  
561  
562  
563  
564  
565  
566  
567  
568  
569  
570  
571  
572  
573  
574  
575  
576  
577  
578  
579  
580  
581  
582  
583  
584  
585  
586  
587  
588  
589  
590  
591  
592  
593  
594  
595  
596  
597  
598  
599  
600  
601  
602  
603  
604  
605  
606  
607  
608  
609  
610  
611  
612  
613  
614  
615  
616  
617  
618  
619  
620  
621  
622  
623  
624  
625  
626  
627  
628  
629  
630  
631  
632  
633  
634  
635  
636  
637  
638  
639  
640  
641  
642  
643  
644  
645  
646  
647  
648  
649  
650  
651  
652  
653  
654  
655  
656  
657  
658  
659  
660  
661  
662  
663  
664  
665  
666  
667  
668  
669  
670  
671  
672  
673  
674  
675  
676  
677  
678  
679  
680  
681  
682  
683  
684  
685  
686  
687  
688  
689  
690  
691  
692  
693  
694  
695  
696  
697  
698  
699  
700  
701  
702  
703  
704  
705  
706  
707  
708  
709  
710  
711  
712  
713  
714  
715  
716  
717  
718  
719  
720  
721  
722  
723  
724  
725  
726  
727  
728  
729  
730  
731  
732  
733  
734  
735  
736  
737  
738  
739  
740  
741  
742  
743  
744  
745  
746  
747  
748  
749  
750  
751  
752  
753  
754  
755  
756  
757  
758  
759  
760  
761  
762  
763  
764  
765  
766  
767  
768  
769  
770  
771  
772  
773  
774  
775  
776  
777  
778  
779  
780  
781  
782  
783  
784  
785  
786  
787  
788  
789  
790  
791  
792  
793  
794  
795  
796  
797  
798  
799  
800  
801  
802  
803  
804  
805  
806  
807  
808  
809  
810  
811  
812  
813  
814  
815  
816  
817  
818  
819  
820  
821  
822  
823  
824  
825  
826  
827  
828  
829  
830  
831  
832  
833  
834  
835  
836  
837  
838  
839  
840  
841  
842  
843  
844  
845  
846  
847  
848  
849  
850  
851  
852  
853  
854  
855  
856  
857  
858  
859  
860  
861  
862  
863  
864  
865  
866  
867  
868  
869  
870  
871  
872  
873  
874  
875  
876  
877  
878  
879  
880  
881  
882  
883  
884  
885  
886  
887  
888  
889  
890  
891  
892  
893  
894  
895  
896  
897  
898  
899  
900  
901  
902  
903  
904  
905  
906  
907  
908  
909  
910  
911  
912  
913  
914  
915  
916  
917  
918  
919  
920  
921  
922  
923  
924  
925  
926  
927  
928  
929  
930  
931  
932  
933  
934  
935  
936  
937  
938  
939  
940  
941  
942  
943  
944  
945  
946  
947  
948  
949  
950  
951  
952  
953  
954  
955  
956  
957  
958  
959  
960  
961  
962  
963  
964  
965  
966  
967  
968  
969  
970  
971  
972  
973  
974  
975  
976  
977  
978  
979  
980  
981  
982  
983  
984  
985  
986  
987  
988  
989  
990  
991  
992  
993  
994  
995  
996  
997  
998  
999  
1000

In general, the increase of the extraction temperature enhances the mass transfer. Nevertheless, the temperature increase produces less cavitation energy conversion due to the decrease in surface tension and the increase in vapour pressure

1 (Kuijpers et al. 2002). However, usually, the observed increase in the extraction yield may  
2 be linked to the increase of the solvent diffusivity into the cells and the enhancement of  
3 desorption and solubility. Hemwimol et al. (2006) observed this behaviour in the USAE  
4 extraction of anthraquinones from roots of *Morinda citrifolia* (frequency: 38.5 kHz; solvent:  
5 ethanol; extraction time: 90 min; ratio raw material/solvent: 0.1 g / 10 mL;  $\Pi$ : 0.007 W /  
6  $\text{cm}^3$ ). At 45°C and 60 °C, the initial extraction rate was so high that the extraction yield  
7 remained almost constant after 30 minutes of USAE. This yield value could not be  
8 reached when working at 25 °C, even in USAE periods of 90 minutes, and a slow  
9 extraction rate was evidenced.

10 It is also common to observe a decrease in the yield as the temperature rises,  
11 especially in the case of unstable or volatile compounds. During the USAE of epimedin C  
12 from fresh leaves of *Epimedium* (frequency: 24 kHz; solvent: methanol; extraction time:  
13 25 min; ratio raw material/solvent: 1g / 30 mL;  $\Pi$ : 0.44 W /  $\text{cm}^3$ ) the extraction yield rises  
14 as the extraction temperature goes up from 15 to 45 °C, at this temperature reaching the  
15 peak value (Zhang et al. 2009). However, when the extraction temperature was above  
16 this threshold temperature (45 °C), the extraction yield decreased. Furthermore, the use  
17 of higher temperatures above the threshold temperature might result in the acceleration  
18 of solvent volatilization, higher energy costs and the enhancement of the extraction of  
19 impurities.

20 As it could be infer from the previous comments, the optimum values of the  
21 operational variables are interrelated. Therefore the higher boiling point of some solvents  
22 permits the use of higher temperatures (limited by the product stability) and so, a greater  
23 efficiency could be achieved. Thus, an optimum extraction temperature value exists at  
24 which the extraction yield can reach its maximum level, and this temperature value may  
25 be different if other working parameters change, i.e. the applied power, the kind or the  
26 amount of solvent. Thus, a multivariate optimization of the process is recommended.

### 39 **3.4 Reactor characteristics.**

40 The design of the reactor is an important factor to be taken into account. Thus, in  
41 the early years of sonochemistry, experiments were performed in open flasks immersed  
42 in ultrasonic cleaning baths and it could be observed how important the accuracy of the  
43 geometry of the reactor is for the reproducibility of the results (Weissler et al. 1950).  
44 Advances in the construction of acoustic generators allowed the use of transducers that  
45 were dipped directly in the bulk solution. This technique allowed higher energy to be  
46 transmitted into the medium, but it increases the chance that the samples may be  
47 contaminated with undesired materials (mainly from the degradation of the ultrasonic  
48 probes). In this approach, the study of the chemical and physical compatibility of all the  
49 materials and chemicals involved is mandatory. Further advances have been made by  
50 taking into account the lack of homogeneity of the pressure field in the reactor in order to  
51 optimize the process efficiency. It is necessary to calculate the optimum reactor  
52 dimensions and the position of the elements inside in relation to the transducer to attain

1 the maximum energy transferred to the fluid (Esclapez et al. 2010). For that purpose, it is  
2 necessary to test different dimensions of the reactor and the relative position of the  
3 transducers. Mapping techniques that allow the identification of the position where  
4 cavitation effects are at their maximum can be used (Kanthale et al. 2003).  
5

6 Some authors have optimized their designs by taking into account the effect of tip  
7 immersion in extraction flasks, obtaining substantial differences in the results. During the  
8 extraction of all-trans- $\beta$ -carotene from citrus peels, Sun et al. (2011) observed that the  
9 extraction yield decreased significantly with an increase in the liquid height. For this  
10 purpose, an ultrasonic tip was immersed 1 mm in a tube containing the sample liquid (3  
11 cm diameter, 10-20 cm of height). Under USAE, the extraction yield at heights of 2 cm  
12 and 12 cm were, respectively, 84.5% and 12.3% higher than the extraction yield for  
13 conventional extraction (2.51  $\mu\text{g} / \text{g}$ ). However, for those kinds of experiments, the power  
14 density should remain constant for valid results just related to geometry, maintaining the  
15 applied power/volume ratio, changing the geometry of the flask, or alternatively, the total  
16 applied power. The same happens in ultrasonic baths, where it is known that the acoustic  
17 stationary field is inhomogeneous and affected by the liquid height (Mulet et al. 2003).  
18

19 Nowadays, the most widely used extraction systems on laboratory scale are  
20 ultrasonic baths or probes working in batch or in flow mode (Figure 4). Efforts have been  
21 made to systematically characterize sonoreactors (Sáez et al. 2005a; Sáez et al. 2005b),  
22 even developing several numerical simulation models for the behaviour of systems with  
23 different geometries (Louisnard et al. 2009). Some research groups use continuous,  
24 specifically designed, devices for leaching with an optimized alternating solvent flow  
25 direction like the one used by Roldán-Gutiérrez et al. (2008) for the extraction of  
26 polyphenols from vegetal tissues. In that case, the sample was placed in a leaching  
27 chamber connected to the manifold flow and immersed in a transmitting fluid where the  
28 ultrasonic probe is dipped. Other groups have adapted this same idea, but by immersing  
29 the cell in an ultrasonic bath (Lou et al. 2010). That approach has focused mainly on the  
30 improvement of analytical procedures, but is an interesting first step for the scale-up. For  
31 the next few years expect to witness further research in this area, bearing in mind that the  
32 interaction between the pressure map inside a reactor and a coupled hydrodynamic flow  
33 is now under research with the aim of building more efficient sonoreactors (Esclapez et  
34 al. 2010). In any case, the reactors must be carefully characterized in order to provide  
35 useful and reproducible results.  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49

### 50 **3.5 Solvent-sample interaction.**

51 Careful consideration should be given to the choice of an appropriate solvent  
52 because the optimum solvent for traditional extractions may not be the best for USAE.  
53 Solvent selection is usually based on achieving high molecular affinity between the  
54 solvent and solute, but factors affecting cavitation, such as solvent vapour pressure or  
55 surface tension, need to be considered as well (Li et al. 2004). At high vapour pressures,  
56 the volatilized solvent will penetrate the cavitation bubbles more easily, cushioning their  
57  
58  
59  
60  
61  
62  
63  
64  
65

collapse (Price 1990). In this sense, cheaper solvents such as water mixtures could work better than traditional volatile pure solvents used for conventional extraction.

For a fixed amount of solid matrix, the more solvent used, the greater concentration gradient effect is achieved between the vegetal issue and the solvent, thus obtaining a faster extraction rate (Stanisavljevic et al 2007). However, for large solvent/solid ratios the improvement in extraction is negligible because the increase in the gradient is also negligible (Yang and Zhang 2008) and more time and energy consumption are needed for post-treatment concentration. Nevertheless, Barbero et al. (2008) found no influence of the solvent/solid ratio in the USAE of capsaicinoids from pepper (frequency: not shown; extraction temperature: 50°C; solvent: methanol; extraction time: 10 min; raw material: 1 g; power applied: 360 W in ultrasonic bath). For that purpose, extractions were carried out with different volumes (15, 25, 40 and 50 mL). No statistical differences in the extraction yield were found for the extraction volumes tested. Those results may be linked to fast extraction kinetics or long observation times that did not permit distinguish among treatments. Nevertheless, in this case, the lower solvent ratio should be chosen.

The extraction yield can be influenced by the particle size too, because extraction is linked to the surface exposed and the length of the path that the molecules should move along. In the extraction of oil from tobacco seeds (frequency: 40 kHz; extraction temperature: 25 °C; solvents: hexane or petroleum ether; extraction time: 60 min; raw material/solvent volume ratio: 1g / 3 mL; power applied: 150 W in an ultrasonic bath), it was observed that the highest extraction yield was achieved for particle size 0.4 - 0.5 mm (Stanisavljevic et al. 2007). The oil yield from the largest particles (>0.5 mm) class was lower than that from the small ones for the same period of time. The authors concluded that a smaller amount of oil could be transported from the interior of larger seed particles to the bulk of liquid extract compared to the smaller ones, due to a greater resistance to the oil transport, linked to longer paths. Other authors (Zhao et al. 2007) found the same behaviour in their system, also suggesting that when the particle is small enough, most of the cell walls are broken by the application of ultrasound, and diffusion would not be a significant step in the extraction of such small particles. A further decrease in the size would not result in a corresponding increase in the extraction rate.

#### **4. Extraction optimization.**

For any extraction process there is an optimum extraction time. Beyond this time, the extractable compounds of interest can be degraded or the achieved yield does not compensate the energy and processing costs. The intention behind the use of ultrasound is that it achieves very high extraction efficiency at the beginning of the process (Orozco-Solano et al. 2010). An example of this approach was described by Wang et al (2008) when studying, in the optimization of the USAE of phenolic compounds from wheat bran (frequency: 40 kHz; extraction temperature: 50 °C; solvent: 70% (v/v) ethanol; raw

1 material/solvent volume ratio: 5 g / 100 mL;  $\Pi$ : 0.025 W / cm<sup>3</sup>), different extraction  
2 experiments with 5.0 g of wheat bran macerated and sonicated for times ranging from 10  
3 to 50 min. A marked increase of the total phenolic content was observed up to 30 min,  
4 then remaining constant until 50 min. Then, the use of ultrasound could be a good way to  
5 increase the extraction yield in the first stage of the process, which is quite appealing for  
6 industrial applications. Despite the great amount of interest in optimizing the operation for  
7 industrial purposes, few USAE kinetics for natural products can be found in the literature,  
8 this being a field of research that, in the future, should be paid attention.  
9  
10  
11  
12  
13  
14

## 15 **5. Industrial approaches.**

16  
17  
18  
19 Soria and Villamiel (2010), considered that, of all the applications of high power-  
20 low frequency ultrasound in the food industry, ultrasound assisted extraction is the most  
21 feasible and economically profitable in the large scale. It is easy to install, has competitive  
22 energy costs and is low maintenance, mainly due to the absence of moving pieces as in  
23 the mechanical stirring methods. However, as was previously shown, it is not a standard  
24 technology and the optimisation of all the considering variables implied in the process is  
25 necessary. For that purpose, it is important to analyse and to optimize the laboratory  
26 scale extraction process as a first step and consider the research and development for  
27 every single application because the processes are case sensitive. For this reason, the  
28 particular scale-up studies could be a profitable source of patents. Scale-up effects were  
29 reported by Boonkird et al. (2008). They found an important decrease in the process yield  
30 for the extraction of capsaicinoids from *Capsicum frutescens* when moving from  
31 laboratory to pilot plant scale. This was a consequence of important changes in the  
32 operational parameters of particle size, power, frequency, geometry and mixing  
33 efficiency, although the kind of solvent, the solvent to material ratio and temperature were  
34 the same.  
35  
36  
37  
38  
39  
40  
41  
42

43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65  
66  
67  
68  
69  
70  
71  
72  
73  
74  
75  
76  
77  
78  
79  
80  
81  
82  
83  
84  
85  
86  
87  
88  
89  
90  
91  
92  
93  
94  
95  
96  
97  
98  
99  
100  
101  
102  
103  
104  
105  
106  
107  
108  
109  
110  
111  
112  
113  
114  
115  
116  
117  
118  
119  
120  
121  
122  
123  
124  
125  
126  
127  
128  
129  
130  
131  
132  
133  
134  
135  
136  
137  
138  
139  
140  
141  
142  
143  
144  
145  
146  
147  
148  
149  
150  
151  
152  
153  
154  
155  
156  
157  
158  
159  
160  
161  
162  
163  
164  
165  
166  
167  
168  
169  
170  
171  
172  
173  
174  
175  
176  
177  
178  
179  
180  
181  
182  
183  
184  
185  
186  
187  
188  
189  
190  
191  
192  
193  
194  
195  
196  
197  
198  
199  
200  
201  
202  
203  
204  
205  
206  
207  
208  
209  
210  
211  
212  
213  
214  
215  
216  
217  
218  
219  
220  
221  
222  
223  
224  
225  
226  
227  
228  
229  
230  
231  
232  
233  
234  
235  
236  
237  
238  
239  
240  
241  
242  
243  
244  
245  
246  
247  
248  
249  
250  
251  
252  
253  
254  
255  
256  
257  
258  
259  
260  
261  
262  
263  
264  
265  
266  
267  
268  
269  
270  
271  
272  
273  
274  
275  
276  
277  
278  
279  
280  
281  
282  
283  
284  
285  
286  
287  
288  
289  
290  
291  
292  
293  
294  
295  
296  
297  
298  
299  
300  
301  
302  
303  
304  
305  
306  
307  
308  
309  
310  
311  
312  
313  
314  
315  
316  
317  
318  
319  
320  
321  
322  
323  
324  
325  
326  
327  
328  
329  
330  
331  
332  
333  
334  
335  
336  
337  
338  
339  
340  
341  
342  
343  
344  
345  
346  
347  
348  
349  
350  
351  
352  
353  
354  
355  
356  
357  
358  
359  
360  
361  
362  
363  
364  
365  
366  
367  
368  
369  
370  
371  
372  
373  
374  
375  
376  
377  
378  
379  
380  
381  
382  
383  
384  
385  
386  
387  
388  
389  
390  
391  
392  
393  
394  
395  
396  
397  
398  
399  
400  
401  
402  
403  
404  
405  
406  
407  
408  
409  
410  
411  
412  
413  
414  
415  
416  
417  
418  
419  
420  
421  
422  
423  
424  
425  
426  
427  
428  
429  
430  
431  
432  
433  
434  
435  
436  
437  
438  
439  
440  
441  
442  
443  
444  
445  
446  
447  
448  
449  
450  
451  
452  
453  
454  
455  
456  
457  
458  
459  
460  
461  
462  
463  
464  
465  
466  
467  
468  
469  
470  
471  
472  
473  
474  
475  
476  
477  
478  
479  
480  
481  
482  
483  
484  
485  
486  
487  
488  
489  
490  
491  
492  
493  
494  
495  
496  
497  
498  
499  
500  
501  
502  
503  
504  
505  
506  
507  
508  
509  
510  
511  
512  
513  
514  
515  
516  
517  
518  
519  
520  
521  
522  
523  
524  
525  
526  
527  
528  
529  
530  
531  
532  
533  
534  
535  
536  
537  
538  
539  
540  
541  
542  
543  
544  
545  
546  
547  
548  
549  
550  
551  
552  
553  
554  
555  
556  
557  
558  
559  
560  
561  
562  
563  
564  
565  
566  
567  
568  
569  
570  
571  
572  
573  
574  
575  
576  
577  
578  
579  
580  
581  
582  
583  
584  
585  
586  
587  
588  
589  
590  
591  
592  
593  
594  
595  
596  
597  
598  
599  
600  
601  
602  
603  
604  
605  
606  
607  
608  
609  
610  
611  
612  
613  
614  
615  
616  
617  
618  
619  
620  
621  
622  
623  
624  
625  
626  
627  
628  
629  
630  
631  
632  
633  
634  
635  
636  
637  
638  
639  
640  
641  
642  
643  
644  
645  
646  
647  
648  
649  
650  
651  
652  
653  
654  
655  
656  
657  
658  
659  
660  
661  
662  
663  
664  
665  
666  
667  
668  
669  
670  
671  
672  
673  
674  
675  
676  
677  
678  
679  
680  
681  
682  
683  
684  
685  
686  
687  
688  
689  
690  
691  
692  
693  
694  
695  
696  
697  
698  
699  
700  
701  
702  
703  
704  
705  
706  
707  
708  
709  
710  
711  
712  
713  
714  
715  
716  
717  
718  
719  
720  
721  
722  
723  
724  
725  
726  
727  
728  
729  
730  
731  
732  
733  
734  
735  
736  
737  
738  
739  
740  
741  
742  
743  
744  
745  
746  
747  
748  
749  
750  
751  
752  
753  
754  
755  
756  
757  
758  
759  
760  
761  
762  
763  
764  
765  
766  
767  
768  
769  
770  
771  
772  
773  
774  
775  
776  
777  
778  
779  
780  
781  
782  
783  
784  
785  
786  
787  
788  
789  
790  
791  
792  
793  
794  
795  
796  
797  
798  
799  
800  
801  
802  
803  
804  
805  
806  
807  
808  
809  
810  
811  
812  
813  
814  
815  
816  
817  
818  
819  
820  
821  
822  
823  
824  
825  
826  
827  
828  
829  
830  
831  
832  
833  
834  
835  
836  
837  
838  
839  
840  
841  
842  
843  
844  
845  
846  
847  
848  
849  
850  
851  
852  
853  
854  
855  
856  
857  
858  
859  
860  
861  
862  
863  
864  
865  
866  
867  
868  
869  
870  
871  
872  
873  
874  
875  
876  
877  
878  
879  
880  
881  
882  
883  
884  
885  
886  
887  
888  
889  
890  
891  
892  
893  
894  
895  
896  
897  
898  
899  
900  
901  
902  
903  
904  
905  
906  
907  
908  
909  
910  
911  
912  
913  
914  
915  
916  
917  
918  
919  
920  
921  
922  
923  
924  
925  
926  
927  
928  
929  
930  
931  
932  
933  
934  
935  
936  
937  
938  
939  
940  
941  
942  
943  
944  
945  
946  
947  
948  
949  
950  
951  
952  
953  
954  
955  
956  
957  
958  
959  
960  
961  
962  
963  
964  
965  
966  
967  
968  
969  
970  
971  
972  
973  
974  
975  
976  
977  
978  
979  
980  
981  
982  
983  
984  
985  
986  
987  
988  
989  
990  
991  
992  
993  
994  
995  
996  
997  
998  
999  
1000

2010). Ultrasound equipment manufacturers offer a wide range of devices for different applications, sometimes on an industrial scale (Hielscher 2011, Meinhardt 2011).

## 6. Future prospects.

As has happened in other fields of ultrasound applications, research is expected into the new possibilities of hybrid ultrasonic technologies in the extraction of natural products. Some successful attempts have been made, such as the combination of ultrasound assisted extraction with supercritical carbon dioxide extraction. Riera et al. (2004) implemented this technology for the extraction of almond oil using an ultrasonic transducer working at 20 kHz located inside a high-pressure extractor operating with CO<sub>2</sub> at 280 bar and 55 °C (Figure 5). The results showed that not only was the kinetics faster, but the extraction yield of the oil was enhanced by 20% to 30%. These improvements may be attributed to an increase in the mass transfer coefficient.

This attempt was later scaled up (Riera et al. 2010) to a pilot-plant scale, tested and validated for cocoa cake oil extraction. The power ultrasonic system operated automatically during the extraction process, meaning that no manual intervention by an external operator is required and the performance, reliability and stability of the process are good. The encouraging results support the need for further experimental and theoretical research into this new, promising field.

Microwave has also been used in combination with ultrasound in extraction processes, either applied simultaneously or in serial disposition. Lianfu and Zelong (2008) optimised an ultrasound-microwave extraction method for the extraction of lycopene from tomatoes. A microwave oven with a maximum power of 800 W at a frequency of 2450 MHz, and an ultrasonic transducer with a fixed power of 50 W at a frequency of 40 kHz were used simultaneously, the solvent being ethyl acetate. Under the optimum conditions for the hybrid method they achieved a recovery of 97.4 % in 6.1 min, improving the best result for USAE, which was 89.4 % in 29.1 min. For the extraction and drying of isoflavonoids from *Pueraria*, a serial disposition was used (USAE followed by microwave drying) (Hu et al. 2008). The time required for the extraction was 20 times shorter than the conventional reflux extraction methods were used. Although it cannot be considered as a hybrid approach for the serial design, it is remarkable that the combination of these emerging technologies is leading to significant advances in the attainment of processes that are safer and less time and energy consuming.

## 7. Final remarks.

It has become more and more commonplace to use USAE in the food industry as the technique for extracting natural valuable compounds thanks to its easy installation and maintenance costs which can be paid back in a short time. It requires less energy and solvent usage and is easier to handle.

1 The variables of the process have a strong influence on the extraction yield and  
2 may be first optimized in a laboratory step for any process. One of the key points for the  
3 scale up and development of the technique is the establishment of a valid method for the  
4 characterization of the power applied and the design of a well-defined reactor. The  
5 research in the scale-up, the design of improved sonoreactors and the combination with  
6 other techniques is expected to significantly enhance the development and  
7 commercialization of this technique.  
8  
9

10 Although this article is mainly focused on food products, they could be easy  
11 extrapolated to the pharmaceutical industry.  
12  
13

#### 14 Acknowledgements.

15  
16  
17 The authors thank the Generalitat Valenciana for their financial support in project  
18 PROMETEO/2010/062 and the Caja de Ahorros del Mediterráneo for M.D. Esclapez's pre-  
19 doctoral grant.  
20  
21  
22  
23

#### 24 References.

- 25  
26  
27  
28 Abad Romero B, Bou-Maroun E, Reparet JM, Blanquet J, Cayot N (2010) Impact of lipid  
29 extraction on the dearomatisation of an *Eisenia foetida* protein powder. *Food Chem* 119: 459–466  
30  
31 Adewuyi YG (2001) Sonochemistry: Environmental Science and Engineering Applications. *Ind*  
32 *Eng Chem Res* 40:4681-4715  
33  
34 Archley AA, Crum LA (1998) In: Suslick KS (ed) *Ultrasound, its chemical physical, and*  
35 *biological effects*, VHS Publishers, Weinheim.  
36  
37 Arnold G, Leiteritz L, Zahn S, Rohm H (2009) Ultrasonic cutting of cheese: Composition affects  
38 cutting work reduction and energy demand. *Int Dairy J* 19:314–320  
39  
40 Barbero GF, Liazid A, Palma M, Barroso CG (2008) Ultrasound-assisted extraction of  
41 capsaicinoids from peppers. *Talanta* 75:1332–1337  
42  
43 Benedito J, Carcel JA, Sanjuan N, Mulet A (2000) Use of ultrasound to assess Cheddar cheese  
44 characteristics. *Ultrasonics* 38:727–730  
45  
46 Benedito J, Carcel JA, Rossello C, Mulet A (2001) Composition assessment of raw meat mixtures  
47 using ultrasonics. *Meat Sci* 57:365-370  
48  
49 Bhaskaracharya RK, Kentish S, Ashokkumar M (2009) Selected Applications of Ultrasonics in  
50 Food Processing. *Food Eng Rev* 1:31–49  
51  
52 Boonkird S, Phisalaphong C, Phisalaphong M (2008) Ultrasound-assisted extraction of  
53 capsaicinoids from *Capsicum frutescens* on a lab- and pilot-plant scale. *Ultrason Sonochem* 15:  
54 1075–1079  
55  
56 Cárcel JA, Benedito J, Bon J, Mulet A (2007a) High intensity ultrasound effects on meat brining.  
57 *Meat Sci* 76:611–619  
58  
59  
60  
61  
62  
63  
64  
65

1 Cárcel JA, Benedito J, Rosselló C, Mulet A (2007b) Influence of ultrasound intensity on mass  
2 transfer in apple immersed in a sucrose solution. *J Food Eng* 78:472–479  
3 Cavitus (2009) Grape colour and flavour extraction (Pat. Pend.) for red must extraction  
4 <http://www.cavitus.com>. Crafers. Accessed: 10 Jan 2011  
5  
6 Chea Chua S, Ping Tan C, Mirhosseini H, Ming Lai O, Long K, Sham Baharin B (2009)  
7 Optimization of ultrasound extraction condition of phospholipids from palm-pressed fiber. *J Food*  
8 *Eng* 92:403–409  
9  
10 Chena R, Menga F, Zhang S, Liu Z (2009) Effects of ultrahigh pressure extraction conditions on  
11 yields and antioxidant activity of ginsenoside from ginseng. *Sep Purif Technol* 66:340–346  
12 Chivate MM, Pandit AB (1995) Quantification of cavitation intensity in fluid bulk, *Ultrason*  
13 *Sonochem* 2:19-25  
14  
15 Da Porto C, Decorti D (2009) Ultrasound-assisted extraction coupled with under vacuum  
16 distillation of flavour compounds from spearmint (carvone-rich) plants: Comparison with  
17 conventional hydrodistillation. *Ultrason Sonochem* 16 :795–799  
18  
19 Da Porto C, Decorti D, Kikic I (2009) Flavour compounds of *Lavandula angustifolia* L. to use in  
20 food manufacturing: Comparison of three different extraction methods *Food Chem.* 112:1072–  
21 1078  
22  
23 Domínguez H, Núñez MJ, Lema JM (1994) Enzymatic pretreatment to enhance oil extraction from  
24 fruits and oilseeds: a review. *Food Chem* 49:271-286  
25  
26 Dong J, Liu Y, Liang Z, Wang W (2010) Investigation on ultrasound-assisted extraction of  
27 salvianolic acid B from *Salvia miltiorrhiza* root. *Ultrason Sonochem* 17: 61–65  
28  
29 Entezari MH, Kruus P (1994) Effect of frequency on sonochemical reactions. I: Oxidation of  
30 iodide. *Ultrason Sonochem* 1:75-79  
31  
32 Esclapez MD, Sáez V, Milán-Yáñez D, Tudela I, Louisnard O, González-García J (2010)  
33 Sonoelectrochemical treatment of water polluted with trichloroacetic acid: From sonovoltammetry  
34 to pre-pilot plant scale. *Ultrason Sonochem* 17:1010–1020  
35  
36 Ferraro V, Cruz IB, Ferreira R, Malcata JFX, Pintado ME, Castro PML (2010) Valorisation of  
37 natural extracts from marine source focused on marine by-products: review. *Food Res Int* 43:2221-  
38 2233  
39  
40 Fischer CH, Hart EJ, Henglein AJ (1986) Hydrogen/deuterium isotope exchange in the hydrogen  
41 deuteride-water system under the influence of ultrasound. *Phys Chem* 90: 3059-3060  
42  
43 García-Noguera J, Weller CL, Oliveira FIP, Rodrigues S, Fernandes FAN (2010) Dual-stage sugar  
44 substitution in strawberries with a Stevia-based sweetener. *Innovative Food Sci Emerg Technol*  
45 11: 225–230  
46  
47 García-Pérez JV, Cárcel JA, de la Fuente-Blanco S, Riera-Franco de Sarabia E (2006) Ultrasonic  
48 drying of foodstuff in a fluidized bed: Parametric study. *Ultrasonics* 44:539–543  
49  
50 García-Pérez JV, García-Alvarado MA, Carcel JA, Mulet A (2010) Extraction kinetics modeling  
51 of antioxidants from grape stalk (*Vitis vinifera* var. Bobal): Influence of drying conditions. *J Food*  
52 *Eng* 101:49–58  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65



1 González-García J, Sáez V, Tudela I, Díez-García MI, Esclapez MD, Louisnard O (2010)  
2 Sonochemical Treatment of Water Polluted by Chlorinated Organocompounds. A Review. *Water*  
3 2:28-74  
4 Handa SS, Preet S, Khanuja S, Longo G, Rakesh DD (2008) Extraction Technologies for  
5 Medicinal and Aromatic Plants. United Nations Industrial Development Organization and the  
6 International Centre for Science and High Technology, Trieste  
7  
8 Hemwimol S, Pavasant P, Shotipruk A (2006) Ultrasound-assisted extraction of anthraquinones  
9 from roots of *Morinda citrifolia*. *Ultrason Sonochem* 13:543–548  
10  
11 Hielscher (2011) Teltow <http://www.hielscher.com>. Accessed 10 Jan 2011  
12  
13 Hu Y, Wang T, Wang M, Han S, Wan P, Fan M (2008) Extraction of isoflavonoids from *Pueraria*  
14 by combining ultrasound with microwave vacuum. *Chem Engin Process* 47:2256–2261  
15  
16 Ince NH, Tezcanli G, Belen RK, Apikyan PG (2001) Ultrasound as a catalyzer of aqueous reaction  
17 systems: the state of the art and environmental applications. *Appl Catal B* 29: 167–176  
18  
19 Jadhav D, Rekha BN, Gogate PR, Rathod VK (2009) Extraction of vanillin from vanilla pods: A  
20 comparison study of conventional soxhlet and ultrasound assisted extraction. *J Food Eng* 93:421–  
21 426  
22  
23 Ji J-b, Lu X-h, Cai M-q, Xu C-c (2006) Improvement of leaching process of Geniposide with  
24 ultrasound. *Ultrason Sonochem* 13:455–462  
25  
26 Kanthale PM, Gogate PR, Pandit AB, Wilhelm AM (2003) Mapping of an ultrasonic horn: link  
27 primary and secondary effects of ultrasound *Ultrason Sonochem* 10:331–335  
28  
29 Karki B, Lamsal BP, Jung S, van Leeuwen JH, Pometto III AL, Grewell D, Khanal SK (2010)  
30 Enhancing protein and sugar release from defatted soy flakes using ultrasound technology. *J Food*  
31 *Eng* 96:270–278  
32  
33 Kardos N, Luche J-L (2001) Sonochemistry of carbohydrate compounds. *Carbohydr Res* 332:115–  
34 131  
35  
36 Kotronarou A, Mills G, Hoffmann MR, (1991) Ultrasonic Irradiation of para-Nitrophenol in  
37 Aqueous Solution. *J Phys Chem* 95:3630-3638  
38  
39 Kuijpers MWA, Kemmere MF, Keurentjes JTF (2002) Calorimetric study of the energy efficiency  
40 for ultrasound-induced radical formation. *Ultrasonics* 40:675–678  
41  
42 Leighton TG (2007) What is ultrasound? *Prog Biophys Mol Biol* 93:3–83  
43  
44 Leonelli C, Mason TJ (2010) Microwave and ultrasonic processing: Now a realistic option for  
45 industry. *Chem Eng Process* 49:885–900  
46  
47 Li H, Pordesimo L, Weiss J (2004) High intensity ultrasound-assisted extraction of oil from  
48 soybeans. *Food Res Int* 37:731–738  
49  
50 Liu J, Li J-W, Tang J (2010) Ultrasonically assisted extraction of total carbohydrates from *Stevia*  
51 *rebaudiana* Bertoni and identification of extracts. *Food Bioprod Process* 88:215–221  
52  
53 Lianfu Z, Zelong L (2008) Optimization and comparison of ultrasound/microwave assisted  
54 extraction (UMAE) and ultrasonic assisted extraction (UAE) of lycopene from tomatoes. *Ultrason*  
55 *Sonochem* 15:731–737  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

1 Liazid A, Schwarz M, Varela RM, Palma M, Guillén DA, Brigui J, Macías FA, Barroso CG  
2 (2010) Evaluation of various extraction techniques for obtaining bioactive extracts from pine  
3 seeds. *Food Bioprod Process* 88:247–252  
4 Londoño-Londoño J, Rodrigues de Lima V, Lara O, Gil A, Crecsynski Pasa TB, Arango GJ,  
5 Ramirez Pineda JR (2010) Clean recovery of antioxidant flavonoids from citrus peel: Optimizing  
6 an aqueous ultrasound-assisted extraction method. *Food Chem* 119:81–87  
7 Lou Z, Wang H, Zhang M, Wang Z (2010) Improved extraction of oil from chickpea under  
8 ultrasound in a dynamic system. *J Food Eng* 98:13–18  
9 Louisnard O, González-García J, Tudela I, Klima J, Sáez V, Vargas-Hernández Y (2009) FEM  
10 simulation of a sono-reactor accounting for vibrations of the boundaries. *Ultrason Sonochem*  
11 16:250-259  
12 Luque de Castro MD, Priego-Capote F (2007) *Analytical Applications of Ultrasound, Vol. 26,*  
13 *Techniques and Instrumentation in Analytical Chemistry, Elsevier Science, Amsterdam*  
14 Ma Y, Ye X, Hao Y, Xu G, Xu G, Liu D (2008) Ultrasound-assisted extraction of hesperidin from  
15 Ma Y, Chen J-C, Dong-Hong Liu, Ye X-Q (2009) Simultaneous extraction of phenolic compounds  
16 of citrus peel extracts: Effect of ultrasound. *Ultrason Sonochem* 16:57–62  
17 Penggan (*Citrus reticulata*) peel. *Ultrason Sonochem* 15:227–232  
18 Makino K, Mossoba MM, Riesz P (1982) Chemical effects of ultrasound on aqueous solutions.  
19 Evidence for hydroxyl and hydrogen free radicals (.cnddot.OH and .cnddot.H) by spin trapping. *J*  
20 *Chem Soc* 104: 3537-3539  
21 Margulis MA, Margulis IM (2003) Calorimetric method for measurement of acoustic power  
22 absorbed in a volume of liquid. *Ultrason Sonochem* 10:343-345  
23 Martin CJ, Law ANR (1983) Design of thermistor probes for measurement of ultrasound intensity  
24 distributions. *Ultrasonics* 21:85-90  
25 Mason TJ, Lorimer JP, Bates DM, Zhao Y (1994) Dosimetry in sonochemistry: the use of aqueous  
26 terephthalate ion as a fluorescence monitor. *Ultrason Sonochem* 1:91-95  
27 Meinhardt (2011) Leipzig. <http://www.meinhardt-ultraschall.de>. Accessed 10 Jan 2011,  
28 Montalbo-Lombay M, Khanal SK, van Leeuwen JH, Raman DR, Dunn Jr L Jr., Grewell D (2010)  
29 Ultrasonic pretreatment of corn slurry for saccharification: A comparison of batch and continuous  
30 Systems. *Ultrason Sonochem* 17: 939–946  
31 Mulet A, Cárcel JA, Sanjuán N, Bon J (2003) New food drying technologies. Use of ultrasound.  
32 *Food Sci Technol Int* 9: 215-221  
33 Naguleswaran S, Vasanthan T (2010) Dry milling of field pea (*Pisum sativum* L.) groats prior to  
34 wet fractionation influences the starch yield and purity *Food Chemistry* 118:627–633  
35 Orozco-Solano M, Ruiz-Jiménez J, Luque de Castro MD (2010) Ultrasound-assisted extraction  
36 and derivatization of sterols and fatty alcohols from olive leaves and drupes prior to determination  
37 by gas chromatography–tandem mass spectrometry. *J Chromatogr A* 1217:1227–1235  
38 Patist A, Bates D (2008). Ultrasonic innovations in the food industry: from the laboratory to  
39 commercial production. *Innovative Food Sci Emerg Technol* 9:147-154  
40 Price GJ (1990) In Mason TJ (ed) *The use of ultrasound for the controlled degradation of polymer*  
41 *solutions. Advances in sonochemistry, Jai Press, Cambridge*

1 Riener J, Noci G, Cronin DA, Morgan DJ, Lyng JG (2010) A comparison of selected quality  
2 characteristics of yoghurts prepared from thermosonicated and conventionally heated milks. *Food*  
3 *Chem* 119:1108–1113

4 Riera E, Golás Y, Blanco A, Gallego JA, Blasco M, Mulet A (2004) Mass transfer enhancement in  
5 supercritical fluids extraction by means of power ultrasound. *Ultrason Sonochem* 11:241–244

6 Riera E, Blanco A, García J, Benedito J, Mulet A, Gallego-Juárez JA, Miguel Blasco (2010) High-  
7 power ultrasonic system for the enhancement of mass transfer in supercritical CO<sub>2</sub> extraction  
8 processes. *Physics Procedia* 3:141-146

9 Roldán-Gutiérrez JM, Ruiz-Jiménez J, Luque de Castro MD (2008) Ultrasound-assisted dynamic  
10 extraction of valuable compounds from aromatic plants and flowers as compared with steam  
11 distillation and superheated liquid extraction. *Talanta* 75:1369–1375

12 Romdhane M, Gourdon C (2002) Investigation in solid–liquid extraction: influence of ultrasound.  
13 *Chem Eng J* 87:11–19

14 Rong L, Kojima Y, Koda S, Nomura H (2008) Simple quantification of ultrasonic intensity using  
15 aqueous solution of phenolphthalein. *Ultrason Sonochem* 8:11-15

16 Sáez V, Frias-Ferrer A, Iniesta J, Gonzalez-Garcia J, Aldaz A, Riera E (2005) Characterization of a  
17 20 kHz sonoreactor. Part I: analysis of mechanical effects by classical and numerical methods.  
18 *Ultrason Sonochem* 12:59–65

19 Sáez V, Frias-Ferrer A, Iniesta J, Gonzalez-Garcia J, Aldaz A, Riera E (2005) Characterization of  
20 a 20 kHz sonoreactor. Part II: analysis of chemical effects by classical and electrochemical  
21 methods. *Ultrason Sonochem* 12:67-72

22 Sahena F, Zaidul ISM, Jinap S, Karim AA, Abbas KA, Norulaini NAN, Omar AKM (2009)  
23 Application of supercritical CO<sub>2</sub> in lipid extraction – A review. *J Food Eng* 95:240–253  
24 Science Direct Database. [www.sciencedirect.com](http://www.sciencedirect.com) (Data of consulting: February 2011)

25 Soria AC, Villamiel M (2010) Effect of ultrasound on the technological properties and bioactivity  
26 of food: a review. *Trends Food Sci Technol* 21:323-331

27 Starmans DAJ, Nijhuis HH (1996) Extraction of secondary metabolites from plant material: A  
28 review. *Trends Food Sci Technol* 7:191-197

29 Sivakumar V, Lakshmi Anna J, Vijayeeswarri J, Swaminathan G (2009) Ultrasound assisted  
30 enhancement in natural dye extraction from beetroot for industrial applications and natural dyeing  
31 of leather. *Ultrason Sonochem* 16:782–789

32 Stanisavljevic IT, Lazic ML, Veljkovic VB (2007) Ultrasonic extraction of oil from tobacco  
33 (*Nicotiana tabacum* L.) seeds. *Ultrason Sonochem* 14:646–652

34 Sun Y, Liu D, Chen J, Ye X, Yu D (2011) Effects of different factors of ultrasound treatment on  
35 the extraction yield of the all-trans-β-carotene from citrus peels. *Ultrason Sonochem* 18: 243–249

36 Suslick K S (2001) In: Meyers RA (ed) *Sonoluminescence and Sonochemistry Encyclopedia of*  
37 *Physical Science and Technology*, 3rd edn. Academic Press, San Diego

38 Trabelsi F, Ait-Iyazidi H, Berlan J, Fabre PL, Delmas H, Wilhelm AM (1996) Electrochemical  
39 determination of the active zones in a high-frequency ultrasonic reactor. *Ultrason Sonochem*  
40 3:125-130

1 Veillet S, Tomao V, Chemat F (2010) Ultrasound assisted maceration: An original procedure for  
2 direct aromatisation of olive oil with basil. *Food Chem* 123:905–911

3 Velickovic DT, Milenovic DM, Ristic MS, Veljkovic VB (2008) Ultrasonic extraction of waste  
4 solid residues from the *Salvia* sp. Essential oil hydrodistillation. *Biochem Eng J* 42:97–104

5 Vercet A, Burgos J, Crelier S, Lopez-Buesa P (2001) Inactivation of proteases and lipases by  
6 ultrasound. *Innovative Food Sci Emerg Technol* 2:139-150

7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

Vilkhu K, Mawson R, Simons L, Bates D (2008) Applications and opportunities for ultrasound  
assisted extraction in the food industry — A review. *Innovative Food Sci Emerg Technol* 9:161–  
169

Vinatoru M (2001) An overview of the ultrasonically assisted extraction of bioactive principles  
from herbs. *Ultrason Sonochem* 8:303–313

Virost M, Tomao V, Le Bourvellec C, Renard CMCG, Chemat F (2010) Towards the industrial  
production of antioxidants from food processing by-products with ultrasound-assisted extraction.  
*Ultrason Sonochem* 17:1066–1074

Wang J, Sun B, Cao Y, Tian Y, Li X (2008) Optimisation of ultrasound-assisted extraction of  
phenolic compounds from wheat bran. *Food Chem* 106:804–810

Wang L, Weller CL (2006) Recent advances in extraction of nutraceuticals from plants. *Trends*  
*Food Sci Technol* 17:300–312

Wei X, Chen M, Xiao Ja, Liu Y, Yu L, Zhang H, Wang Y (2010) Composition and bioactivity of  
tea flower polysaccharides obtained by different methods. *Carbohydr Polym* 79:418–422

Weissler A, Cooper HW, Snyder S (1950) Chemical effects of ultrasonic waves: oxidation of  
potassium iodide solution by carbon tetrachloride. *J Am Chem Soc* 72: 1769-1775.

Wulff-Pérez M, Torcello-Gómez A, Gálvez-Ruiz MJ, Martín-Rodríguez A (2009) Stability of  
emulsions for parenteral feeding: Preparation and characterization of o/w nanoemulsions with  
natural oils and Pluronic f68 as surfactant. *Food Hydrocolloids* 23:1096–1102

Yang B, Yang H, Li J, Li Z, Jiang Y (2011) Amino acid composition, molecular weight  
distribution and antioxidant activity of protein hydrolysates of soy sauce lees. *Food Chem*  
124:551–555

Yang Y, Zhang F (2008) Ultrasound-assisted extraction of rutin and quercetin from *Euonymus*  
*alatus* (Thunb.) Sieb. *Ultrason Sonochem* 15:308–313

Zhang Z-S, Wang L-J, Li D, Jiao S-S, Chena XD, Maoa Z-H (2008) Ultrasound-assisted  
extraction of oil from flaxseed. *Sep Purif Technol* 62:192–198

Zhang H-F, Yang X-H, Zhao L-D, Wang Y (2009) Ultrasonic-assisted extraction of epimedin C  
from fresh leaves of *Epimedium* and extraction mechanism. *Innovative Food Sci Emerg Technol*  
10:54–60

Zhang Q-A, Zhang Z-Q, Yue X-F, Fan X-H, Li T, Chen S-F (2009) Response surface optimization  
of ultrasound-assisted oil extraction from autoclaved almond powder. *Food Chem* 116:513–518

Zhao S, Kwok K-C, Liang H (2007) Investigation on ultrasound assisted extraction of  
saikosaponins from *Radix Bupleuri*. *Sep Purif Technol* 55:307–312

Zhu KX, Sun X-H, Zhou H-M (2009) Optimization of ultrasound-assisted extraction of defatted  
wheat germ proteins by reverse micelles. *J Cereal Sci* 50:266–271

Zheng L, Sun D-W (2006) Innovative applications of power ultrasound during food freezing processes—a review. *Trends Food Sci Technol* 17:16–23

Zou Y, Xie C, Fan G, Gu Z, Han Y (2010) Optimization of ultrasound-assisted extraction of melanin from *Auricularia auricula* fruit bodies. *Innovative Food Sci Emerg Technol* 11:611–615

1  
2  
3  
4  
5  
6  
7  
8  
9  
10  
11  
12  
13  
14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29  
30  
31  
32  
33  
34  
35  
36  
37  
38  
39  
40  
41  
42  
43  
44  
45  
46  
47  
48  
49  
50  
51  
52  
53  
54  
55  
56  
57  
58  
59  
60  
61  
62  
63  
64  
65

## Figure Captions

**Fig. 1** Number of peer reviewed articles published in the last 30 years dealing about ultrasound assisted extraction in food science ([www.sciencedirect.com](http://www.sciencedirect.com) 2011).

**Fig. 2** Schematic representation of the rarefaction (A) and compression cycle (B) of a cavitating bubble that eventually collapses near the solution/biological tissue interface generating shock waves and micro-jets (C) that can disrupt the cell wall releasing the intra cellular material into the solution (D).

**Fig. 3.** Relationship between the percentage of electrical input to the ultrasonic generator and ultrasonic intensity measured using a calorimetric method in a sucrose solution of 30 Brix at 30 °C. From (Cárcel et al. 2007b)

**Fig. 4** Most typical reactor configurations: ultrasonic bath (a), transducer dipped in batch configuration (b), transducer immersed in a flow reactor (c). From González-García et al. (2010).

**Fig. 5** Scheme of the supercritical fluid extractor coupled with an ultrasonic transducer. Adapted from Riera et al. (2004).

Table

Table 1. Operating conditions for the extraction of some natural compounds.

Extract	Raw material	Freq. (kHz)	$\Pi$ (W/cm <sup>3</sup> ) <sup>b</sup>	t (min)	T (°C)	Raw material/total solvent ratio	Solvent	Equipment	Extraction yield	Reference
Total carbohydrates	Stevia	20 <sup>a</sup>	0.06	32	68	100g dry/1 L <sup>a</sup>	Water, pH 7 <sup>a</sup>	2	17 g/100g	Liu et al. 2010
Flavonoids	Citrus peel	60	**	30	40	1 g dry/10 mL	basified water	1	40 mg/g	Londoño-Londoño et al. 2010
Oil	Chickpea	40 <sup>a</sup>	250 W	20	50	1 g/8.5 mL	3:1 hexane-isopropanol	3 (5mL/min flow)	81% of soxhlet leaching	Lou et al. 2010
Salvianolic acid B	Salvia miltiorrhiza root	45 <sup>a</sup>	100 W <sup>a</sup>	28	30	1 g/20 mL	60:40 EtOH-H <sub>2</sub> O	1	33.9 mg/g <sup>1</sup>	Dong et al. 2010
Equiv. catequin	Apple pomace	24 <sup>a</sup>	0.142 W/g	45	40.1	15 g/100 mL	1:1 EtOH-H <sub>2</sub> O	1	964 mg/100 g	Virost et al. 2010
Dye	Beetroot	20 <sup>a</sup>	1.6	180	45 <sup>a</sup>	1 g/50 mL <sup>a</sup>	1:1 EtOH-H <sub>2</sub> O	1	0.19 g/g	Sivakumar et al. 2009
Cinnamic acids	Citrus peel	60 <sup>a</sup>	0.02	40	30	2 g/40 mL <sup>a</sup>	4:1 EtOH-H <sub>2</sub> O	1	Depending on compound	Ma et al. 2009
Benzoic acids	Citrus peel	60 <sup>a</sup>	0.02	10	40	2 g/40 mL <sup>a</sup>	4:1 EtOH-H <sub>2</sub> O	1	Depending on compound	Ma et al. 2009
Proteins	Defatted wheat germ flour	20 <sup>a</sup>	7.26	24	Ice bath <sup>a</sup>	0.5 g/50 mL <sup>a</sup>	Reverse micellar solution	2 (pulsed 2.4s:2s)	57 %	Zhu et al. 2009 <sup>1</sup>
Phosphatidylethanolamine	Palm-pressed fiber	24 <sup>a</sup>	1.53	30	25 <sup>a</sup>	15 g/60 mL	1:3 HCCl <sub>3</sub> -MeOH <sup>a</sup>	2 (pulsed 0.2 W / s)	12570 mg/kg	Chea Chua et al. 2009
Phosphatidylcholine	Palm-pressed fiber	24 <sup>a</sup>	1.53	30	25 <sup>a</sup>	15 g/60 mL	1:3 HCCl <sub>3</sub> -MeOH <sup>a</sup>	2 (pulsed 0.2 W s <sup>-1</sup> )	5426 mg/kg	Chea Chua et al, 2009
Vainillin	Vanilla pods	22.4 <sup>a</sup>	2.40	60	Water bath <sup>a</sup>	0.5 g/100 mL	1:1 EtOH-H <sub>2</sub> O	2 (pulsed 5 s:5 s <sup>a</sup> )	140 ppm in leachant	Jadhav et al. 2009
Phenolic compounds	Wheat bran	40 <sup>a</sup>	0.025	25	60	5 g/100 mL	64:36 EtOH-H <sub>2</sub> O	1	3.12 mg gallic ac. equiv/g	Wang et al. 2008
Oil	Flaxseed	20 <sup>a</sup>	50 W	30	30	16.7 g/100 mL	n-hexane	2	85 % of soxhlet leaching	Zhang et al. 2008
Nordihydrocapsaicin	Cayenne	** <sup>a</sup>	360 W <sup>a</sup>	10	50	1 g/25 mL	MeOH	1	94 μmol/kg	Barbero et al. 2008

Extract	Raw material	Freq. (kHz)	$\Pi$ (W/cm <sup>3</sup> ) <sup>b</sup>	t (min)	T (°C)	Raw material/total solvent ratio	Solvent	Equipment	Extraction yield	Reference
Nordihydrocapsaicin	Bolilla Redondo pepper	**a	360W <sup>a</sup>	10	50	1 g/25 mL	MeOH	1	40 $\mu$ mol/kg	Barbero et al. 2008
Nordihydrocapsaicin	Bolilla Largo Pepper	**a	360 W <sup>a</sup>	10	50	1 g/25 mL	MeOH	1	25 $\mu$ mol/kg	Barbero et al. 2008
Capsaicin	Cayenne	**a	360 W <sup>a</sup>	10	50	1 g/25 mL	MeOH	1	448 $\mu$ mol/kg	Barbero et al. 2008
Capsaicin	Bolilla Redondo pepper	** a	360 W <sup>a</sup>	10	50	1 g/25 mL	MeOH	1	370 $\mu$ mol/kg	Barbero et al. 2008
Capsaicin	Bolilla Largo pepper	** a	360 W <sup>a</sup>	10	50	1 g/25 mL	MeOH	1	275 $\mu$ mol/kg	Barbero et al. 2008
Dihydrocapsaicin	Cayenne	** a	360 W <sup>a</sup>	10	50	1 g/25 mL	MeOH	1	265 $\mu$ mol/kg	Barbero et al. 2008
Dihydrocapsaicin	Bolilla Redondo pepper	** a	360 W <sup>a</sup>	10	50	1 g/25 mL	MeOH	1	190 $\mu$ mol/kg	Barbero et al. 2008
Dihydrocapsaicin	Bolilla Largo pepper	** a	360 W <sup>a</sup>	10	50	1 g/25 mL	MeOH	1	122 $\mu$ mol/kg	Barbero et al. 2008
Essential oils	Laurel	20 <sup>a</sup>	45 W <sup>a</sup>	10	25	1 g/8.5 mL	EtOH	3 (pulsed 30% <sup>a</sup> , 4 mL/min),	Depending on compound	Roldan-Gutierrez et al. 2008
Hesperidin	Penggan peel	60	3.2	60	40	1 g/40 mL	MeOH	1	57 mg/g	Ma et al. 2008
Rutin	Euonymus alatus	50 <sup>a</sup>	0.05 <sup>a</sup>	30(x3)	Amb. <sup>a</sup>	0.5 g/20 mL	70:30 EtOH-H <sub>2</sub> O	1	0.299 mg/g	Yang et al. 2008
Quercetin	Euonymus alatus	50 <sup>a</sup>	0.05 <sup>a</sup>	30(x3)	Amb. <sup>a</sup>	0.5 g/20 mL	70:30 EtOH-H <sub>2</sub> O	1	0.013 mg/g	Yang et al, 2008

<sup>a</sup> Non-optimized value.

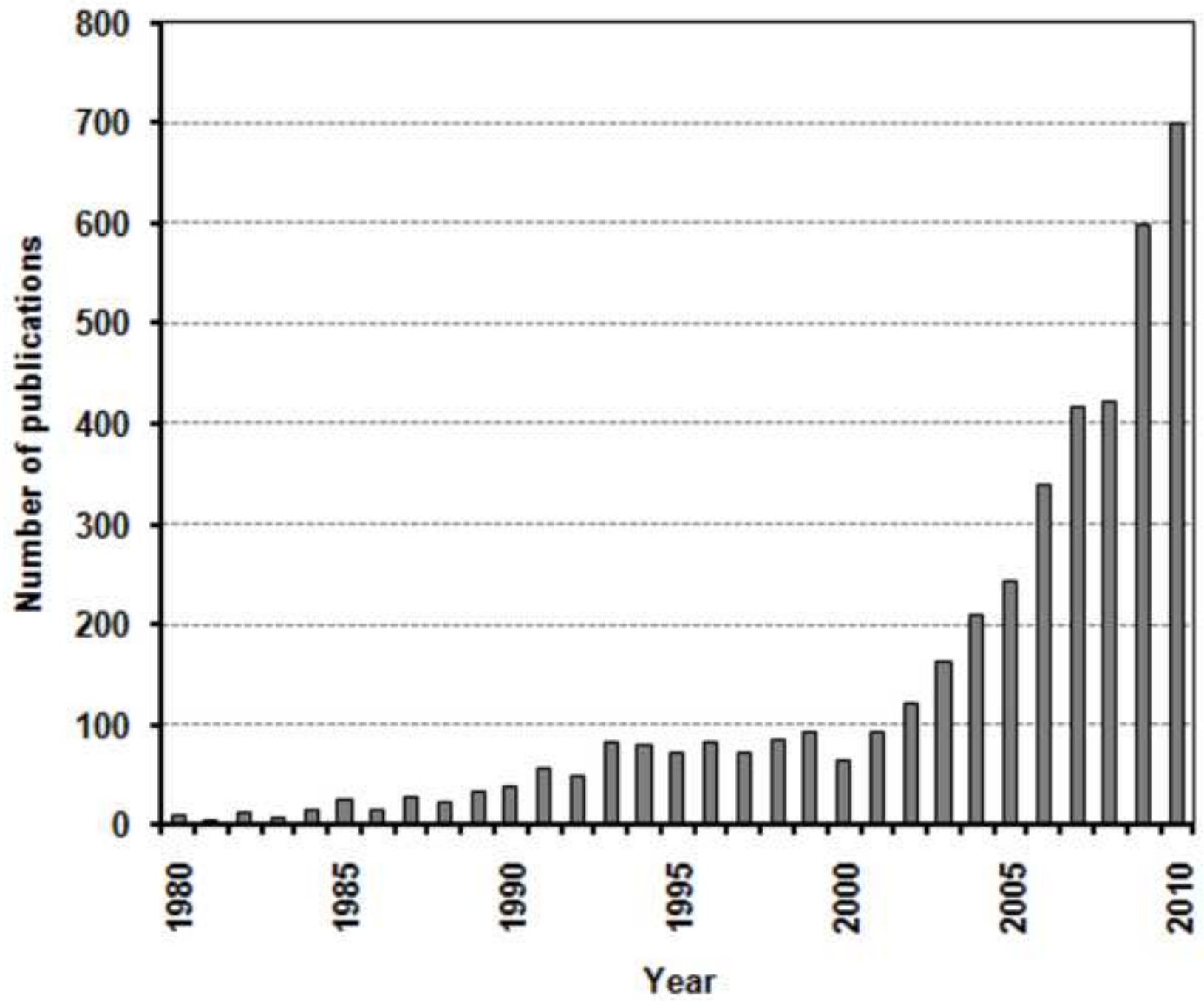
<sup>b</sup>  $\Pi$  was calculated when possible. In other cases, total power applied is shown, (W).

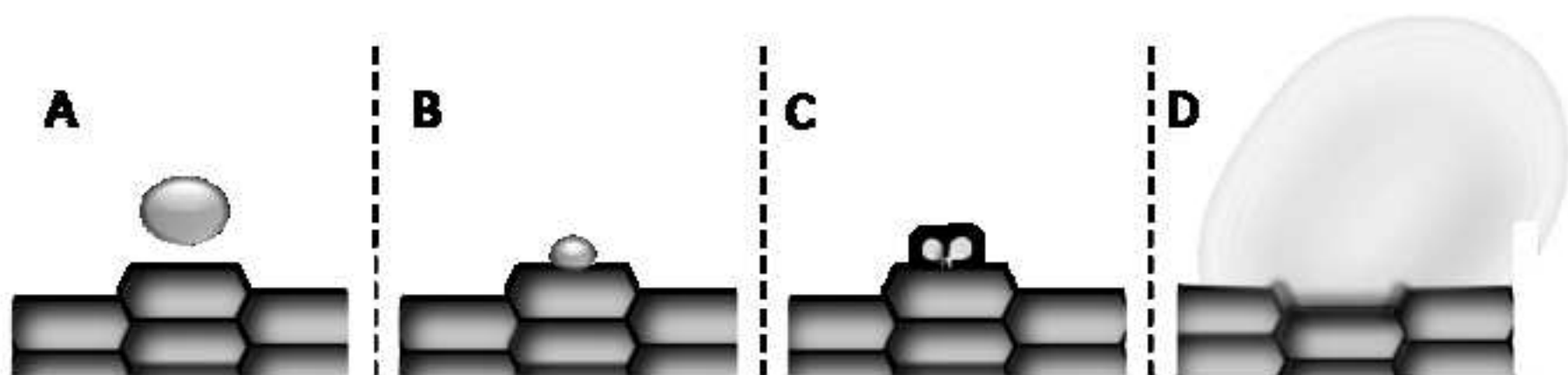
1 Ultrasonic cleaning bath.

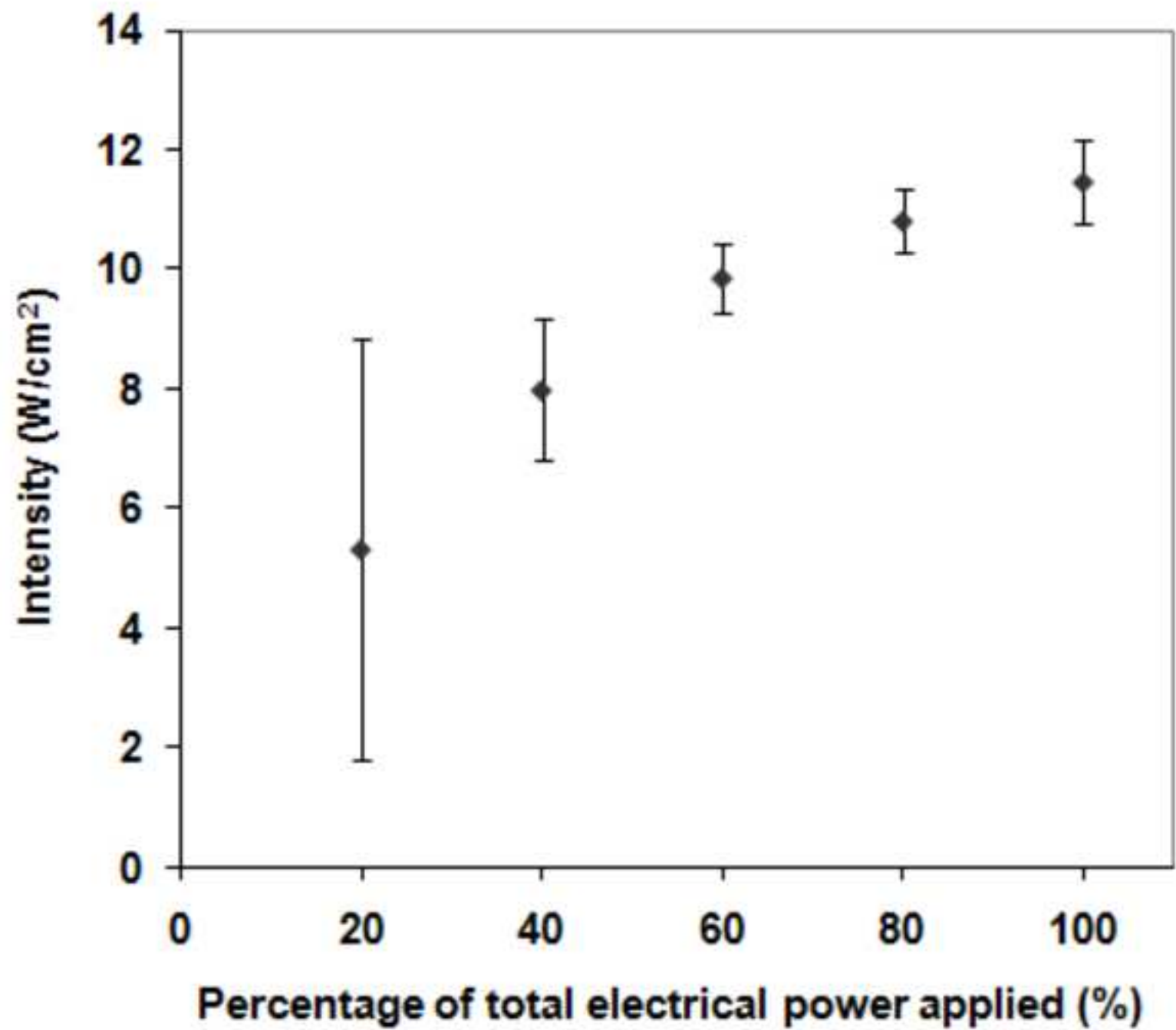
2 Ultrasonic probe.

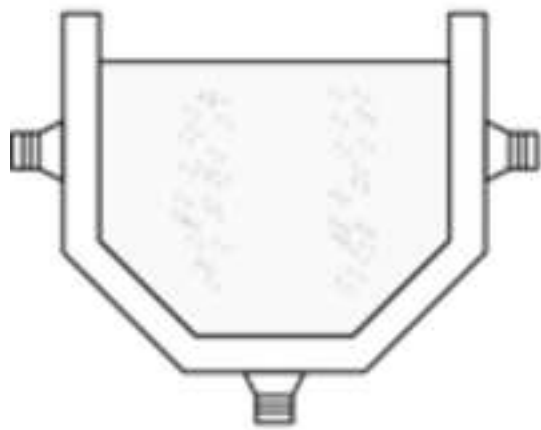
3 Specifically designed reactor.



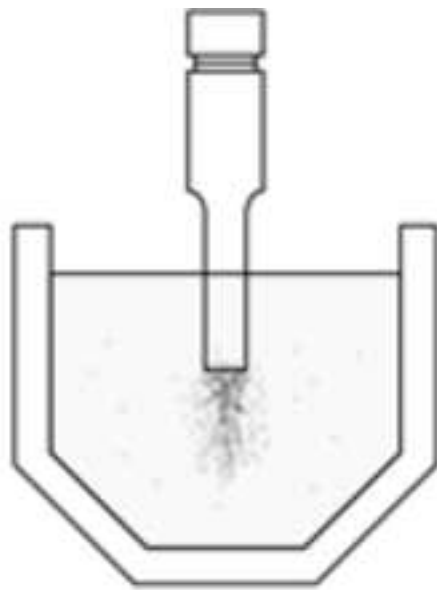




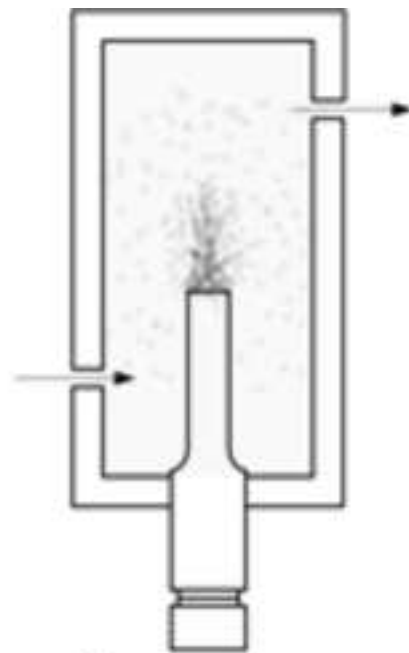




a



b



c

